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EXPERT REVIEW

Current and Emerging Therapies in T-cell Acute Lymphoblastic Leukemia

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Abstract: Current studies of T-cell acute lymphoblastic leukemia (T-ALL) show promise for improving the treatment for this previously difficult to cure neoplasm. This has been facilitated by the increased understanding of the molecular basis of leukemogenesis in T-cells as well as advanced analytical technologies in modern molecular biology. Combined with intensive conventional therapies, novel drugs as well as improved diagnostic protocols have been developed for the rational therapy of T-ALL, resulting in a cure rate up to 80%. An understanding of the detailed molecular mechanism of action for T-ALL specific alterations in gene expression has led to the design and development of targeted molecular therapies of this aggressive cancer. Several novel, target-specific drugs are now available, including mitotic inhibitors with more specificity and potency for growth inhibition, γ -secretase inhibitors (GSIs) for leukemic NOTCHI signaling blockage in T-ALL as well as growth inhibition, tyrosine kinase inhibitors for correction of uncontrolled proliferation caused by certain genetic lesions, and inhibitors of BCL2 proteins leading to sensitization of leukemic cells to apoptosis induction. Some of these drugs are under development in the different phase of clinic trials, but others are currently included in successful therapeutic protocols, and, as hoped, they cause the remission of patients that suffered from certain types of T-ALLs, and who previously were poor responders using conventional treatment protocols. However, these new approaches still often need to be combined with highly toxic conventional treatment to obtain satisfactory overall outcome. The encouraging achievements in targeted molecular therapy challenge researchers to achieve no-event complete remission in T-ALL patient by developing novel therapies with even more specific targeting and efficacy to minimize complications. Finally, with the development of targeted molecular therapy, rational approaches for individually tailored treatment of T-ALL patients in the near future now may be an achievable goal.

Keywords: T-acute lymphoblastic leukemia, chemotherapy, apoptosis

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Introduction

Leukemia is a fairly common cancer, with an incidence of 30-40 per million. It is estimated that 43,050 new cases of leukemia were diagnosed in the United States in 2010. The major types of leukemia include acute lymphoblastic (or lymphoid) leukemia, chronic lymphocytic leukemia, acute myeloid leukemia. chronic myelogenous (or myeloid) leukemia, and other, rarer forms of leukemia. Acute lymphoblastic leukemia (ALL) is the most common pediatric malignancy, with over 70% of diagnosed cases occurring in children and adolescents less than 20 years old.¹⁻⁴ ALL can progress quickly if untreated, and it is responsible for about 1,400 deaths a year in the U.S. Approximately, 2,900 new ALLs are diagnosed each year in the United States, and in the past several years, there has been a gradual increase in the incidence of ALL.5-7 Acute lymphoblastic leukemia is a heterogeneous disease. It originates as a single B- or T-lymphocyte progenitor, and it is further characterized as pre-B, B, or T-cell ALL. T cell ALL (T-ALL) is diagnosed in 15%-20% of children and adults with ALL, while about 80%-85% of ALL cases are of the B-cell lineage. Current treatment for T- and B-ALL mainly depends on multi-agent combination chemotherapy, and over the past six decades, overall cure rates for pediatric ALL have significantly improved to long term (5 years event-free) survival rates of nearly 80%-90%.^{2,8-12}

T-ALL is a malignancy of thymocytes (T-cell progenitor) that develops mainly in children and adolescents, which is much less common or rare in adults. Upon spreading throughout the body, rapid progression of this aggressive tumour leads to high peripheral blood cell counts, increased numbers of blast cells, CNS dissemination and, frequently, to large mediastinal masses that cause tracheal compression and respiratory distress at diagnosis, and which is fatal without treatment. Comparing to B-cell ALL, T-cell precursor acute lymphoblastic leukemia is clinically regarded as a high risk disease with a high frequency of treatment failure and a relapse rate of about 30%-40%.^{4,13–16} T-ALL is now treated the same way as highrisk B-progenitor ALL. With appropriately intensive therapy, children with T-ALL have an estimated 5-year event-free survival rate at 75%-80%.^{1,14,17,18} Nevertheless, T-ALL patients have increased risk for remission induction failure, early relapse (2 years,



>50%), and, especially, isolated CNS relapse.¹⁹⁻²¹ Treatment outcome is poor among those patients who relapse on current frontline chemo-regimens and over the past two decades, survival rates after relapse have been consistently low. Furthermore, much less success has been achieved in adult T-ALL patients, with a long term survival rate at only about 30% to 40% (<60 years of age) or 10% (>60 years of age), at most.^{16,20-28} Moreover, during or after intensive treatment, many T-ALL patients develop serious acute and late complications because of the side effects of the highly toxic chemotherapeutic reagents. These realities have challenged investigators to develop novel approaches that incorporate salvage regimens and frontline therapeutic strategies for these high-risk patients.^{12,21,25,28-30} This is accomplished by treatment protocols that more efficiently and specifically target the leukemic cells to improve the rate of complete remission (relapse free) as well as the quality of life of patients.

Commonly Used Chemotherapies for T-ALL

Using standard intensive chemotherapy, the current cure rate of T-ALL has risen to about 70%–80% in children and 40%–50% in adults, with the expectation of better overall outcome in the near future. This significant progress in outcome largely depends on the selective use and combination of the anti-leukemia regimens together with stringent applications of risk-directed therapy (by prognostic factors) in clinical trials. And in general, a combined modality (directed by different phases of treatment), intensive chemotherapy is currently used to treat T-ALL in a manner that is essentially the same as high risk ALLs.^{4,16,18,21,22,26–28,30–35} Various protocols based on different combinations of drugs have been developed for T-ALL treatment at various phases.

1. Induction therapy: Initially, different combinations of various cytoctoxic drugs are used to elimanate the leukemic cells by inducing cell lysis and growth inhibition. There are over 10 different drugs currently available and generally used in combinations for intensive treatment. The cytoxicity of these drugs can be grouped according to the manner in which they interfere with the biological functions of leukemic cells, for example, as a



metabolic inhibitor for an essential metabolistic pathway or as signaling molecules that promote apoptosis or lysis of the cancer cells. Table 1 lists the commonly used chemotherapeutic drugs for the current treatment of ALL and their biological effects on leukemic cells. These drugs include DNA damaging agents (anthracyclines,³⁶ alkylating agents,³⁷ and epipodophyllotoxins^{38,39}), anti-metabolites (nucleotides analogues such as 6-mercaptopurine⁴⁰ and methotrexate^{41,42}), mitotic inhibitors (vincristine^{43,44}), protein synthesis inhibitors (specifically, asparaginase and its more potent derivatives, PEG-asparaginase or Oncaspar[®],^{45–49} etc.) and corticosteroids (glucocorticoid analogues, prednisone and dexamethasone).^{47,50–55}

- 2. Consolidation: Treatment is directed at leukemic cells in the central nervous system (CNS). High dose systemic methotrexate or cranial irradiation with continued systemic chemotherapy (mercaptopurine and vincristine) is adopted to prevent CNS relapses. The treatment may last for 4 weeks.
- 3. Interim maintenance and delayed intensification: At this stage, the phase of interim maintenance

followed by delayed intensification is repeated. Interim maintenance is designed to adopt less intense therapies after the induction therapy and between the courses of delayed intensification, which allow for normal cell recovery. Delayed intensification is a repeat of more intense induction therapy, in attempts to destroy any trace of leukemic cells surviving the induction therapy. Multiple drugs are administered in an intensive schedule to intensify the remission. This phase is usually two months long and consists of primarily oral medications given at home.

- 4. Maintenance: A 1–2 year maintenance phase of treatment is required. The treatment in this phase is important because it aims at eliminating any remaining, slow dividing leukemia initiating cells, thus prevent relapse. Maintenance treatment consists of oral administration of 6-mercaptopurine and methotrexate, monthly visits for pulses of vincristine and corticosteroids (dexamethasone) and intrathecal methotrexate every 12 weeks.
- 5. Prognosis and risk-directed therapy: Early treatment response is the most important prognostic factor to stratify risk-directed therapy. This strategy

Agent	Mechanism of action		
Prednisone	Small signaling molecular to activate glucoccorticoid signaling pathway; Binds to		
Dexamethasone	and activates glucocorticoid receptor, a member of nuclear steroid receptor and transcription factor, to regulate gene expression; Induces sensitive apoptosis of ALL leukemic cells in vitro and in vivo.		
Vincristine	Binds to tubulin protein and inhibits formation of mitotic spindle, leading to growth an and cell death.		
Daunorubicin	Binds to DNA and inhibits the progression of the enzyme topoisomerase II by stabilizin		
Doxorubicin	the topoisomerase II complex after it has broken the DNA chain for replication, leadin		
Idarubicin	to double-stranded DNA breaks and stop of DNA replication, which causes DNA damage and apoptosis.		
Etoposide Teniposide	Forms a ternary complex with DNA and the topoisomerase II enzyme and prevents re-ligation of the DNA strands, leading to DNA damage and apoptosis.		
Asparaginase	catalyzes the hydrolysis of asparagine to aspartic acid, inhibits protein synthesis in ALL leukemic cells by depriving the circulating asparagine since ALL leukemic cells ar unable to synthesize the non-essential amino acid asparagine.		
Methotrexate	Inhibits the synthesis of DNA, RNA, thymidylates, and proteins by inhibiting dihydrofolate reductase (DHFR) involved in the de novo synthesis of the nucleoside thymidine and purine; leading to growth inhibition and cell death.		
6-Thioguanine	Interferes with purine nucleotides synthesis; incorporates into RNA and DNA, leading		
6-Mercaptopurine Azathioprine	to genotoxic stress (growth inhibition and cell death).		
Cytarabine	Inhibits the synthesis of DNA by incorporating into DNA, leading to DNA damage, growth inhibition (cell cycle arrest) and cell death.		
Cyclophosphamide	Forms DNA crosslinks (irreversible) between and within DNA strands at guanine N-7 positions, leading to cell death.		

