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## STUDIES ON THE SYNTHESIS OF NICOTINAMIDE NUCLEOSIDE AND NUCLEOTIDE ANALOGUES AND THEIR INHIBITIONS TOWARDS CD38 NADASE

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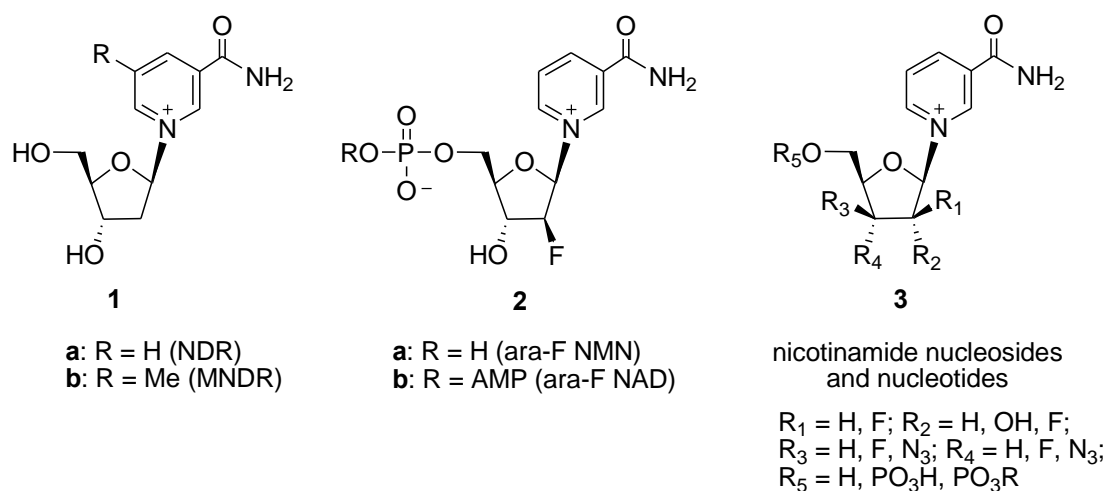
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**Abstract** – Nicotinamide adenine dinucleotide (NAD) analogues inhibit the NADase activity of CD38. In the current study, efficient protocols for the synthesis of substituted-nicotinamide nucleosides and nucleotides were developed. The one-pot phosphorylation–esterification strategy provides a convenient way of obtaining nicotinamide nucleoside phosphodiester from the corresponding nucleosides. Structure–activity relationship information revealed that replacement of 3'-hydroxy group with F or N<sub>3</sub> led to the considerably decrease of activity as compared with ara-F NMN. Phosphodiesterification of nicotinamide nucleosides lowers their inhibitory activities in some extent.

### INTRODUCTION

Nicotinamide adenine dinucleotide (NAD) analogues are gaining importance in the discovery of therapeutic agents because of their broad spectrum of pharmacological and biological activities. Thiazole-4-carboxamide adenine dinucleotide, benzamide adenine dinucleotide, and mycophenolic adenine dinucleotide have been reported to be potent inhibitors of inosine monophosphate dehydrogenase.<sup>1-3</sup> Nicotinamide hypoxanthine dinucleotide and nicotinamide guanine dinucleotide are potent inhibitors of ADP-ribosyl cyclase.<sup>4</sup> Carba-NAD has been shown to have inhibitory activities against NAD-dependent histone deacetylase.<sup>5</sup> Our research aimed to discover inhibitors of CD38 which is the main mammalian ADP-ribosyl cyclase and signaling enzyme responsible for catalyzing the synthesis

of two  $\text{Ca}^{2+}$  messengers, i.e., cyclic ADP-ribose (cADPR) and nicotinic acid adenine dinucleotide phosphate (NAADP).<sup>6</sup> CD38 inhibitors can be classified as covalent and noncovalent types based on their modes of binding to CD38. The former often displays stronger activity than the latter. Among the covalent inhibitors (**1**, **2**) (Figure 1),<sup>7-9</sup> fluorinated NAD analogues are the most intensively studied because derivatization with fluorine enhances the activity, lipophilicity, metabolic stability, and so on.<sup>10-13</sup> 1-(2'-Deoxy-2'-fluoro- $\beta$ -D-arabinofuranosyl) nicotinamide 5'-phosphate (**2a**, ara-F NMN) inhibits the NADase activity of CD38 in the nanomolar range.<sup>8</sup> 2'-Deoxy-2'-fluoro-NAD (**2b**, ara-F NAD) was used to capture the enzymatic reaction intermediates of mutated CD38 and label the extracellular domain of human CD38.<sup>14,15</sup>



**Figure 1.** Structures of nicotinamide nucleoside and nucleotide analogues (**1-3**)

The synthesis of nucleotides that carry positively charged constituents still remains challenging even though many strategies have been developed to assemble nucleotides using either P(III) or P(V) reagents in nucleic acid chemistry.<sup>16-19</sup> The existing strategies often resulted in tedious purification work and cleavage of the anomeric C-N glycosidic bond. Oppenheimer *et al.* firstly synthesized the 2'-substituted arabino-NAD analogues using 1,2:5,6-di-*O*-isopropylidene- $\alpha$ -D-allofuranose as a starting material, which allowed the stereospecific introduction of the fluoro and azido substituents.<sup>20</sup> Recently, Saue *et al.* described a strategy for diastereocontrolled electrophilic fluorination to synthesize 2'-fluoro-substituted NAD analogues.<sup>21</sup> Esterification of nucleotide enhances its lipophilicity and enzymatic stability. The esterification of ribonofuranosyl nicotinamide mononucleotide was investigated by using *N,N'*-dicyclohexylcarbodiimide (DCC)/4-dimethylaminopyridine (DMAP)<sup>22</sup> or the 2,4,6-triisopropylbenzenesulfonyl chloride (TIPS-Cl)/pyridine<sup>23</sup> coupling system. However, the research of the phosphoesters of nicotinamide nucleoside either on the structural diversity or on the biological property is far from well studied.