 Table 1. Mechanism of commonly used chemotherapy for ALLs.

introduces a stringent assessment of the risk of relapse for subgroups of patients, which is critical in selecting therapy that will avoid excessive toxicity but maintain a high cure rate, as well as efficiently designing experimental therapy for very high risk patients. Currently, risk classification is based on apparent clinical features of patients and characteristics of leukemia subtypes; interestingly, the response to a single week of mono-treatment with steroids strongly predicts outcome of treatment. In fact, steroid analogues have been a mainstay in all protocols developed to treat ALL including T-ALL.^{53–55}

Nevertheless, for rare cases of adult T-ALL, even with the most intensified protocol, the outcome is much less favourable,⁵⁶ and in childhood T-ALL, about 20% of patients are still uncured.^{2,4,21,35} Currently there are no effective ways to treat these patients; very high doses of cytoxic drug combinations must be used for treatment, which exerts severe side-effects on patients during or after the treatment. These serious short and long-term complications that result from the toxicity of these therapies must be considered. Thus, there remains a need for more effective agents to be incorporated into T-ALL treatment regimens, and novel reagents/strategies are needed to obtain cures for patients when commonly used therapies fail.

An important trend in anti-cancer drug development is to understand the molecular basis of leukemogenesis (e.g., genetic and epigenetic changes, alterations to the proteome) and therapeutic resistance in leukemia. These studies have led to the development of new agents against relevant molecular targets, and fostered the concept of "molecular targeted therapy" for the highly specific treatment of T-ALL.^{56–59} An excellent example of this is the development of imatinib for chemotherapeutic regimens now widely used to treat Philadelphia chromosome-positive (Ph+) ALL (B-ALLs and a rare portion of T-ALLs) and chronic myelogenous leukemia (CML).59,60 Based on the presentation of molecular abnormalities of a patient's leukemic subtype, more effective, and potentially less toxic, individually tailored treatment protocols for T-ALL are under development.

The Biology of T-ALL

The biological knowledge of T-ALL has been greatly expanded in the past several years. T-ALL originates



from immature, stem-cell-like thymocytes. In normal T-cell development, lymphoid progenitor cells migrate from the bone marrow towards the thymus, where CD4 CD8 double-negative progenitor cells developed into mature T-cell receptor (TCR) β or δ positive T-cells. Neoplastic transformation of the lymphoblast during this process leads to leukemogenesis of these progenitor cells as T-ALL.61-63 Recent studies have revealed that this requires multi-step mutagenesis and genetic/genomic alterations. These changes cause the deregulation of certain genes involved in the critical events of normal T-cell development, e.g., cell-cycle control, differentiation, cell survival and proliferation, which ultimately result in developmental arrest at nearly all stages of T-cell maturation.63,64 Clarification of the molecular bases and cytogenetic changes specifically associated with T-ALL has provided novel opportunities for developing highly efficient, targeted therapy of this cancer.

T-ALL is characterized by various chromosomal translocations and aneuploidy.⁶¹ Using animal models together with advanced detection techniques in molecular biology, the molecular-genetic abnormalities in T-ALL, including chromosomal translocations, deletions, amplifications, duplications and point mutations, have been intensively investigated. T-ALLs may be grouped into distinct subgroups, TAL/LOM, LYL1, TLX1 (HOX11), TLX3 (HOX11L2), MYB, HOXA, or other defects (e.g., PICALM-MLLT10 and SET-NUP214), which are characterized by ectopic expression of certain genes. This may represent an alternative to conventional karyotyping for diagnostic classification and therapy selection. Gene expression analysis in specific T-cell ALLs has revealed that 70% of these genetic alterations involve altered expression of transcription factors. In childhood T-ALL, deregulation of MYB, TAL/SCL or the homebox gene TLX3 (HOX11L2) is observed in over 50% of patients, while in adults the expression of TLX1 (HOX11) or SCL is found in two major groups.^{65–74} It is notable that these transcription factors are "master" transcription factors that regulate the development, differentiation, and proliferation of T-lymphoblast cells, which determines the gene signature and the characteristic T-ALL subsets.75 In addition, homozygous deletions of tumour suppressor genes, e.g., genes coded by the CDKN2A/2B loci (p16INK4A, p15INK4B and p14ARF), occur in 90%



of the cases of leukemic T-lineage lymphoblasts.⁶¹ These subsets of different genetic alterations possibly are of clinical relevance. For example, expression of TLX1 (HOX11) in T-ALLs is associated with favourable treatment outcomes, while unfavourable results have been observed with TAL1 and LYL1 expression.^{69,72} Potentially, based on the genetic markers associated with the clinical presentations, these studies may provide a means of T-ALL classification for rational treatment.

The molecular basis or mechanisms of genetic alterations in T-ALL have been intensively explored. Gene expression profiling, as well as several genomewide approaches, have shown that tyrosine kinases (e.g., BCR-ABL1, EML1-ABL1, ETV6-ABL1 and ETV6-JAK2), and certain important signaling pathways, including NOTCH1, PTEN, and (pre)TCR signaling, are specifically associated with development of thymocytes and are deregulated in T-ALL. Activating NOTCH1 mutations have now been identified in more than 50% of T-ALL, resulting in constitutive NOTCH signaling in these cells.^{76–82} In one study, the presence of activating NOTCH1 mutations was associated with a favourable early treatment response. Targeting the signaling pathway of NOTCH1 has been suggested to be a potential therapeutic strategy in T-ALL.⁸³⁻⁸⁶ Furthermore, multiple components of the (pre)TCR signaling pathway are targeted by either mutations or chromosomal rearrangements in T-ALL.⁶¹ The RAS protein is involved in the transmission of TCR signaling from membrane receptors to the ERK protein, which is commonly mutated in a wide variety of malignancies. In T-ALL, activating RAS mutations have been identified in 4%-10% of cases.⁶⁴ These patients would potentially benefit from additional treatment with RAS inhibitors.

Recent studies in T-ALL also have focused on understanding apoptosis in lymphoblast cells. Apoptosis plays important roles in the normal development and homeostasis of lymphoid lineages.^{87,88} The cytotoxicity of the most chemotherapeutic agents currently in use largely depends on their ability to induce a killing response in leukemic cells at different phases of the cell cycle.^{89,90} Chemotherapeutic drugs stimulate an apoptosis response in leukemic cells using different upstream mechanisms. Methotrexate, cytosine arabinoside, camptothecin, etoposide and adriamycin preferentially induce apoptosis in S phase. Prednisone (or dexamethasone) and 6-mercaptopurine effectively induce apoptosis in the G1 phase and G1 + S phases, respectively. Another cytotoxic agent used to treat leukemia, cyclophosphamide, induces apoptosis with no cell cycle phase specificity.91 However, these agents eventually induce apoptosis through a common, mitochondrial dependent (intrinsic pathway) downstream death machinery mechanism, involving a cascade of caspase activation that is conserved in eukaryotic cells across species. This downstream process is fundamentally regulated by BCL-2 family members.⁹²⁻⁹⁴ The BCL-2 family of proteins consists of three subgroups, anti-apoptotic proteins (e.g., Bcl-2, Bcl-xL, Mcl-1) that promote cell survival, pro-apoptotic proteins (BH3 only proteins, e.g., Bid, Bim, PUMA) that trigger cell death, and proapoptotic effector proteins (Bax, Bak and Bok, having intrinsic pore-forming activity) that act downstream of BH3-only and pro-survival Bcl-2 members to induce mitochondrial outer membrane permeabilization (MOMP). This ultimately leads to the activation of caspase effectors. The balance of anti-apoptotic and pro-apoptotic BCL-2 family members determines the survival of lymphoblast cells during development of the normal lymphoid lineage and, in T-ALL, the ratio of BCL-2/BIM affects apoptosis in response to multiple chemotherapeutic reagents. In fact, possibly due to downregulation of antiapoptotic genes, TLX1 (HOX11)+ in adult T-ALL is associated with a favourable outcome of treatment. Conversely, the poor prognosis found in T-ALL subtypes expressing TAL1 or LYL1 is thought to be caused by the concomitant upregulation of antiapoptotic genes that confer resistance to chemotherapy.^{65,72} It has been shown that, frequently, overexpression of pro-survival BCL-2 proteins may contribute to the profound resistance to chemotherapy in T-ALL poor responders.^{95,96} These studies have highlighted the clinical relevance of the BCL-2 family members as potential targets for novel molecular targeting therapy in T-ALL.