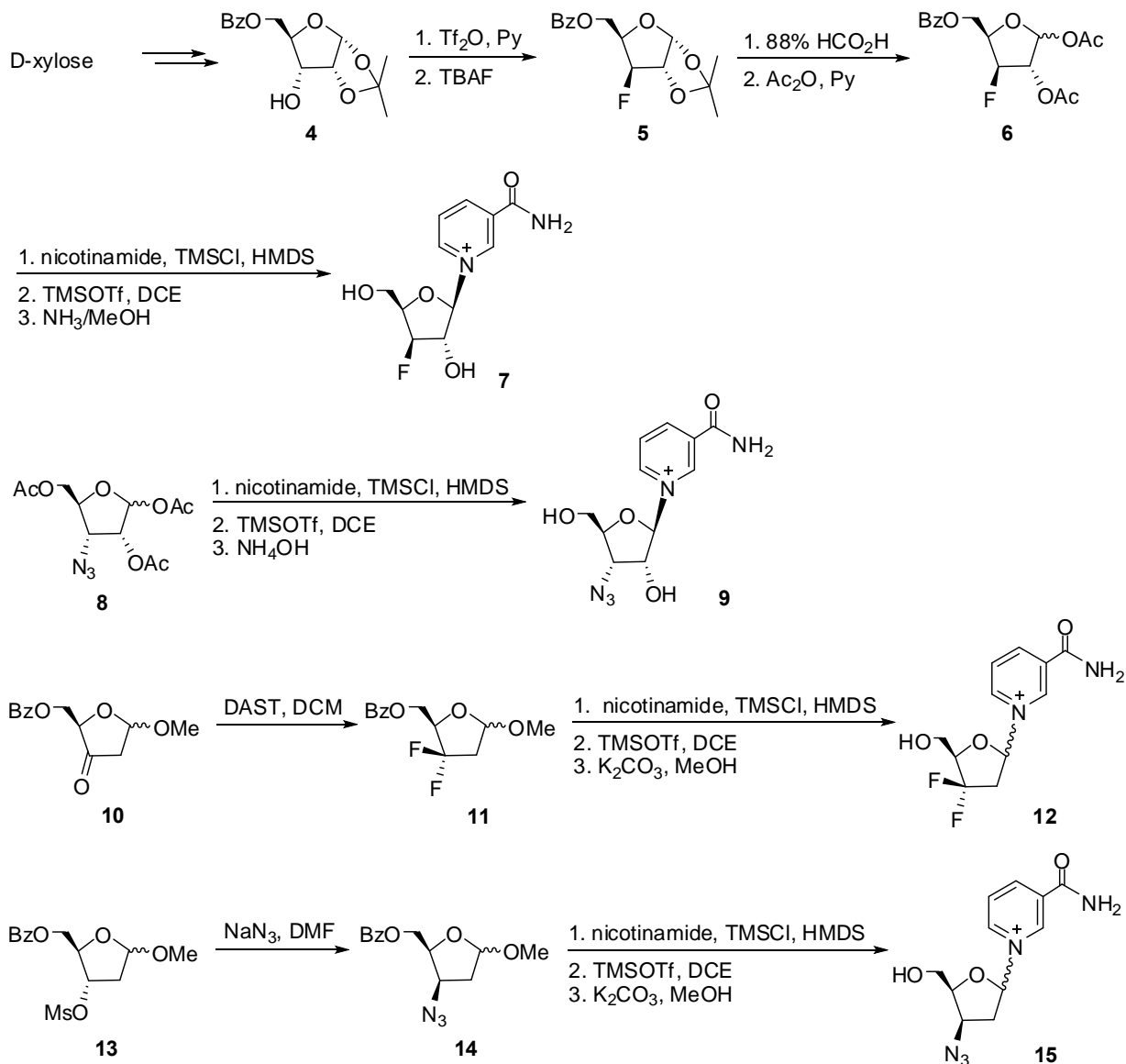
In this study, for providing compounds to investigate the effect of 3'-substitution on biological activity, the synthesis of 3'-substituted NAD analogues (**3**) is reported for the first time. A simple and efficient method of preparing nicotinamide nucleosides phosphodiester (**20-23**) is provided, in which phosphorylation and esterification of nucleosides were carried out in a one-pot reaction. These structurally diverse NAD analogues were used to study the inhibition to NADase of CD38. Findings in this study help in providing compounds to elucidate the structure-activity relationships of NAD analogues and investigate the physiological properties of human CD38.

## RESULTS AND DISCUSSION

Treatment of **4**<sup>24</sup> with trifluoromethanesulfonic anhydride (Tf<sub>2</sub>O) in pyridine at -10 °C followed by fluorination gave fluoroxylose **5** in 64% in two steps. The fluorination using tetrabutylammonium fluoride (TBAF) afforded fluororibose *via* S<sub>N</sub>2 reaction and resulted in the change of configuration of C-3. This fluorinating condition is milder than those using HF, DAST or other fluorinating reagents. The configuration of C-3 was confirmed by <sup>1</sup>H and <sup>19</sup>F NMR spectra, which are in accordance with the previous reports.<sup>24</sup> After hydrolysis of **5** with aqueous formic acid and the following acetylation, an anomeric mixture ( $\alpha:\beta = 1:1$ ) of compound **6** was obtained. Condensation of **6** with trimethylsilylated nicotinamide,<sup>25</sup> and the subsequent deprotection of the condensation product with NH<sub>3</sub>/MeOH provided 3'-deoxy-3'-fluoro- $\beta$ -D-xylofuranosylnicotinamide nucleoside (**7**) (Scheme 1). 3'- $\beta$ -D-azido substituted ribofuranosylnicotinamide nucleoside (**9**) was synthesized *via* directed coupling of **8**<sup>26</sup> with trimethylsilylated nicotinamide, followed by deprotection using a similar procedure as that of nucleoside **7**. Treatment of ketone **10**<sup>27</sup> with DAST afforded difluorodeoxyfuranoside **11** in 44% yield. Condensation of difluorosugar **11** with trimethylsilylated nicotinamide in the presence of TMSOTf afforded the corresponding nucleoside which was followed by deprotection using K<sub>2</sub>CO<sub>3</sub>/MeOH to provide 2',3'-dideoxy-3',3'-difluoro-D-*glycero*-pentofuranosylnicotinamide nucleoside (**12**) in 47% yield (Scheme 1). Compound **12** was obtained as an anomeric mixture that could be separated into the  $\alpha$ - and  $\beta$ -isomers ( $\alpha:\beta = 1:1$ ). 3'-Azido-2',3'-dideoxy-D-*thero*-furanosylnicotinamide nucleoside (**15**), an anomeric mixture ( $\alpha:\beta = 1:1$ ), was obtained using a similar procedure starting with azidofuranoside **14** which was prepared from the azidation of **13**.<sup>28</sup> Nucleoside **15** was not stable and decomposed during purification, which resulted in low yields (33%).

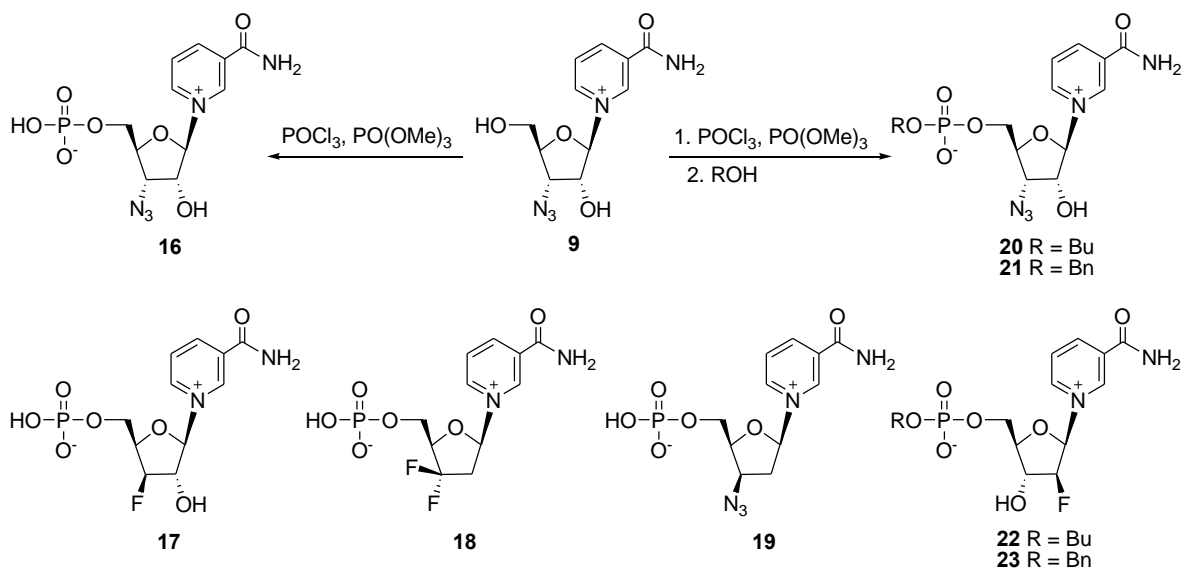
The phosphorylation of nicotinamide nucleoside analogues (**7**, **9**, **12** and **15**) was performed based on a modification of the procedure developed by Yoshikawa, which is generally used to synthesize nucleoside 5'-monophosphates (Scheme 2).<sup>29</sup> These nucleosides were dissolved in trimethyl phosphate and reacted with phosphoryl chloride, and then hydrolyzed to yield the corresponding mononucleotides **16-19** in