Moleular Targets in T-ALL and Emerging New Therapeutic Compounds

Gowth arrest and inhibition of cell cycle progression Nelarabine: Nelarabine is an ana

Nelarabine: Nelarabine is an analog of arabinosylguanine and acts as a prodrug of ara-G.⁹⁷ By becoming demethylated and coverted into ara-GTP intracellularly, nelarabine induces rapid cell cycle arrest (G1-S or S phase), elevated expression of FasL and apoptosis of leukemic cells via both the extrinsic and intrinsic pathways. Nelarabine is much more effective in T-cell ALL than B-ALL, probably because of a genetic deficiency of purine nucleoside phosphorylase in T-cells leading to a selective sensitivity of T-ALL to deoxyguanosine.98 Phase II trials on refractory or relapsed T-cell malignancies were conducted.^{99,100} In the pediatric study, an initial nelarabine dose of 1,200 mg/m²/d was administered for 5 consecutive days was de-escalated due to neurotoxicity, resulting in the final dose of 650 mg/m²/d, with further de-escalation to $400 \text{ mg/m}^2/\text{d}$ for patients with CNS complications. A complete remission was achieved in 46% of 39 patients in first relapse, 25% of 40 patients in second relapse, and 21% of 28 with CNS disease.¹⁰¹ In the adult trial, 26 patients were given nelarabine on an alternate day schedule (days 1, 3, and 5) at 1,500 mg/m² per day. 31% of them achieved complete remission.¹⁰²⁻¹⁰⁴ Nelarabine had been approved by the US Food and Drug Administration (FDA) for third-line treatment of patients with T-cell leukemia or lymphoma.

Flavopiridol: Flavopiridol (FP) is a flavonoid derived from an indigenous plant from India. FP is a potent inhibitor of cyclin dependent kinases (cdks) 1, 2, 4 and 7 in vitro, which induces transient cell cycle arrest at G1/S and/or G2/M and apoptosis.105 In ALL cells, FP treatment suppresses the activity of cdks. This decreases the expression of phosphorylated retinoblastoma protein (ser-795/807/811) and effectively leads to cell cycle arrest. Furthermore, FP also reduces the expression of Mcl-1 and phosphorylated forms of the C-terminal domain of RNA polymerase II.¹⁰⁶⁻¹⁰⁸ In primary ALL blasts, FP treatment increases cell death by approximately two-fold over baseline.¹⁰⁶ FP was more potent in vitro than glucocorticoids and thiopurines and at doses that recent phase I experience predicts will translate into clinical efficacy.¹⁰⁹ FP should be widely effective in ALL if sufficient plasma levels can be achieved clinically.

Inhibition of NOTCH1 signaling

NOTCH signaling is deregulated in the majority of T-cell acute lymphoblastic leukemias as a result



of activating mutations in NOTCH1.76-79 NOTCH signalling is activated by a series of proteolytic cleavage events, leading to the release of the intracellular domain (NICD) via ligand-dependent cleavage mediated by the gamma secretase (GS) complex. Upon GS cleavage, NICD translocates to the nucleus, binds to the transcription factor CSL and other protein cofactors, and activates the transcription of NOTCH target genes including HES1, HEY1, MYC, PTCRA, DTX1 and certain protein components in NFkB signaling pathway. Also, activated NOTCH1 can regulate the activity of the mTOR signaling pathway by phosphorylation.^{78,83,86} The high frequency of NOTCH1 mutations found in associating to human T-ALLs makes the NOTCH signaling pathway the best available rational target in treatment of this disorder.

Gamma-secretase inhibitors (GSIs): GSIs comprise a class of small molecules that interfere with the proteolytic cleavage of the receptor and inhibit the release of NICD.⁸⁶ Potentially, by suppressing the NOTCH signaling pathways, GSIs may induce growth arrest in T-ALL cells and cause prolonged cell cycle arrest and apoptosis.84,110 The potency of GSIs in T-ALL treatment has been shown by multiple pre-clinical studies. A reversible, noncompetitive, gamma-secretase inhibitor, PF-03084014, has been investigated for the treatment of T-ALL and advanced solid tumours in phase I clinical trials. PF-03084014 selectively inhibits gamma-secretase and reduces endogenous NICD levels, which results in the downregulation of Notch target genes Hes-1 and c-Myc in T-ALL cell lines in vitro and in vivo. In T-ALL, PF-03084014 treatment also causes cell growth inhibition through cell cycle arrest and induction of apoptosis. Animal models have demonstrated the broad antitumor efficacy of PF-03084014 at well-tolerated dose levels. Treatment of PF-03084014 induces gastrointestinal toxicity, while glucocorticoids abrogate PF-03084014-induced gastrointestinal toxicity.85,111 These studies show promise for the development of an approach using PF-03084014 and steroid therapy for Notch receptor-dependent T-ALL and other cancers. In another study, GSI developed by Merck (MRK-003) effectively suppresses Notch1 target gene expression and causes apoptosis of T-ALL leukemic cells in vivo, and significantly extended the survival of leukemic cell grafted mice in animal study.¹¹²



However, during phase I clinical trial, MRK-003 failed to induce favourable responses in relapsed/ refractory T-ALL patients. One possible explaination might be attributed to the finding that the dysfunction of Notch1 signalling is more likely a secondary (acquired) leukemogenic event in T-ALLs.¹¹³ Several other pre-clinical studies and clinical trials are testing the anti-tumour use of GSIs. These attempts should verify the therapeutic promise of GSIs in treatment of T-ALL and certain other cancers.^{114–116}

Resveratrol: Resveratrol (RES), a potential chemopreventive agent is a natural phytoalexin.¹¹⁷ The cytotoxicity of resveratrol is shown by its ability to inhibit cell proliferation and induce apoptosis in a variety of cancer cell lines. In MOLT-4 acute lymphoblastic leukemia cells, RES inhibits the survival and induces apoptosis by (1) inhibiting Notch signaling pathways and their down stream effectors; (2) increasing the pro-apoptotic protein p53 and its effectors p21^{waf} and Bax; and, (3) inhibiting the PI3K/Akt pathway and activating Gsk-3beta.¹¹⁸ Thus, future studies may further indicate a role for RES as an inhibitor of the NOTCH signaling pathway and as an apoptosis inducer in the treatment of T-ALL.

Tyrosine kinase inhibitors

The c-ABL proto-oncogene product, ABL1, is a tyosine kinase that regulates key processes including cell growth and survival, differentiation, stress and cell migration. The c-ABL gene is frequently one of the targets of mutations and genetic lesions in various cancers. Chromosome translocation t(9;22)(q34;q11)results in the formation of a Philadelphia chromosome (Ph) and generates an active chimeric BCR-ABL tyrosine kinase.¹¹⁹ Typically, BCR-ABL is found in chronic myelogenous leukemia (CML) and precursor B-acute lymphoblastic leukemia (pre B-ALL), but it is exceptionally rare in patients with T-ALL (less than 1%). Another active fusion of ABL1 has been identified, ETV6-ABL1. It is caused by an acquired del(6)(q15q23), which is also preferentially associated with CML and B-ALL, and rarely in T-ALL (<1%). Furthermore, a unique T-ALL case had identified an EML1-ABL1 fusion due to a cryptic t(9;14) (q34;q32). However, in T-ALLs, another type of genetic lesion, episomal fusion of NUP214 to ABL1 (NUP214-ABL), resulted by episomal ABL1 gene amplication, and this has been identified in 6% of T-ALL patients.^{73,120} Overexpression of the ABL fusion gene activates a number of downstream signaling cascades including the mitogen activated protein kinase (MAPK) and Janus kinase signal transducer and activator of transcription (JAK-STAT) signaling pathways, leading to growth-factor independent proliferation.¹²¹ It also effectively promotes the survival of leukemic cells in the face of apoptosis induction. Until the recent introduction of tyrosine kinase inhibitors (imatinib and its more potent analogues), these genetic lesions had been one of the most unfavourable prognosis subsets among all types of acute lymphoblastic leukemia, with very poor outcome and high rate of refractoriness to treatment.^{122,123}

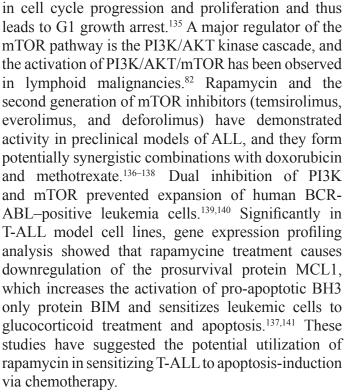
Imatinib mesylate (Imatinib, STI-571): Imatinib is a tyrosine kinase inhibitor that inhibits the ABL, KIT and PDGFR kinases, and which has considerable activity against active BCR-ABL1 fusions in CML and ALL. Imatinib is used in the treatment of Ph+ associated leukemia, and it is a dramatic example of targeted molecular therapy against a specific genetic/ molecular lesion.^{60,124,125} Several studies have shown that imatinib combined with chemotherapy induces complete remission rates at levels of 90% to 100% and dramatically improves disease-free survival duration in patients with Ph+ ALL. In another study, combined with stem cell transplantation, administration of imatinib improved the treatment outcome of Ph+ALL patients with a 3 year event-free remission of over 78%.126 NUP214-ABL1 T-ALL is sensitive to the tyrosine kinase inhibitor imatinib in vitro, which indicates a potential new therapeutic approach to these cases. However, in a clinical study of one NUP214-ABL1 T-ALL case, imatinib failed to induce remission of patient.^{127,128} This might suggest a resistance mechanism of NUP214-ABL1 T-ALL to imatinib treatment in vivo.

Nilotinib and dasatinib: Nilotinib and dasatinib were developed as second-generation tyrosine kinase inhibitors, which are significantly more potent than imatinib. While nilotinib is the aminopyrimidine modified analogue of imatinib, dasatinib has no structural similarity to imatinib. Dasatinib is a dual Src/Abl kinase inhibitor that is 300-fold more potent against active ABL1 kinase than imatinib, both in vitro and in vivo. Both nilotinib and dasatinib seem to be well tolerated and show activity against imatinib-resistant Ph+ leukemias, except in resistant leukemias with a T315I mutation in the ABL1 gene.^{129,130} Subsequent clinical studies have shown that dasatinib induces a complete hematological response in 43% of patients and a cytogenetic response in 32% of 23 patients with CML.¹³¹ Patients with initial imatinib intolerance had more favourable responses to dasatinib than did patients with initial imatinib resistance. A combination of imatinib with dasatinib is expected to overcome drug resistance.^{132,133} However, the optimal dose schedule of dasatinib in combination with Ph+ ALL regimens remains to be determined.^{25,134} Remarkably, in a clinic study, complete hematologic and cytogenetic remission was achieved on a NUP214-ABL1 positive T-ALL patient after 3 weeks of dasatinib monotherapy treatment (70 mg of dasatinib two times daily).¹²⁸ These studies have raised the hope of treating these rare deadly cases of T-ALLs with this enhanced tyrosine kinase inhibitor.

Sensitizing apoptosis by targeted activation of pro-apoptotic BCL-2 family proteins

Activation of pro-apoptotic members of BCL-2 family proteins (BH3-only proteins) is a key event that commits cancer cells to apoptosis in response to many apoptosis-inducing chemotherapeutic agents. This is also strongly regulated by pro-survival BCL-2 proteins (e.g., BCL-2, MCL1) that bind to pro-apoptotic proteins and prevent apoptisis in normal conditions. The BH3-only protein, BIM, plays a pivotal role in regulating apoptosis of lymphoblast cells. Overexpression of prosurvival protein BCL-2 or downregulation of BIM effectively results in resistance to apoptosis-inducing agents, including radiation, corticosteroids, and chemotherapy. Therefore, several drugs targeting these proteins may potentially benefit the treatment of T-ALL by sensitizing the leukemic cells to apoptosis.

Rapamycin and mTOR kinase inhibitors: The mammalian target of rapamycin (mTOR) is a serine-threonine protein kinase that belongs to the phosphoinositide3-kinase(PI3K)-relatedkinasefamily. Inhibition of mTOR kinase results in dephosphorylation of its two major downstream signaling components p70 S6 kinase (p70S6K) and eukaryotic initiation factor 4E binding protein 1 (4E-BP1), which in turn inhibits the translation of specific mRNAs involved



Obatoclax (GX15-070): The small molecule obatoclax is an antagonist of BCL-2 like proteins. It is an indole bipyrrole compound that is designed to target all BCL-2 family proteins, and it is especially potent against MCL1. Although obatoclax is currently used in developing single-agent therapy or in combination in phase I/II clinical trials directed at leukemia, the molecular mechanisms of cell death induced by obatoclax are not entirely clear. Studies in ALL model cell lines have revealed that obatoclax induces cell death through BAK-dependent apoptosis (by disrupting the MCL1/BAK complex) and via ATG5-dependent (becline-1 independent) autophagy.142-146 Unlike the other small-molecule BCL-2 inhibitors, e.g., ABT-737 and ABT-263, obatoclax can overcome the resistance conferred by high levels of MCL1.^{142,146} Potentially, obatoclax can be developed as a monotherapeutic agent against T-ALL.

ABT-737: ABT-737 (an orally available derivate ABT-263) ws developed to selectively target BCL-2-like proteins. ABT-737 has effective cytotoxicity against many hematological malignancies including ALL, acute myeloid leukemia (AML), multiple myeloma, lymphoma and chronic lymphocytic leukemia (CLL). It is able to bind BCL-2 family proteins including BCL-2, BCL-XL and BCL-w, but not for





MCL1.^{147,148} Potently, its binding activity disrupts the interactions of these pro-survival proteins with their pro-apoptotic counterparts, leading to activation of BH3-only proteins including BIM and BID, which in turn sensitize cells to or trigger apoptosis. This mechanism involves an increase in cleaved fragments of PARP, and caspase-8 and cytochrome C levels shortly after treatment with ABT-737 at low micromolar concentrations.¹⁴⁹ BH3 profiling analysis of ALL cell lines as well as primary samples from ALL patients reveals the BCL-2 dependence of ALL, supporting the potential usefulness of ABT-737, as antagonist of BCL-2, in sensitizing leukemic cells for apoptosis induction.^{148,150,151}

Conclusions

Current progress in studies on T-ALL using the technology of modern molecular biology has greatly improved cure rates for this type of cancer. Fundamentally, the goal of modern cancer research is to develop more effective therapies that specifically target the cancer cell with minimal effects on normal cells and tissues. Conventional therapies for T-ALL currently used clinically depend on cytotoxic anticancer drugs that induce apoptosis of malignant cells, but which inevitably cause considerable short or long term side effects and damage to the health of the patient. Furthermore, the non-selective cytotoxicity of anticancer drugs adversely affects the goals of treatment. After intensive chemotherapy, stem cell transplantation has to be performed for some patients, and, in many cases, secondary malignancies occur as the result of the treatment. These situations have prompted the development of novel therapeutic approaches with more specific targeting and efficiency. Based on the elucidation of the molecular basis of T-ALL, a few targets have been identified and drugs and protocols that target these molecules have been developed (summarized in Table 2). However, the specificity and efficiency of these treatments are uncertain, due to the relatively short period that they have been available for clinic applications. Also, it is notable that, in the most cases, these new approaches still have to be combined with conventional treatment to obtain satisfactory outcome. Other novel therapeutic approaches under development include RNA interference to silence leukemic oncogene expression of the T-ALL specific mRNA and immunotherapy that exploits the antitumor activity of cytotoxic T lymphocytes or natural killer cells.^{2,152,153} Moreover, protein therapy appears to be another interesting approach. Leukemogenesis is associated with the deregulated expression transcription factors. Specific targeting of transcription factors that are associated with the transformation of certain T-ALLs could potentially lead to strategies for rational antileukemia therapy.

Table 2. Selected new therapies for T-acute lymphoblastic leukemia.

Agent	Mechanism of action	Targeting subtype of leukaemia
Clofarabine	Inhibits DNA polymerase and ribonucleotide reductase; disrupts mitochrondria membrane	All types
Nelarabine	Inhibits ribonucleotide reductase and DNA synthesis	T-cell
Flavopiridol	Serine-threonine cyclin-dependent kinase inhibition	All types
Forodesine	Inhibits purine nucleoside phosphorylase	T-cell
γ-secretase inhibitor (PF-03084014)	Inhibit γ -secretase, an enzyme required for NOTCH1 signaling	T-cell
Resveratrol	Inhibits NOTCH1 signaling; Inhibit PIK3/AKT signaling pathway	T-cell
Imatinib mesilate; Nilotinib	ABL kinase inhibition	BCR-ABL-positive
Dasatinib	BCR-ABL kinase inhibition	BCR-ABL-positive
Rapmycine; Sirolimus; temsirolimus; everolimus; AP-23573	Mammalian target-of-rapamycin (mTOR) inhibition	All types
Obatoclax	Inhibits BCL-2 anti-apoptosis proteins; Disrupt MCL-1/BAK complex	All types
ABT-737 (ABT-263)	Inhibits binding of BCL-2 anti-apoptosis proteins to pro-apoptotic BH3 proteins BIM and BID	All types

Successful techniques of protein transduction mediated by a protein transduction domain (PTD) have made this possible by rendering the cell permeable to any protein molecule. Because of the transient activity of endogenous proteins used for treatment (due to eventual cellular degradation of proteins), very low side effects are expected for this strategy. Using model T-ALL cell lines, a few related studies have been successfully conducted in vitro.¹⁵⁴ More work clearly needs to be done to better exploit this technology. In summary, in the past several years, targeted molecular therapy has emerged as a major focus of interest in T-ALL treatment. Based on an improved future understanding of the molecular mechanism of the leukogenosis, additional treatment targets that are T-ALL specific are expected to be identified. It is hoped that an optimal combination of these target specific drugs will lead to rational treatment of T-ALL patients to obtain complete remission with the lowest possible instances of non-specific side effects.