76-80% yield. The synthesis of ara-F NMN (**2a**) and its nucleoside moiety, 2'-deoxy-2'-fluoro- $\beta$ -D-arabinofuranosylnicotinamide (**3a**), was carried out according to the reported method.<sup>20</sup>



Scheme 1

Mononucleotides can be esterified by the DCC/DMAP<sup>22</sup> or the TIPS-Cl/pyridine<sup>23</sup> strategy. However, the process often requires a long reaction time and results in low yields. In the current study, a one-pot strategy was used to synthesize nicotinamide nucleoside phosphodiester. The nucleoside **9** was phosphorylated at 5'-OH with  $\text{POCl}_3$  at 0 °C. Afterward, butanol or benzyl alcohol was added to the reaction mixture, which was then stirred for 1-6 h, and then quenched with water and neutralized with 1 N NaOH, the corresponding nicotinamide nucleoside phosphodiester **20** and **21** were obtained, respectively. The butyl and benzyl esters (**22**, **23**) could be also prepared under the similar conditions in 56-62% yield.



Scheme 2

**Table 1.** CD38 NADase inhibitory activities of nicotinamide nucleosides (**2a**, **7**, **9**, **12**, **15**) and nucleotides (**16-23**)

Compound	IC <sub>50</sub> (μM)	Compound	IC <sub>50</sub> (μM)
<b>2a</b>	0.61	<b>17</b>	250
<b>3a</b>	>1000	<b>18(α)</b>	>1000
<b>7</b>	>1000	<b>18(β)</b>	>1000
<b>9</b>	>1000	<b>19(α)</b>	929
<b>12(α)</b>	>1000	<b>19(β)</b>	457
<b>12(β)</b>	>1000	<b>20</b>	>1000
<b>15(α)</b>	321	<b>21</b>	>1000
<b>15(β)</b>	843	<b>22</b>	2.17
<b>16</b>	100	<b>23</b>	3.06

The synthesized compounds have been tested for their inhibitory activities towards CD38 NADase (Table 1). The results show that nicotinamide nucleosides (**2a**, **3a**, **7**, **9**, **12**, **15**) do not have or only have very weak inhibitory activities. Phosphorylation of nicotinamide nucleosides improves activities as compounds **16**, **17** and **19** show, but in general the nicotinamide nucleoside monophosphates with modification at 3'-position only give very weak inhibition to CD38 NADase. By comparing with ara-F NMN (**2a**), it clearly shows that the modification at 3'-position will dramatically lower inhibitory activity. Esterification of nicotinamide nucleotide also leads to obvious decrease of inhibitory activity (**20-23**).

Considering the increase of stability towards the hydrolysis by phosphatase and the increase of membrane permeability after esterification, it may provide probe for investigating the physiological property of CD38.

## CONCLUSION

In summary, efficient protocols for the synthesis of nicotinamide 3'-substituted nucleosides and nucleotides have been developed. The one-pot phosphorylation–esterification strategy provides a convenient way of producing nicotinamide nucleoside phosphodiester from the corresponding nucleosides. Biological investigation indicates that the 3'-hydroxy group of D-ribose is crucial for the inhibitory activity. Phosphodiesterification of nicotinamide nucleosides obviously lowers their inhibitory activities. The newly synthesized compounds and related findings help to understand the structure–activity relationship of NAD analogues for the inhibition to CD38 NADase. The active compounds may be used as probes to elucidate the physiological properties of CD38.

## EXPERIMENTAL

HR-ESI-MS (electrospray ionization) were performed with Bruker BIFLEX III.  $^1\text{H}$ NMR and  $^{13}\text{C}$  NMR were recorded with a Bruker AVANCE III 400 or a JEOLAL300 spectrometer,  $\text{CDCl}_3$  or  $\text{D}_2\text{O}$  was used as a solvent. Chemical shifts are reported in parts per million downfield from TMS ( $^1\text{H}$  and  $^{13}\text{C}$ ).  $^{31}\text{P}$  NMR spectra were recorded at room temperature by use of JEOL AL300 spectrometer (122 MHz); Orthophosphoric acid (85%) was used as an external standard.  $^{19}\text{F}$  NMR spectra were recorded on a Varian VXR-500 spectrometer (470 MHz). Chemical shifts of  $^{19}\text{F}$  NMR are reported in ppm with reference to  $\text{CF}_3\text{CO}_2\text{H}$  (-75.6 relative to  $\text{CF}_3$ ) as an external standard. Most of the target compounds were purified twice on an Alltech preparative C18 reversed-phase column ( $2.2 \times 25$  cm) using a Gilson HPLC buffer system:  $\text{H}_2\text{O}/\text{MeOH}$ , 0.1%  $\text{CF}_3\text{CO}_2\text{H}$  (pH 2-3).

### 5-*O*-Benzoyl-3-deoxy-3-fluoro-1, 2-*O*-(isopropylidene)- $\alpha$ -D-xylofuranose (5)

Compound **4** (5.50 g, 18.69 mmol) was dissolved in 50 mL DCM which contains pyridine (6.30 mL, 7.53 mmol) and cooled to  $-30$  °C.  $\text{Ti}_2\text{O}$  (6.32 mL, 37.38 mmol) was added dropwise and then was stirred at  $-10$  °C for 2 h. 100 mL DCM was added to the reaction solution, and was washed with dilute HCl solution and saturated aqueous  $\text{NaHCO}_3$  solution, respectively. After dried with anhydrous  $\text{MgSO}_4$  and evaporated, the residue was dissolved in 80 mL THF and then TBAF· $3\text{H}_2\text{O}$  (11.79 g, 37.37 mmol) was stirred. The mixture was stirred at  $60$  °C for 10 h, and then was evaporated and purified by silica gel column chromatography (PE:EtOAc = 3:1) to give compound **5** as a colorless oil (3.54 g, 64% yield).  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.06 (d, 2H,  $J = 8.4$  Hz, Ar-H), 7.59-7.56 (m, 1H, Ar-H), 7.45 (t, 2H,  $J = 8.4$

Hz, Ar-H), 6.04 (d, 1H,  $J_{1,2} = 3.6$  Hz, H-1), 5.05 (dd, 1H,  $J_{3,4} = 2.0$ ,  $J_{3,F} = 50.8$  Hz, H-3), 4.76-4.70 (m, 1H, H-2), 4.65-4.52 (m, 3H, H-4, H-5), 1.51 (s, 3H, -CH<sub>3</sub>), 1.34 (s, 3H, -CH<sub>3</sub>). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 166.1, 133.2, 129.8, 129.6, 128.4, 112.4, 105.1, 94.0 (d,  $J_{C-3-F} = 138.0$  Hz, C-3), 82.5 (d,  $J_{C-4-F} = 24.0$  Hz, C-4), 77.7 (d,  $J_{C-2-F} = 14.3$  Hz, C-2), 61.3 (C-5), 26.7, 26.2. <sup>19</sup>F NMR (470 MHz, CDCl<sub>3</sub>, CF<sub>3</sub>Ph): δ -209.7 (m, CF). ESI: [M + Na]<sup>+</sup> 319.1, [M + K]<sup>+</sup> 335.0. MS (ESI-TOF<sup>+</sup>)  $m/z$  319.06 (M + Na)<sup>+</sup>, 335.03 (M + K)<sup>+</sup>.