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References

- 1. Tucci F, Arico M. Treatment of pediatric acute lymphoblastic leukemia. *Haematologica*. 2008;93(8):1124–8.
- Pui CH, Evans WE. Treatment of acute lymphoblastic leukemia. N Engl J Med. 2006;354:166–78.



- 3. Advani AS, Hunger SP, Burnett AK. Acute Leukemia in Adolescents and Young Adults. *Seminars in Oncology*. 2009;36(3):213–26.
- Pui C-H. Acute Lymphoblastic Leukemia: Introduction. Seminars in Hematology. 2009;46(1):1–2.
- Ries LAG, Kosary CL, Hankey BF, Miller BA, Clegg L, Edwards BK. SEER Cancer Statistics Review, 1973–1996: National Cancer Institute, Bethesda, MD; 1999.
- Shah A, Coleman MP. Increasing incidence of childhood leukaemia: a controversy re-examined. *Br J Cancer*. 2007;97(7):1009–12.
- Smith MA, Ries LAG, Gurney JG, Ross JA, editors. *Leukemia*.: National Cancer Institute, Bethesda, MD; 1999. Smith MA, Ries LAG, Gurney JG, Ross JA, editors. Cancer incidence and survival among children and adolescents: United States SEER Program 1975–1995.
- 8. Armstrong GT, Liu Q, Yasui Y, et al. Late mortality among 5-year survivors of childhood cancer: a summary from the Childhood Cancer Survivor Study. *J Clin Oncol.* 2009;27(14):2328–38.
- 9. Margolin JF, Steuber CP, Poplack DG. *Acute lymphoblastic leukemia*. 15th. ed; 2006.
- Pui CH, Campana D, Pei D, et al. Treating childhood acute lymphoblastic leukemia without cranial irradiation. *New England Journal of Medicine*. 2009;360(26):2730–41.
- Gaynon PS, Angiolillo AL, Carroll WL, et al. Long-term results of the children's cancer group studies for childhood acute lymphoblastic leukemia 1983–2002: A children's oncology group report. *Leukemia*. 2009;24(2): 285–97.
- Pui C-H, Cheng C, Leung W, et al. Extended follow-up of long-term survivors of childhood acute lymphoblastic leukemia. N Engl J Med. 2003;349(7):640–9.
- Möricke A, Reiter A, Zimmermann M, et al. Risk-adjusted therapy of acute lymphoblastic leukemia can decrease treatment burden and improve survival: treatment results of 2169 unselected pediatric and adolescent patients enrolled in the trial ALL-BFM 95. *Blood.* 2008;111(9):4477–89.
- Conter V, Arico M, Valsecchi MG, et al. Intensive BFM chemotherapy for childhood ALL: interim analysis of the AIEOP-ALL 91study. Associazione Italiana Ematologia Oncologia Pediatrica. *Haematologica*. 1998;83(9): 791–9.
- Pui C-H, Boyett JM, Rivera GK, et al. Long-term results of Total Therapy studies 11, 12 and 13A for childhood acute lymphoblastic leukemia at St Jude Children's Research Hospital. *Leukemia*. 2000;14(12):2286–94.
- Hoelzer D, Thiel E, Loffler H, et al. Prognostic factors in a multicenter study for treatment of acute lymphoblastic leukemia in adults. *Blood*. 1988;71(1): 123–31.
- Aricò M, Valsecchi MG, Rizzari C, et al. Long-term results of the AIEOP-ALL-95 Trial for Childhood Acute Lymphoblastic Leukemia: insight on the prognostic value of DNA index in the framework of Berlin-Frankfurt-Muenster based chemotherapy. *J Clin Oncol.* 2008;26(2):283–9.
- Hunger SP, Devidas M, Camitta B. Improved survival for children with acute lymphoblastic leukemia (ALL) from 1990–2005: A report from the Children's Oncology Group. 40th Congress of the International Society of Pediatric Oncology, Berlin, Germany [abstract]. *Pediatr Blood Cancer*. 2008;52.
- Coustan-Smith E, Mullighan CG, Onciu M, et al. Early T-cell precursor leukaemia: a subtype of very high-risk acute lymphoblastic leukaemia. *Lancet Oncol.* 2009;10(2):147–56.
- Ko RH, Ji L, Barnette P, et al. Outcome of patients treated for relapsed or refractory acute lymphoblastic leukemia: A therapeutic advances in childhood leukemia consortium study. *J Clin Oncol.* 2010;28(4):648–54.
- Bassan R, Hoelzer D. Modern therapy of acute lymphoblastic leukemia. *J Clin Oncol.* 2011;29(5):532–43.
- 22. Hunault M, Harousseau J-L, Delain M, et al. Better outcome of adult acute lymphoblastic leukemia after early genoidentical allogeneic bone marrow transplantation (BMT) than after late high-dose therapy and autologous BMT: a GOELAMS trial. *Blood*. 2004;104(10):3028–37.
- 23. Freyer DR, Devidas M, La M, et al. Postrelapse survival in childhood acute lymphoblastic leukemia is independent of initial treatment intensity: A report from the Children's Oncology Group. *Blood*. 2010;117(11):3010–5.



- Wofford MM, Smith SD, Shuster JJ, Johnson W, Buchanan GR, Wharam MD. Treatment of occult or late overt testicular relapse in children with acute lymphoblastic leukemia: a Pediatric Oncology Group study. *J Clin Oncol.* 1992;10:624–30.
- Hunger SP, Raetz EA, Loh ML, Mullighan CG. Improving outcomes for high-risk ALL: Translating new discoveries into clinical care. *Pediatr Blood Cancer*. 2011;56(6):984–93.
- Hermine O, Wattel E, Gessain A, Bazarbachi A. Adult T Cell Leukaemia: A Review of Established and New Treatments. *Bio Drugs*. 1998;10(6): 447–62.
- 27. Goldstone AH, Richards SM, Lazarus HM, et al. In adults with standardrisk acute lymphoblastic leukemia, the greatest benefit is achieved from a matched sibling allogeneic transplantation in first complete remission, and an autologous transplantation is less effective than conventional consolidation/maintenance chemotherapy in all patients: final results of the International ALL Trial (MRC UKALL XII/ECOG E2993). *Blood.* 2008; 111(4):1827–33.
- Takeuchi J, Kyo T, Naito K, et al. Induction therapy by frequent administration of doxorubicin with four other drugs, followed by intensive consolidation and maintenance therapy for adult acute lymphoblastic leukemia: The JALSG-ALL93 Study. *Leukemia*. 2002;16(7):1259–66.
- Oeffinger KC, Hudson MM. Long-term complications following childhood and adolescent cancer: foundations for providing risk-based health care for survivors. *CA Cancer J Clin.* July 1, 2004;54(4):208–36.
- Tanimoto M, Miyawaki S, Ino T, et al. Response-oriented individualized induction therapy followed by intensive consolidation and maintenance for adult patients with acute lymphoblastic leukemia: the ALL-87 study of the Japan Adult Leukemia Study Group (JALSG). *Int J Hematol.* 1998;68(4): 421–9.
- Pui CH, Ochs J, Kalwinsky DK, Costlow ME. Impact of treatment efficacy on the prognostic value of glucocorticoid receptor levels in childhood acute lymphoblastic leukemia. *Leukemia Res.* 1984;8:345–50.
- Vrooman LM, Silverman LB. Childhood acute lymphoblastic leukemia: update on prognostic factors. *Current Opinion in Pediatrics*. 2009;21(1):1–8 10.1097/MOP.1090b1013e32831f32831f32824.
- Vecchi V, Pession A, Paolucci G, et al. Risk-directed therapy for childhood acute lymphoblastic leukemia. Results of the associazione italiana ematologia oncologia pediatrica '82 studies. *Cancer*. 1993;72(8): 2517–24.
- Tzortzatou-Stathopoulou F, Papadopoulou AL, Moschovi M, Botsonis A, Tsangaris GT. Low relapse rate in children with acute lymphoblastic leukemia after risk-directed therapy. *J Pediatr Hematol Oncol.* 2001;23(9): 591–7.
- 35. Pui CH. Risk assessment in acute lymphoblastic leukemia: beyond leukemia cell characteristics. *J Pediatr Hematol Oncol*. 2001;23(7):405–8.
- Klumper E, Pieters R, den Boer ML, Huismans DR, Loonen AH, Veerman AJ. In vitro anthracycline cross-resistance pattern in childhood acute lymphoblastic leukaemia. *Br J Cancer*. 1995;71(6):1188–93.
- Gerson SL, Willson JK. O6-alkylguanine-DNA alkyltransferase. A target for the modulation of drug resistance. *Hematol Oncol Clin North Am.* 1995; 9(2):431–50.
- Brenner DE. Antitumor antibiotics, epipodophyllotoxins, and vinca alkaloids: clinical developments. *Current Opinion in Oncology*. 1990;2(6): 1115–8.
- Chow KC, Macdonald TL, Ross WE. DNA binding by epipodophyllotoxins and N-acyl anthracyclines: implications for mechanism of topoisomerase II inhibition. *Mol Pharmacol.* 1988;34(4):467–73.
- Bökkerink JPM, Stet EH, De Abreu RA, et al. 6-Mercaptopurine: Cytotoxicity and biochemical pharmacology in human malignant T-lymphoblasts. *Biochemical Pharmacology*. 1993;45(7):1455–63.
- McGuire JJ. Anticancer antifolates: current status and future directions. Curr Pharm Des. 2003;9(13):2593–613.
- 42. McGuire JJ, Russell CA, Bolanowska WE, Freitag CM, Jones CS, Kalman TI. Biochemical and Growth Inhibition Studies of Methotrexate and Aminopterin Analogues Containing a Tetrazole Ring in Place of the gamma-Carboxyl Group. *Cancer Research*. Mar 15, 1990;50(6):1726–31.