### 1,2-Di-*O*-acetyl-5-*O*-benzoyl-3-deoxy-3-fluoro- $\alpha,\beta$ -D-xylofuranose (6)

Compound **5** (1.87 g, 5.50 mmol) in 88% HCO<sub>2</sub>H (10 mL) was reacted at 80 °C for 2 h, and then the solvent was removed under reduced pressure. The residue was dissolved in pyridine (10 mL), and was added Ac<sub>2</sub>O (5.20 mL, 55 mmol) dropwise at 0 °C. The reaction mixture was stirred at rt for 24 h and then cooled to 0 °C. The mixture was quenched by MeOH (1 mL) and partitioned between H<sub>2</sub>O and DCM. The organic layer was washed with cold dilute HCl, saturated aqueous NaHCO<sub>3</sub>, and brine, respectively, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and evaporated. The residue was purified by silica gel column chromatography (20–30% EtOAc in PE) to give compound **6** as a colorless oil (1.74 g, 81%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.07-8.03 (m, 2H, Ar-H), 7.58 (t,  $J = 7.2$  Hz, 1H, Ar-H), 7.46 (t,  $J = 7.2$  Hz, 2H, Ar-H), 6.57-6.20 (m, 1H, H-1), 5.57-5.08 (m, 2H, H-2, H-3), 4.69-4.49 (m, 3H, H-4, H-5), 2.13-2.05 (m, 6H, -COMe).

### 1-(3'-Deoxy-3'-fluoro- $\beta$ -D-xylofuranosyl) nicotinamide (7)

To a solution of dry nicotinamide (0.49 g, 4.01 mmol) in HMDS (4 mL) was added TMSCl (1.03 mL, 8.02 mmol) under argon, and the mixture was stirred at 120 °C for approximately 6 h, then the reaction mixture was evaporated to dryness at 90 °C under vacuum. A solution of **6** (1.36 g, 4.01 mmol) in DCE (6 mL) was added to the above residue at 45 °C and then TMSOTf (1.85 mL, 8.02 mmol) was added portionwise over 5 min. The resulting mixture was stirred at 45 °C for 2 h, and then was cooled to 0 °C and MeOH (2 mL) was added. The mixture was concentrated in vacuo, and then dissolved in methanolic ammonium solution (30 mL) and stirred at -10 °C for 24 h. After filtration of the solution, the filtrate was neutralized with cold dilute HCl and evaporated. The residue was partitioned between CHCl<sub>3</sub> and H<sub>2</sub>O. The aqueous layer was evaporated, and the residue was dissolved in 0.1% CF<sub>3</sub>CO<sub>2</sub>H buffer (1.0 mL), which was applied to a C18 reversed-phase column (2.2 × 25 cm). The column was developed using a linear gradient of 0–40% MeOH in CF<sub>3</sub>CO<sub>2</sub>H buffer (pH 3.5) within 38 min to give **7** as a colorless oil. Yield: 62%. <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O) δ 9.39 (s, 1H, H-2), 9.17 (d, 1H,  $J_{5,6} = 6.0$  Hz, H-6), 8.97 (d, 1H,  $J_{4,5} = 8.0$  Hz, H-4), 8.27 (dd, 1H, H-5), 6.36 (s, 1H, H-1'), 5.15 (d, 1H,  $J_{3',F} = 50.4$  Hz, H-3'), 4.75 (d, 1H,  $J_{4',F} = 11.6$  Hz, H-4'), 4.67-4.56 (m, 1H, H-2'), 3.93-3.91 (m, 2H, H-5'). <sup>13</sup>C NMR (100 MHz, D<sub>2</sub>O) δ

162.3, 145.1, 142.2, 141.0, 133.8, 128.1, 100.8 (C-1'), 94.5 (d,  $J_{C-3'-F} = 181.0$  Hz, C-3'), 86.0 (d,  $J_{C-4'-F} = 7.2$  Hz, C-4'), 79.9 (d,  $J_{C-2'-F} = 10.8$  Hz, C-2'), 57.7 (d,  $J_{C-5'-F} = 3.6$  Hz, C-5').  $^{19}\text{F}$  NMR (470 MHz,  $\text{D}_2\text{O}$ ,  $\text{CF}_3\text{CO}_2\text{H}$ ):  $\delta$  -201.1 (m, C-F). HRMS (ESI-TOF<sup>+</sup>) calcd for  $\text{C}_{11}\text{H}_{14}\text{N}_2\text{O}_4\text{F}$  ( $\text{M}^+$ ), 257.0932; found, 257.0928.

### 1-(3'-Azido-3'-deoxy- $\beta$ -D-ribofuranosyl) nicotinamide (9)

**9** was synthesized by a procedure similar to that for **7**, colorless oil. Yield: 62%.  $^1\text{H}$  NMR (400 MHz,  $\text{D}_2\text{O}$ )  $\delta$  9.48 (s, 1H, H-2), 9.14 (d, 1H,  $J_{5,6} = 6.4$  Hz, H-6), 8.87 (d, 1H,  $J_{4,5} = 8.0$  Hz, H-4), 8.16 (dd, 1H, H-5), 6.15 (d, 1H,  $J_{1',2'} = 4.4$  Hz, H-1'), 4.67 (t, 1H,  $J_{2',1'} = 4.4$  Hz, H-2'), 4.41-4.38 (m, 1H, H-3'), 4.22 (t, 1H,  $J = 5.2$  Hz, H-4'), 3.96 (dd, 1H, H-5',  $J = 13.2$  Hz, H-5'), 3.78 (dd, 1H, H-5').  $^{13}\text{C}$  NMR (100 MHz,  $\text{D}_2\text{O}$ )  $\delta$  166.1, 146.3, 143.1, 140.9, 134.5, 128.9, 100.1, 86.2, 78.2, 61.4, 60.7; HRMS (ESI-TOF<sup>+</sup>) calcd for  $\text{C}_{11}\text{H}_{14}\text{N}_5\text{O}_4$  ( $\text{M} + \text{H}$ )<sup>+</sup>, 280.1040; found, 280.1037.