- Perez EA. Microtubule inhibitors: Differentiating tubulin-inhibiting agents based on mechanisms of action, clinical activity, and resistance. *Mol Cancer Ther.* 2009;8(8):2086–95.
- Pasquier E, Kavallaris M. Microtubules: A dynamic target in cancer therapy. *IUBMB Life*. 2008;60(3):165–70.
- Keating MJ, Holmes R, Lerner S, Ho DH. L-Asparaginase and PEG Asparaginase—Past, Present, and Future. *Leukemia & Lymphoma*. 1993; 10(s1):153–7.
- Schrey D, Speitel K, Lanvers-Kaminsky C, Gerss J, Möricke A, Boos J. Five-year single-center study of asparaginase therapy within the ALL-BFM 2000 trial. *Pediatric Blood & Cancer*. 2011;57(3):378–84.
- 47. Gahrton G, Engstedt L, Franzén S, et al. Induction of remission with l-asparaginase, cyclophosphamide, cytosine arabinoside, and prednisolone in adult patients with acute leukemia. *Cancer*. 1974;34(2):472–9.
- Asselin BL, Ryan D, Frantz CN, et al. In Vitro and in Vivo Killing of Acute Lymphoblastic Leukemia Cells by L-Asparaginase. *Cancer Research*. 1989; 49(15):4363–8.
- Asselin BL. L-asparaginase for treatment of childhood acute lymphoblastic leukemia: What have we learned? *Pediatr. Blood Cancer*. 2011;57:357–58.
- Planey SL, Litwack G. Glucocorticoid-induced apoptosis in lymphocytes. Biochem Biophys Res Commun. 2000;279:307–12.
- Csoka M, Bocsi J, Falus A, et al. Glucocorticoid-induced apoptosis and treatment sensitivity in acute lymphoblastic leukemia of children. *Pediatr Hematol Oncol.* 1997;14:433–42.
- Thompson EB, Nazareth LV, Thulasi R, Ashraf J, Harbour D, Johnson BH. Glucocorticoids in malignant lymphoid cells: gene regulation and the minimum receptor fragment for lysis. *J. Steroid Biochem Mol Biol.* 1992;41:273–82.
- 53. Aricò M, Basso G, Mandelli F, et al. Good steroid response in vivo predicts a favorable outcome in children with T-cell acute lymphoblastic leukemia. The Associazione Italiana Ematologia Oncologia Pediatrica (AIEOP). *Cancer.* 1995;75(7):1684–93.
- 54. Labar B, Suciu S, Willemze R, et al. Dexamethasone compared to prednisolone for adults with acute lymphoblastic leukemia or lymphoblastic lymphoma: final results of the ALL-4 randomized, phase III trial of the EORTC Leukemia Group. *Haematologica*. 2010;95(9):1489–95.
- Bansal N, Houle A, Melnykovych G. Apoptosis: mode of cell death induced in T-cell leukemia lines by dexamethasone and other agents. *FASEB J*. 1991;5:211–6.
- Thomas X. Chemotherapy of acute leukemia in adults. Expert Opin Pharmacother. 2009;10(2):221–37.
- 57. Bourquin JP, Izraeli S. Where can biology of childhood ALL be attacked by new compounds? *Cancer Treat Rev.* 2010;36(4):298–306.
- Fullmer A, O'Brien S, Kantarjian H, Jabbour E. Novel therapies for relapsed acute lymphoblastic leukemia. *Current Hematologic Malignancy Reports*. 2009;4(3):148–56.
- Stegmeier F, Warmuth M, Sellers WR, Dorsch M. Targeted Cancer Therapies in the Twenty-First Century: Lessons From Imatinib. *Clin Pharmacol Ther*. 2010;87(5):543–52.
- Druker BJ. Imatinib as a Paradigm of Targeted Therapies. Adv Cancer Res. 2004;91:1–30.
- Tremblay CS, Hoang T, Hoang T. Early T cell differentiation: lessons from T-cell acute lymphoblastic leukemia. *Progress in Molecular Biology and Translational Science*. 2010;92:121–56.
- Bhandoola A, von Boehmer H, Petrie HT, Zúñiga-Pflücker JC. Commitment and developmental potential of extrathymic and intrathymic T cell precursors: plenty to choose from. *Immunity*. 2007;26(6):678–89.
- Staal F, van Dongen J, Langerak A. Novel insights into the development of T-cell acute lymphoblastic leukemia. *Curr Hematol Malig Rep.* 2007; 2(3):176–82.
- Van Vlierberghe P, Pieters R, Beverloo HB, Meijerink JPP. Moleculargenetic insights in paediatric T-cell acute lymphoblastic leukaemia. *Br J Haematol*. 2008;143(2):153–68.
- Ferrando AA, Neuberg DS, Staunton J, et al. Gene expression signatures define novel oncogenic pathways in T cell acute lymphoblastic leukemia. *Cancer Cell*. 2002;1(1):75–87.



- Armstrong SA, Look AT. Molecular Genetics of Acute Lymphoblastic Leukemia. J Clin Oncol. 2005;23(26):6306–15.
- Mellentin JD, Smith SD, Cleary ML. lyl-1, a novel gene altered by chromosomal translocation in T cell leukemia, codes for a protein with a helix-loop-helix DNA binding motif. *Cell*. 1989;58(1):77–83.
- Soulier J, Clappier E, Cayuela J-M, et al. HOXA genes are included in genetic and biologic networks defining human acute T-cell leukemia (T-ALL). *Blood*. 2005;106(1):274–86.
- 69. Van Grotel M, Meijerink JP, Beverloo HB, et al. The outcome of molecularcytogenetic subgroups in pediatric T-cell acute lymphoblastic leukemia: a retrospective study of patients treated according to DCOG or COALL protocols. *Haematol.* 2006;91(9):1212–1.
- Clappier E, Cuccuini W, Kalota A, et al. The C-MYB locus is involved in chromosomal translocation and genomic duplications in human T-cell acute leukemia (T-ALL), the translocation defining a new T-ALL subtype in very young children. *Blood*. 2007;110(4):1251–61.
- Xia Y, Brown L, Yang CY, et al. TAL2, a helix-loop-helix gene activated by the (7;9)(q34;q32) translocation in human T-cell leukemia. *Proc Natl Acad Sci U S A*. 1991;88(24):11416–20.
- Ferrando AA, Neuberg DS, Dodge RK, et al. Prognostic importance of TLX1 (HOX11) oncogene expression in adults with T-cell acute lymphoblastic leukaemia. *Lancet*. 2004;363(9408):535–6.
- Graux C, Cools J, Melotte C, et al. Fusion of NUP214 to ABL1 on amplified episomes in T-cell acute lymphoblastic leukemia. *Nat Genet.* 2004; 36(10):1084–9.
- Look AT. Oncogenic transcription factors in the human acute leukemias. Science. 1997;278(5340):1059–64.
- 75. Rabbitts TH. Translocations, master genes, and differences between the origins of acute and chronic leukemias. *Cell*. 1991;67(4):641–4.
- Weng AP, Ferrando AA, Lee W, et al. Activating mutations of NOTCH1 in human T cell acute lymphoblastic leukemia. *Science*. 2004;306(5694): 269–71.
- Mansur MB, Emerenciano M, Splendore A, Brewer L, Hassan R, Pombo-de-Oliveira MS. T-cell lymphoblastic leukemia in early childhood presents NOTCH1 mutations and MLL rearrangements. *Leukemia Res.* 2010;34(4):483–6.
- Ferrando AA. The role of NOTCH1 signaling in T-ALL. *Hematology*. Jan 1, 2009;(1):353–61.
- Pear WS, Aster JC. T cell acute lymphoblastic leukemia/lymphoma: a human cancer commonly associated with aberrant NOTCH1 signaling. *Current Opinion in Hematology*. 2004;11(6):426–33.
- Palomero T, Sulis ML, Cortina M, et al. Mutational loss of PTEN induces resistance to NOTCH1 inhibition in T-cell leukemia. *Nat Med.* 2007;13(10):1203–10.
- Jiang BH, Liu LZ. Chapter 2 PI3K/PTEN signaling in angiogenesis and tumorigenesis. Adv Cancer Res. 2009;102:19–65.
- Steelman LS, Abrams SL, Whelan J, et al. Contributions of the Raf/MEK/ ERK, PI3K/PTEN/Akt/mTOR and Jak/STAT pathways to leukemia. *Leukemia*. 2008;22(4):686–707.
- Palomero T, Ferrando A. Therapeutic targeting of NOTCH1 signaling in T-cell acute lymphoblastic leukemia. *Clin Lymphoma Myeloma*. 2009;9(S3): S205–10.
- Rao SS, O'Neil J, Liberator CD, et al. Inhibition of NOTCH Signaling by Gamma Secretase Inhibitor Engages the RB Pathway and Elicits Cell Cycle Exit in T-Cell Acute Lymphoblastic Leukemia Cells. *Cancer Research*. Apr 1, 2009;69(7):3060–8.
- Paganin M, Ferrando A. Molecular pathogenesis and targeted therapies for NOTCH1-induced T-cell acute lymphoblastic leukemia. *Blood reviews*. 2010;25(2):83–90.
- Yin L, Velazquez OC, Liu Z-J. Notch signaling: Emerging molecular targets for cancer therapy. *Biochemical Pharmacology*. 2010;80(5):690–701.
- Sohn SJ, Rajpal A, Winoto A. Apoptosis during lymphoid development. *Current Opinion in Immunology*. 2003;15(2):209–16.
- Giovannetti A, Pierdominici M, Di Iorio A, et al. Apoptosis in the homeostasis of the immune system and in human immune mediated diseases. *Curr Pharm Des.* 2008;14(3):253–68.

- Smets LA. Programmed cell death (apoptosis) and response to anti-cancer drugs. Anti-Cancer Drugs. 1994;5(1):3–9.
- Hannun YA. Apoptosis and the Dilemma of Cancer Chemotherapy. Blood. 1997;89(6):1845–53.
- Tsurusawa M, Saeki K, Fujimoto T. Differential induction of apoptosis on human lymphoblastic leukemia Nalm-6 and Molt-4 cells by various antitumor drugs. *Int J Hematol.* 1997;66(1):79–88.
- Budihardjo I, Oliver H, Lutter M, Luo X, Wang X. Biochemical pathways of caspase activation during apoptosis. *Ann Rev Cell Dev Biol.* 1999; 15(1):269–90.
- Gross A, McDonnell JM, Korsmeyer SJ. BCL-2 family members and the mitochondria in apoptosis. *Genes Dev.* 1999;13(15):1899–911.
- Brunelle JK, Letai A. Control of mitochondrial apoptosis by the Bel-2 family. J Cell Sci. 2009;122(4):437–41.
- Schmidt S, Rainer J, Ploner C, Presul E, Riml S, Kofler R. Glucocorticoidinduced apoptosis and glucocorticoid resistance: molecular mechanisms and clinical relevance. *Cell Death Differ*. 2004;11(S1):S45–55.
- Haarman EG, Kaspers GJ, Pieters R, et al. BCL-2 expression in childhood leukemia versus spontaneous apoptosis, drug induced apoptosis, and in vitro drug resistance. *Adv Exp Med Biol*. 1999;457:325–3.
- 97. Gandhi V, Plunkett W. Clofarabine and nelarabine: two new purine nucleoside analogs. *Current Opinion in Oncology*. 2006;18(6):584–90.
- Rodriguez CO, Stellrecht CM, Gandhi V. Mechanisms for T-cell selective cytotoxicity of arabinosylguanine. *Blood*. 2003;102(5):1842–8.
- Cooper TM. Role of nelarabine in the treatment of T-cell acute lymphoblastic leukemia and T-cell lymphoblastic lymphoma. *Ther Clin Risk Manag.* 2007;3(6):1135–41.
- Cohen MH, Johnson JR, Justice R, Pazdur R. FDA Drug Approval Summary: Nelarabine (Arranon) for the Treatment of T-Cell Lymphoblastic Leukemia/Lymphoma. *Oncologist*. 2008;13(6):709–14.
- 101. Commander LA, Seif AE, Insogna IG, Rheingold SR. Salvage therapy with nelarabine, etoposide, and cyclophosphamide in relapsed/refractory paediatric T-cell lymphoblastic leukaemia and lymphoma. *British Journal* of Haematology. 2010;150(3):345–51.
- Reilly KM, Kisor DF. Profile of nelarabine: use in the treatment of T-cell acute lymphoblastic leukemia. *Onco Targets Ther.* 2009;2:219–28.
- 103. DeAngelo DJ, Yu D, Johnson JL, et al. Nelarabine induces complete remissions in adults with relapsed or refractory T-lineage acute lymphoblastic leukemia or lymphoblastic lymphoma: Cancer and Leukemia Group B study 19801. *Blood.* 2007;109(12):5136–42.
- Buie LW, Epstein SS, Lindley CM. Nelarabine: A novel purine antimetabolite antineoplastic agent. *Clinical Therapeutics*. 2007;29(9):1887–99.
- Chahar M, Sharma N, Dobhal M, Joshi Y. Flavonoids: A versatile source of anticancer drugs. *Pharmacognosy Reviews*. 2011;5(9):1–12.
- Jackman KM, Frye CB, Hunger SP. Flavopiridol displays preclinical activity in acute lymphoblastic leukemia. *Pediatric Blood & Cancer*. 2008;50(4):772–8.
- 107. Gojo I, Zhang B, Fenton RG. The Cyclin-dependent Kinase Inhibitor Flavopiridol Induces Apoptosis in Multiple Myeloma Cells through Transcriptional Repression and Down-Regulation of Mcl-1. *Clinical Cancer Research*. 2002;8(11):3527–38.
- Chen R, Keating MJ, Gandhi V, Plunkett W. Transcription inhibition by flavopiridol: mechanism of chronic lymphocytic leukemia cell death. *Blood.* 2005;106(7):2513–9.
- Beesley AH, Palmer M-L, Ford J, et al. In vitro cytotoxicity of nelarabine, clofarabine and flavopiridol in paediatric acute lymphoblastic leukaemia. *British Journal of Haematology*. 2007;137(2):109–16.
- Real PJ, Tosello V, Palomero T, et al. Gamma-secretase inhibitors reverse glucocorticoid resistance in T cell acute lymphoblastic leukemia. *Nat Med.* 2009;15(1):50–8.
- 111. Wei P, Walls M, Qiu M, et al. Evaluation of Selective r-Secretase Inhibitor PF-03084014 for Its Antitumor Efficacy and Gastrointestinal Safety to Guide Optimal Clinical Trial Design. *Molecular Cancer Therapeutics*. 2010;9(6):1618–28.
- Cullion K, Draheim KM, Hermance N, et al. Targeting the Notch1 and mTOR pathways in a mouse T-ALL model. *Blood*. 2009;113(24):6172–81.