### Methyl 5-O-benzoyl-2,3-dideoxy-3,3-difluoro- $\alpha,\beta$ -D-glycero-pentofuranoside (11)

To a solution of compound **10** (6.90 g, 27.6 mmol) in dry DCM (120 mL) were added DAST (15 mL, 113.2 mmol). The reaction mixture was stirred under reflux for 2 days and then cooled to 0 °C. The mixture was added to saturated aqueous  $\text{NH}_4\text{Cl}$  portionwise and stirred. Layers were separated, and the aqueous solution was extracted with DCM. The combined organic layers were washed with a saturated aqueous solution of  $\text{NaHCO}_3$  and then with brine, dried over  $\text{Na}_2\text{SO}_4$ , and concentrated to dryness. The residue was purified by silica gel column (10–15% EtOAc in PE) to give compound **11** as a colorless oil. Yield: 44%.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.07 (m, 2H, Ar-H), 7.56 (m, 1H, Ar-H), 7.44 (m, 2H, Ar-H), 5.16 (t, 1H, H-1), 4.62-4.41 (m, 3H, H-4, H-5), 3.42 and 3.40 (2 $\times$ s, 3H, -OCH<sub>3</sub>), 2.74-2.41 (m, 2H, H-2). MS (ESI-TOF<sup>+</sup>)  $m/z$  295.12 ( $\text{M} + \text{Na}$ )<sup>+</sup>.

### 1-(2', 3'-Dideoxy-3',3'-difluoro- $\alpha,\beta$ -D-glycero-pentofuranosyl) nicotinamide (12)

**12** was synthesized by a procedure similar to that for **7**, colorless oil. Yield: 47%. **12**( $\alpha$ ):  $^1\text{H}$  NMR (400 MHz,  $\text{D}_2\text{O}$ )  $\delta$  9.46 (s, 1H, H-2), 9.14 (t, 1H,  $J_{5,6} = 6.4$  Hz, H-6), 8.85 (d, 1H,  $J_{4,5} = 8.0$  Hz, H-4), 8.14 (dd, 1H, H-5), 6.62 (dd, 1H,  $J_{1',2'} = 4.8$ ,  $J_{1',F} = 6.8$  Hz, H-1'), 4.53-4.46 (m, 1H, H-4'), 3.98-3.89 (m, 2H, H-5'), 3.48-3.35 (m, 1H, H-2'), 3.00-2.88 (m, 1H, H-2').  $^{13}\text{C}$  NMR (100 MHz,  $\text{D}_2\text{O}$ )  $\delta$  166.2, 146.3, 143.5, 141.5, 134.4, 128.9, 94.9 (d,  $J_{C-1'-F} = 5.0$  Hz, C-1'), 83.8 (t,  $J_{C-4'-F} = 25.0$  Hz, C-4'), 58.3 (C-5'), 42.3 (t,  $J_{C-2'-F} = 25.0$  Hz, C-2'). HRMS (ESI-TOF<sup>+</sup>) calcd for  $\text{C}_{11}\text{H}_{13}\text{N}_2\text{O}_3\text{F}_2$  ( $\text{M}^+$ ), 259.0889; found, 259.0895. **12**( $\beta$ ):  $^1\text{H}$  NMR (400 MHz,  $\text{D}_2\text{O}$ )  $\delta$  9.29 (s, 1H, H-2), 9.06 (d, 1H,  $J_{5,6} = 6.0$  Hz, H-6), 8.85 (d, 1H,  $J_{4,5} = 8.0$  Hz, H-4), 8.16 (dd, 1H, H-5), 6.74 (d, 1H,  $J = 7.2$  Hz, H-1'), 4.80-4.76 (m, 1H, H-4'), 3.88-3.79 (m, 2H, H-5'), 3.46-3.31 (m, 1H, H-2'), 3.08-3.00 (m, 1H, H-2').  $^{13}\text{C}$  NMR (100 MHz,  $\text{D}_2\text{O}$ )  $\delta$  166.1, 146.0,



142.8, 140.6, 134.5, 129.0, 96.0 (d,  $J_{C-1'-F} = 8.0$  Hz, C-1'), 84.8 (dd,  $J_{C-4'-F} = 30.0, 25.0$  Hz, C-4'), 59.1 (t,  $J_{C-5'-F} = 5.0$  Hz, C-5'), 42.7 (t,  $J_{C-2'-F} = 25.0$  Hz, C-2'). HRMS (ESI-TOF<sup>+</sup>) calcd for C<sub>11</sub>H<sub>13</sub>N<sub>2</sub>O<sub>3</sub>F<sub>2</sub> (M<sup>+</sup>), 259.0888; found, 259.0886. <sup>19</sup>F NMR (470 MHz, D<sub>2</sub>O, CF<sub>3</sub>CO<sub>2</sub>H): δ -93.9 and -115.7 (m, C-F).

### Methyl 3-azido-5-O-benzoyl-2,3-dideoxy- $\alpha,\beta$ -D-threo-pentofuranoside (14)

To a solution of compound **13** (4.50 g, 13.62 mmol) in dry DMF (150 mL) was added NaN<sub>3</sub> (4.43 g, 68.1 mmol). The reaction mixture was stirred at 100 °C for 6 h and then cooled to rt. The mixture was filtered and the filtrate was evaporated to dryness. The residue was purified by silica gel column (10–30% EtOAc in PE) to give compound **14** as a yellow syrup. Yield: 90%. **14 $\alpha$** : <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.09-8.03 (m, 2H, Ar-H), 7.57 (t, 1H,  $J = 7.2$  Hz, Ar-H), 7.45 (m, 2H,  $J = 7.6$  Hz, Ar-H), 5.19 (dd, 1H,  $J = 2.8, 5.2$  Hz, H-1), 4.60-4.50 (m, 2H, H-3, H-4), 4.39 (dd, 1H,  $J = 5.2$  Hz, 10.8 Hz, H-5), 4.27-4.23 (m, 1H, H-5), 3.38 (s, 3H, -OCH<sub>3</sub>), 2.39-2.25 (m, 2H, H-2). MS (ESI-TOF<sup>+</sup>)  $m/z$  300.1 (M + Na)<sup>+</sup>. **14 $\beta$** : <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.07 (t, 2H,  $J = 7.2$  Hz), 7.57 (m, 1H,  $J = 7.6$  Hz), 7.45 (t, 2H,  $J = 7.6$  Hz), 5.08 (t, 1H,  $J = 4.8$  Hz), 4.62-4.51 (m, 2H), 4.44 (dd, 1H,  $J = 5.6, 12.4$  Hz), 4.31-4.28 (m, 1H), 3.40 (s, 3H), 2.39-2.42 (m, 2H). <sup>13</sup>C NMR (100 MHz, D<sub>2</sub>O) δ 166.2, 133.1, 129.9, 129.7, 128.4, 128.4, 104.5, 79.3, 64.5, 60.7, 55.4, 38.5. MS (ESI-TOF<sup>+</sup>)  $m/z$  300.1 (M + Na)<sup>+</sup>.