- 113. Chiang MY, Xu L, Shestova O, et al. Leukemia-associated NOTCH1 alleles are weak tumor initiators but accelerate K-ras-initiated leukemia. *J Clin Invest.* 2008;118(9):3181–94.
- Liu H, Chiang M, Pear W. Critical roles of NOTCH1 in acute T-cell lymphoblastic leukemia. *International Journal of Hematology*. 2011;94(2): 118–25.
- 115. Tatarek J, Cullion K, Ashworth T, Gerstein R, Aster JC, Kelliher MA. Notch1 inhibition targets the leukemia-initiating cells in a Tal1/Lmo2 mouse model of T-ALL. *Blood*. 2011;118(6):1579–1590.
- 116. Tammam J, Ware C, Efferson C, et al. Down-regulation of the Notch pathway mediated by a γ-secretase inhibitor induces anti-tumour effects in mouse models of T-cell leukaemia. *British Journal of Pharmacology*. 2009;158(5):1183–95.
- 117. Clument MV, Hirpara JL, Chawdhury S-H, Pervaiz S. Chemopreventive Agent Resveratrol, a Natural Product Derived From Grapes, Triggers CD95 Signaling-Dependent Apoptosis in Human Tumor Cells. *Blood.* 1998;92(3):996–1002.
- Cecchinato V, Chiaramonte R, Nizzardo M, et al. Resveratrol-induced apoptosis in human T-cell acute lymphoblastic leukaemia MOLT-4 cells. *Biochemical Pharmacology*. 2007;74(11):1568–74.
- 119. Kurzrock R, Kantarjian HM, Druker BJ, Talpaz M. Philadelphia Chromosome-positive Leukemias: From Basic Mechanisms to Molecular Therapeutics. *Annals of Internal Medicine*. 2003;138(10):819–30.
- Hagemeijer A, Graux C. ABL1 rearrangements in T-Cell acute lymphoblastic leukemia. *Genes, Chromosomes and Cancer.* 2010;49(4): 299–308.
- 121. Nishihara T, Miura Y, Tohyama Y, et al. Effects of the Tyrosine Kinase Inhibitor Imatinib Mesylate on a Bcr-Abl-Positive Cell Line: Suppression of Autonomous Cell Growth but No Effect on Decreased Adhesive Property and Morphological Changes. *International Journal of Hematology*. 2003;78(3):233–40.
- 122. Schrappe M, Arico M, Harbott J, et al. Philadelphia chromosome-positive (Ph+) childhood acute lymphoblastic leukemia: good initial steroid response allows early prediction of a favorable treatment outcome. *Blood*. 1998;92:2730–41.
- Ottmann OG, Wassmann B. Treatment of Philadelphia Chromosome-Positive Acute Lymphoblastic Leukemia. *Hematology*. 2005;(1):118–22.
- 124. De Labarthe A, Rousselot P, Huguet-Rigal F, et al. Imatinib combined with induction or consolidation chemotherapy in patients with de novo Philadelphia chromosome-positive acute lymphoblastic leukemia: Results of the GRAAPH-2003 study. *Blood.* 2007;109(4): 1408–13.
- 125. Schultz KR, Bowman WP, Aledo A, et al. Improved early event-free survival with imatinib in Philadelphia chromosome-positive acute lymphoblastic leukemia: A Children's Oncology Group Study. *Journal of Clinical Oncology*. 2009;27(31):5175–81.
- Thomas X, Dombret H. Treatment of Philadelphia chromosome-positive adult acute lymphoblastic leukemia. *Leukemia & Lymphoma*. 2008;49(7): 1246–54.
- Quintas-Cardama A, Tong W, Manshouri T, et al. Activity of tyrosine kinase inhibitors against human NUP214-ABL1-positive T cell malignancies. *Leukemia*. 2008;22(6):1117–24.
- 128. Deenik W, Beverloo HB, van der Poel-van de Luytgaarde SCPAM, et al. Rapid complete cytogenetic remission after upfront dasatinib monotherapy in a patient with a NUP214-ABL1-positive T-cell acute lymphoblastic leukemia. *Leukemia*. 2008;23(3):627–9.
- Tauchi T, Ohyashiki K. The Second Generation of BCR-ABL Tyrosine Kinase Inhibitors. *International Journal of Hematology*. 2006;83(4): 294–300.
- Pinilla-Ibarz J, Quintas-Cardama A. New Agents in the Treatment of Chronic Myelogenous Leukemia. *Journal of the National Comprehensive Cancer Network*. 2009;7(9):1028–37.
- 131. Kantarjian H, Cortes J, Kim D-W, et al. Phase 3 study of dasatinib 140 mg once daily versus 70 mg twice daily in patients with chronic myeloid leukemia in accelerated phase resistant or intolerant to imatinib: 15-month median follow-up. *Blood.* 2009;113(25):6322–9.

- 132. Quintas-Cardama A, Kantarjian H, Cortes J. Targeting ABL and SRC kinases in chronic myeloid leukemia: experience with dasatinib. *Future Oncol.* 2006;2(6):655–65.
- Tuma RS. With Targeted Drugs, Chronic Myelogenous Leukemia Therapy May Follow HIV's Model. J Natl Cancer Inst. 2007;99(3):192–4.
- Wong S-F. Dasatinib dosing strategies in Philadelphia chromosome-positive leukemia. *Journal of Oncology Pharmacy Practice*. 2009;15(1):17–27.
- Gibbons JJ, Abraham RT, Yu K. Mammalian target of rapamycin: discovery of rapamycin reveals a signaling pathway important for normal and cancer cell growth. *Semin Oncol.* 2009;36(suppl 3):S3–17.
- Dancey JE, Curiel R, Purvis J. Evaluating Temsirolimus Activity in Multiple Tumors: A Review of Clinical Trials. Semin oncol. 2009;36:S46–58.
- Gu L, Zhou C, Liu H, et al. Rapamycin sensitizes T-ALL cells to dexamethasone-induced apoptosis. J Exp Clin Cancer Res. 2010;29(1):150.
- 138. Pulsipher MA, Wall DA, Grimley M, et al. A Phase I/II study of the safety and efficacy of the addition of sirolimus to tacrolimus/methotrexate graft versus host disease prophylaxis after allogeneic haematopoietic cell transplantation in paediatric acute lymphoblastic leukaemia (ALL). *Bri J Haematol.* 2009;147(5):691–9.
- Chiarini F, Grimaldi C, Ricci F, et al. Activity of the Novel Dual Phosphatidylinositol 3-Kinase/Mammalian Target of Rapamycin Inhibitor NVP-BEZ235 against T-Cell Acute Lymphoblastic Leukemia. *Cancer Res.* 2010;70(20):8097–107.
- 140. Chiarini F, Fal F, Tazzari PL, et al. Dual Inhibition of Class IA Phosphatidylinositol 3-Kinase and Mammalian Target of Rapamycin as a New Therapeutic Option for T-Cell Acute Lymphoblastic Leukemia. *Cancer Res.* 2009;69(8):3520–8.
- 141. Miller A, Garza A, Johnson B, Thompson EB. Pathway interactions between MAPKs, mTOR, PKA, and the glucocorticoid receptor in lymphoid cells. *Cancer Cell International*. 2007;7(1):3.
- 142. Konopleva M, Watt J, Contractor R, et al. Mechanisms of Antileukemic Activity of the Novel Bcl-2 Homology Domain-3 Mimetic GX15–070 (Obatoclax). *Cancer Res.* 2008;68(9):3413–20.
- 143. Schimmer AD, O'Brien S, Kantarjian H, et al. A Phase I Study of the Pan Bcl-2 Family Inhibitor Obatoclax Mesylate in Patients with Advanced Hematologic Malignancies. *Clin Cancer Res.* 2008;14(24):8295–301.
- 144. O'Brien SM, Claxton DF, Crump M, et al. Phase I study of obatoclax mesylate (GX15–070), a small molecule pan-bcl-2 family antagonist, in patients with advanced chronic lymphocytic leukemia. *Blood*. 2009;113(2): 299–305.
- 145. Heidari N, Hicks MA, Harada H. GX15–070 (obatoclax) overcomes glucocorticoid resistance in acute lymphoblastic leukemia through induction of apoptosis and autophagy. *Cell Death Dis.* 2010;1(9):e76.
- 146. Nguyen M, Marcellus RC, Roulston A, et al. Small molecule obatoclax (GX15–070) antagonizes MCL-1 and overcomes MCL-1-mediated resistance to apoptosis. *Proc Natl Acad Sci U S A*. Dec 4, 2007;104(49): 19512–7.
- 147. Chen S, Dai Y, Pei XY, Grant S. Bim Upregulation by Histone Deacetylase Inhibitors Mediates Interactions with the Bcl-2 Antagonist ABT-737: Evidence for Distinct Roles for Bcl-2, Bcl-xL, and Mcl-1. *Mol Cell Biol.* 2009;29(23):6149–69.
- 148. Deng J, Carlson N, Takeyama K, Dal Cin P, Shipp M, Letai A. BH3 Profiling Identifies Three Distinct Classes of Apoptotic Blocks to Predict Response to ABT-737 and Conventional Chemotherapeutic Agents. *Cancer cell*. 08/14 2007;12(2):171–85.
- Iayanthan A, Incoronato A, Singh A, et al. Cytotoxicity, drug combinability, and biological correlates of ABT-737 against acute lymphoblastic leukemia cells with MLL rearrangement. *Pediatr Blood Cancer*. 2010; 56(3):353–60.
- 150. High LM, Szymanska B, Wilczynska-Kalak U, et al. The Bcl-2 Homology Domain 3 Mimetic ABT-737 Targets the Apoptotic Machinery in Acute Lymphoblastic Leukemia Resulting in Synergistic in Vitro and in Vivo Interactions with Established Drugs. *Mol Pharmacol.* 2010;77(3):483–94.
- Del Gaizo Moore V, Schlis KD, Sallan SE, Armstrong SA, Letai A. BCL-2 dependence and ABT-737 sensitivity in acute lymphoblastic leukemia. *Blood.* 2008;111(4):2300–9.



- 152. Pui C-H, Jeha S. New therapeutic strategies for the treatment of acute lymphoblastic leukaemia. *Nat Rev Drug Discov*. 2007;6(2):149–65.
- 153. Advani AS, Lazarus HM, Rosenblatt J, Avigan D. Immunotherapy for Acute Lymphocytic Leukemia. *Adult Acute Lymphocytic Leukemia*: Humana Press; 2010:351–63.
- 154. Geng Cd, Vedeckis WV. Use of Recombinant Cell-Permeable Small Peptides To Modulate Glucocorticoid Sensitivity of Acute Lymphoblastic Leukemia Cells. *Biochemistry*. 09/12 2010;49(41):8892–901.