### 1-(3'-Azido-2',3'-deoxy- $\alpha,\beta$ -D-threo-pentofuranosyl) nicotinamide (15)

**15** was synthesized by a procedure similar to that for **12**, colorless oil. Yield: 33%. **15 $\alpha$** : <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O) δ 9.37 (s, 1H, H-2), 9.12 (d, 1H,  $J_{5,6} = 6.0$  Hz, H-6), 8.81 (d, 1H,  $J_{4,5} = 8.0$  Hz, H-4), 8.11 (dd, 1H, H-5), 6.47 (t, 1H,  $J_{1',2'} = 6.0$  Hz, H-1'), 4.51-4.46 (m, 2H, H-3', H-4'), 4.00-3.99 (m, 2H, H-5'), 3.11-3.04 (m, 1H, H-2'), 2.73-2.69 (m, 1H, H-2'). <sup>13</sup>C NMR (100 MHz, D<sub>2</sub>O) δ 163.3, 145.5, 143.0, 141.0, 134.0, 128.5, 97.3, 87.0, 60.7, 60.2, 41.1. **15 $\beta$** : <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O) δ 9.24 (s, 1H, H-2), 9.00 (d, 1H,  $J_{5,6} = 6.0$  Hz, H-6), 8.82 (d, 1H,  $J_{4,5} = 8.0$  Hz, H-4), 8.13 (dd, 1H, H-5), 6.56 (t, 1H,  $J = 6.0$  Hz, H-1'), 4.63-4.60 (m, 1H, H-3'), 4.53-4.49 (m, 1H, H-4'), 3.84-3.83 (m, 2H, H-5'), 3.14-3.08 (m, 1H, H-2'), 2.69-2.62 (m, 1H, H-2'). <sup>13</sup>C NMR (100 MHz, D<sub>2</sub>O) δ 163.5, 145.7, 140.3, 134.4, 129.0, 97.4, 85.4, 61.9, 60.5, 41.3. HRMS (ESI-TOF<sup>+</sup>) calcd for C<sub>11</sub>H<sub>14</sub>N<sub>5</sub>O<sub>3</sub> (M<sup>+</sup>), 264.1091; found, 264.1092.

### General procedure for preparing 2'- and 3'-substituted nicotinamide-5'-mononucleotides (2a, 16-19)

To a solution of 3'-substituted nicotinamide nucleoside **9** (0.36 mmol) in PO(OMe)<sub>3</sub> (2.0 mL) at 0 °C was added POCl<sub>3</sub> (1.78 mmol), and the mixture was stirred at the same temperature for 1-2 h, then neutralized with 1N NaOH at 0 °C till pH = 7. The crude product was separated from the reaction mixture by addition of Et<sub>2</sub>O, and the precipitated salt was further purified using a C18 reversed-phase column (2.2 × 25 cm).

The column was eluted with a linear gradient of 15-30% MeOH in TFA buffer (pH 3.0) within 40 min to give **16** as colorless foam. Compound **2a** was synthesized according to the reported literature.<sup>20</sup> Mononucleotides **17**, **18** and **19** were synthesized by a procedure similar to that for **16**.

#### **1-(3'-Azido-3'-deoxy-β-D-ribofuranosyl) nicotinamide 5'-phosphate (16)**

Yield: 80%. <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O) δ 9.25 (s, 1H, H-2), 9.06 (d, 1H,  $J_{5,6} = 6.0$  Hz, H-6), 8.81 (d, 1H,  $J_{4,5} = 7.6$  Hz, H-4), 8.11 (dd, 1H, H-5), 6.05 (d, 1H,  $J_{1',2'} = 5.2$  Hz, H-1'), 4.63 (t, 1H, H-2'), 4.46 (d, 1H,  $J = 2.4$  Hz, H-3'), 4.29 (dd, 1H,  $J = 3.6, 5.6$  Hz, H-4'), 4.22-4.17 (m, 1H, H-5'), 4.04-4.00 (m, 1H, H-5'). <sup>13</sup>C NMR (100 MHz, D<sub>2</sub>O) δ 166.1, 146.6, 142.9, 140.3, 134.5, 129.1, 99.9, 85.3, 78.3, 65.1, 62.6. <sup>31</sup>P NMR (122 MHz, D<sub>2</sub>O) δ -0.87(s). HRMS (ESI-TOF<sup>+</sup>) calcd for C<sub>11</sub>H<sub>14</sub>N<sub>5</sub>O<sub>7</sub>P (M + H)<sup>+</sup>, 360.0704; found, 360.0701.

#### **1-(3'-Deoxy-3'-fluoro-β-D-xylofuranosyl) nicotinamide 5'-phosphate (17)**

Yield: 79%. <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O) δ 9.27 (s, 1H, H-2), 9.06 (d, 1H,  $J_{5,6} = 6.0$  Hz, H-6), 8.78 (d, 1H,  $J_{4,5} = 7.6$  Hz, H-4), 8.09 (dd, 1H, H-5), 6.23 (s, 1H, H-1'), 5.14 (dt, 1H,  $J_{3',F} = 50.4$  Hz, H-3'), 4.82-4.73 (m, 2H, H-2', H-4'), 4.33-4.22 (m, 2H, H-5'). <sup>13</sup>C NMR (100 MHz, D<sub>2</sub>O) δ 166.1, 146.2, 142.9, 140.8, 134.3, 128.9, 100.9 (C-1'), 94.5 (d,  $J_{C-3',F} = 183$  Hz, C-3'), 84.1 (d,  $J_{C-4',F} = 19$  Hz, C-4'), 80.3 (d,  $J_{C-2',F} = 28.0$  Hz, C-2'), 62.5 (d,  $J_{C-5',F} = 10$  Hz, C-5'). <sup>19</sup>F NMR (470 MHz, D<sub>2</sub>O, CF<sub>3</sub>CO<sub>2</sub>H) δ -201.6 (m, C-F). <sup>31</sup>P NMR (122 MHz, D<sub>2</sub>O) δ 0.54 (s). HRMS (ESI-TOF<sup>+</sup>) calcd for C<sub>11</sub>H<sub>14</sub>N<sub>2</sub>O<sub>7</sub>F (M + H)<sup>+</sup>, 337.0595; found, 337.0598.

#### **1-(2', 3'-Dideoxy-3',3'-difluoro-α,β-D-glycero-pentofuranosyl) nicotinamide 5'-phosphate (18)**

Yield: 71%. **18(α)**: <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O) δ 9.34 (s, 1H, H-2), 9.16 (d, 1H,  $J_{5,6} = 6.4$  Hz, H-6), 8.84 (d, 1H,  $J_{4,5} = 8.0$  Hz, H-4), 8.16 (dd, 1H, H-5), 6.61 (t, 1H,  $J = 6.4$  Hz, H-1'), 4.71-4.63 (m, 1H, H-4'), 4.22-4.09 (m, 2H, H-5'), 3.52-3.39 (m, 1H, H-2'), 3.07-2.90 (m, 1H, H-2'). <sup>13</sup>C NMR (100 MHz, D<sub>2</sub>O) δ 166.3, 146.4, 143.4, 141.0, 134.4, 128.9, 94.7 (d,  $J_{C-1',F} = 5.0$  Hz, C-1'), 82.8 (dd,  $J_{C-4',F} = 26.0, 9.0$  Hz, C-4'), 62.4 (C-5'), 42.2 (t,  $J_{C-2',F} = 25.0$  Hz, C-2'). <sup>31</sup>P NMR (122 MHz, D<sub>2</sub>O) δ -0.11 (s). <sup>19</sup>F NMR (470 MHz, D<sub>2</sub>O, CF<sub>3</sub>CO<sub>2</sub>H): δ -97.9 and -113.5 (m, C-F). HRMS (ESI-TOF<sup>+</sup>) calcd for C<sub>11</sub>H<sub>13</sub>N<sub>2</sub>O<sub>6</sub>F<sub>2</sub>P (M + H)<sup>+</sup>, 339.0552; found, 339.0550. **18(β)**: <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O) δ 9.31 (s, 1H, H-2), 9.08 (d, 1H,  $J_{5,6} = 6.0$  Hz, H-6), 8.85 (d, 1H,  $J_{4,5} = 8.0$  Hz, H-4), 8.16 (dd, 1H, H-5), 6.80 (d, 1H,  $J = 7.6$  Hz, H-1'), 4.94 (d, 1H,  $J = 15.2$  Hz, H-4'), 4.11-4.05 (m, 2H, H-5'), 3.51-3.36 (m, 1H, H-2'), 3.09-2.94 (m, 1H, H-2'). <sup>13</sup>C NMR (100 MHz, D<sub>2</sub>O) δ 166.2, 146.1, 142.9, 140.8, 134.5, 129.0, 96.3 (d,  $J_{C-1',F} = 10.0$  Hz, C-1'), 83.7 (C-4'), 63.0 (C-5'), 42.6 (t,  $J_{C-2',F} = 25.0$  Hz, C-2'). <sup>31</sup>P NMR (122 MHz, D<sub>2</sub>O) δ 0.49 (s). <sup>19</sup>F NMR (470

MHz, D<sub>2</sub>O, CF<sub>3</sub>CO<sub>2</sub>H):  $\delta$  -92.3 and -116.1 (m, C-F). HRMS (ESI-TOF<sup>+</sup>) calcd for C<sub>11</sub>H<sub>13</sub>N<sub>2</sub>O<sub>6</sub>F<sub>2</sub>P (M + H)<sup>+</sup>, 339.0552; found, 339.0546.

**1-(3'-Azido-2',3'-dideoxy- $\alpha,\beta$ -D-threo-pentofuranosyl) nicotinamide 5'-phosphate (19)**

Yield: 78%. **19**( $\alpha$ ): <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O)  $\delta$  9.32 (s, 1H, H-2), 9.15 (d, 1H,  $J_{5,6}$  = 6.0 Hz, H-6), 8.81 (d, 1H,  $J_{4,5}$  = 7.6 Hz, H-4), 8.11 (dd, 1H, H-5), 6.46 (t, 1H,  $J$  = 7.2 Hz, H-1'), 4.60-4.53 (m, 2H, H-3', H-4'), 4.26-4.21 (m, 2H, H-5'), 3.12-3.05 (m, 1H, H-2'), 2.72-2.68 (m, 1H, H-2'). <sup>13</sup>C NMR (100 MHz, D<sub>2</sub>O)  $\delta$  166.3, 145.6, 143.1, 140.8, 134.0, 128.5, 97.2, 85.3, 63.8, 60.6, 40.9. <sup>31</sup>P NMR (122 MHz, D<sub>2</sub>O)  $\delta$  0.70 (s). HRMS (ESI-TOF<sup>+</sup>) calcd for C<sub>11</sub>H<sub>13</sub>N<sub>5</sub>O<sub>6</sub>P (M + H)<sup>+</sup>, 344.0755; found, 344.0755. **19**( $\beta$ ): <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O)  $\delta$  9.25 (s, 1H, H-2), 9.03 (d, 1H,  $J_{5,6}$  = 6.4 Hz, H-6), 8.82 (d, 1H,  $J_{4,5}$  = 8.4 Hz, H-4), 8.13 (dd, 1H, H-5), 6.58 (t, 1H,  $J_{1',2'}$  = 6.4 Hz, H-1'), 4.78-4.74 (m, 1H, H-3'), 4.58-4.52 (m, 1H, H-4'), 4.10-4.04 (m, 2H, H-5'), 3.14-3.08 (m, 1H, H-2'), 2.68-2.61 (m, 1H, H-2'). <sup>13</sup>C NMR (100 MHz, D<sub>2</sub>O)  $\delta$  166.2, 145.8, 142.7, 140.3, 134.4, 129.0, 97.4, 83.9, 64.0, 61.8, 41.1. <sup>31</sup>P NMR (243 MHz, D<sub>2</sub>O)  $\delta$  0.76 (s). HRMS (ESI-TOF<sup>+</sup>) calcd for C<sub>11</sub>H<sub>14</sub>N<sub>5</sub>O<sub>6</sub>P (M + H)<sup>+</sup>, 344.0755; found, 344.0754.

**1-(2'-Deoxy-2'-fluoro- $\beta$ -D-arabinofuranosyl) nicotinamide 5'-phosphate (2a)**

Yield: 76%. <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O)  $\delta$  9.25 (s, 1H, H-2), 9.08 (t, 1H,  $J_{5,6}$  = 5.6 Hz, H-6), 8.81 (d, 1H,  $J_{4,5}$  = 8.0 Hz, H-4), 8.10 (dd, 1H, H-5), 6.56 (dd, 1H,  $J_{1',2'}$  = 4.4,  $J_{1',F}$  = 10.4 Hz, H-1'), 5.36 (dd, 1H,  $J_{2',F}$  = 61.2 Hz, H-2'), 4.43 (dt, 1H,  $J$  = 17.2 Hz, H-3'), 4.28-4.27 (m, 1H, H-4'), 4.21-4.04 (m, 2H, H-5'). <sup>13</sup>C NMR (100.6 MHz, D<sub>2</sub>O)  $\delta$  165.5, 146.3, 143.6, 141.4, 133.7, 128.2, 94.6 (d,  $J_{C-2',F}$  = 196.0 Hz, C-2'), 94.1 (d,  $J_{C-1',F}$  = 16 Hz, C-1'), 83.9 (t,  $J_{C-4',F}$  = 5.0 Hz, C-4'), 71.4 (d,  $J_{C-3',F}$  = 25.0 Hz, C-3'), 63.4 (C-5'); <sup>19</sup>F NMR (D<sub>2</sub>O, CF<sub>3</sub>CO<sub>2</sub>H):  $\delta$  -199.2 (m, C-F). <sup>31</sup>P NMR (243.0 MHz, D<sub>2</sub>O)  $\delta$  0.76 (s). HRMS (ESI-TOF<sup>+</sup>) calcd for C<sub>11</sub>H<sub>15</sub>N<sub>2</sub>O<sub>7</sub>FP (M + H)<sup>+</sup>, 337.0601; found (M + H)<sup>+</sup>, 337.0591.

**General procedure for preparing 2'- and 3'-substituted nicotinamide nucleoside phosphodiester (20-23)**

To a solution of nicotinamide nucleoside **9** (0.36 mmol) in PO(OMe)<sub>3</sub> (2.0 mL) at 0 °C was added POCl<sub>3</sub> (1.78 mmol), and the mixture was stirred at the same temperature for 1-2 h. Then corresponding alcohol (3.6 mmol) was added and stirred for 1-2 h, then neutralized with 1N NaOH at 0 °C till pH = 7. The crude product was separated from the reaction mixture by addition of Et<sub>2</sub>O, and the precipitated salt was further purified using a C18 reversed-phase column (2.2 × 25 cm). The column was eluted with a linear gradient of 15-60% MeOH in TFA buffer (pH 3.0) within 30-80 min to give phosphodiester (**20** and **21**) as a colorless foam. Compound **22** and **23** were synthesized using a similar method as **20** and **21**.

**1-(3'-Azido-3'-deoxy- $\beta$ -D-ribofuranosyl) nicotinamide 5'-(butyl phosphate) (20)**

Yield: 62%.  $^1\text{H}$  NMR (400 MHz,  $\text{D}_2\text{O}$ )  $\delta$  9.32 (s, 1H, H-2), 9.13 (d, 1H,  $J_{5,6} = 6.4$  Hz, H-6), 8.87 (d, 1H,  $J_{4,5} = 8.4$  Hz, H-4), 8.18 (dd, 1H, H-5), 6.11 (d, 1H,  $J_{1',2'} = 5.6$  Hz, H-1'), 4.70 (t, 1H,  $J = 5.6$  Hz, H-2'), 4.52 (t, 1H,  $J = 2.4$  Hz, H-3'), 4.36 (dd, 1H,  $J = 3.2, 5.2$  Hz, H-4'), 4.22-4.18 (m, 1H, H-5'), 4.05-4.00 (m, 1H, H-5'), 3.76-3.71 (m, 2H,  $-\text{CH}_2-$ ), 1.46-1.41 (m, 2H,  $-\text{CH}_2-$ ), 1.23-1.18 (m, 2H,  $-\text{CH}_2-$ ), 0.75 (t, 3H,  $J = 7.2$  Hz,  $-\text{CH}_3$ ).  $^{13}\text{C}$  NMR (100 MHz,  $\text{D}_2\text{O}$ )  $\delta$  166.1, 146.7, 142.9, 140.2, 134.5, 129.0, 99.8, 85.6, 78.4, 66.8, 65.0, 62.9, 32.4, 18.8, 13.4.  $^{31}\text{P}$  NMR (122 MHz,  $\text{D}_2\text{O}$ )  $\delta$  -0.49 (s). HRMS (ESI-TOF $^+$ ) calcd for  $\text{C}_{15}\text{H}_{22}\text{N}_5\text{O}_7\text{P}$  ( $\text{M}^+$ ), 416.1330; found, 416.1327.

**1-(3'-Azido-3'-deoxy- $\beta$ -D-ribofuranosyl) nicotinamide 5'-(benzyl phosphate) (21)**

Yield: 56%.  $^1\text{H}$  NMR (400 MHz,  $\text{D}_2\text{O}$ )  $\delta$  9.18 (s, 1H, H-2), 8.99 (d, 1H,  $J_{5,6} = 6.4$  Hz, H-6), 8.74 (d, 1H,  $J_{4,5} = 8.4$  Hz, H-4), 8.03 (dd, 1H, H-5), 7.24-7.20 (m, 5H, Ar-H), 5.99 (d, 1H,  $J_{1',2'} = 5.6$  Hz, H-1'), 4.75 (d, 1H,  $J = 8.8$  Hz,  $\text{PhCH}_2-$ ), 4.50 (t, 1H,  $J = 5.6$  Hz, H-2'), 4.42-4.39 (m, 1H, H-3'), 4.13-4.09 (m, 1H, H-4'), 3.97-3.94 (m, 1H, H-5'), 3.91-3.87 (m, 1H, H-5').  $^{13}\text{C}$  NMR (100 MHz,  $\text{D}_2\text{O}$ )  $\delta$  165.9, 146.5, 142.7, 140.0, 134.3, 129.3, 129.0, 128.9, 128.2, 99.7, 85.5, 78.3, 68.4, 65.2, 62.8.  $^{31}\text{P}$  NMR (122 MHz,  $\text{D}_2\text{O}$ )  $\delta$  -0.70 (s). HRMS (ESI-TOF $^+$ ) calcd for  $\text{C}_{18}\text{H}_{20}\text{N}_5\text{O}_7\text{P}$  ( $\text{M}^+$ ), 450.1173; found, 450.1173.

**1-(2'-Deoxy-2'-fluoro- $\beta$ -D-arabinofuranosyl) nicotinamide 5'-(butyl phosphate) (22)**

Yield: 61%.  $^1\text{H}$  NMR (400 MHz,  $\text{D}_2\text{O}$ ):  $\delta$  9.32 (s, 1H, H-2), 9.17 (d, 1H,  $J_{5,6} = 6.4$  Hz, H-6), 8.87 (d, 1H,  $J_{4,5} = 8.0$  Hz, H-4), 8.17 (dd, 1H, H-5), 6.62 (dd, 1H,  $J_{1',2'} = 4.4, J_{1',\text{F}} = 12.8$  Hz, H-1'), 5.42 (dt, 1H,  $J_{2',\text{F}} = 50.8$  Hz, H-2'), 4.50 (dt, 1H,  $J = 5.2, 17.2$  Hz, H-3'), 4.32-4.31 (m, 1H, H-4'), 4.18-4.17 (m, 1H, H-5'), 4.10-4.04 (m, 1H, H-5'), 3.77 (dd, 2H,  $J = 6.8, 13.6$  Hz,  $-\text{CH}_2-$ ), 1.45 (t, 2 H,  $J = 7.2$  Hz,  $-\text{CH}_2-$ ), 1.16-1.22 (m, 2H,  $-\text{CH}_2-$ ), 0.71 (t, 3H,  $J = 7.6$  Hz,  $-\text{CH}_3$ ).  $^{13}\text{C}$  NMR (100 MHz,  $\text{D}_2\text{O}$ )  $\delta$  166.0, 146.9, 144.1, 141.9, 134.2, 128.8, 95.2 (d,  $J_{\text{C}-2',\text{F}} = 197.0$  Hz, C-2'), 94.6 (d,  $J_{\text{C}-1',\text{F}} = 15$  Hz, C-1'), 84.5 (C-4'), 71.9 (d,  $J_{\text{C}-3',\text{F}} = 24.0$  Hz, C-3'), 66.9, 63.6 (C-5'), 32.4, 18.8, 13.4.  $^{31}\text{P}$  NMR (122 MHz,  $\text{D}_2\text{O}$ ):  $\delta$  0.05 (s).  $^{19}\text{F}$  NMR (470 MHz,  $\text{D}_2\text{O}$ ,  $\text{CF}_3\text{CO}_2\text{H}$ ):  $\delta$  -198.8 (m, C-F). HRMS (ESI-TOF $^+$ ): calcd for  $\text{C}_{15}\text{H}_{23}\text{N}_2\text{O}_7\text{FP}$  ( $\text{M} + \text{H}$ ) $^+$  393.1221, found, 393.1224.

**1-(2'-Deoxy-2'-fluoro- $\beta$ -D-arabinofuranosyl) nicotinamide 5'-(benzyl phosphate) (23)**

Yield: 60%.  $^1\text{H}$  NMR (400 MHz,  $\text{D}_2\text{O}$ ):  $\delta$  9.30 (s, 1H, H-2), 9.12 (t, 1H,  $J_{5,6} = 6.4$  Hz, H-6), 8.84 (d, 1H,  $J_{4,5} = 8.0$  Hz, H-4), 8.08 (dd, 1H, H-5), 7.31-7.20 (m, 5H, Ar-H), 6.62 (dd, 1H,  $J_{1',2'} = 4.8, J_{1',\text{F}} = 10.4$  Hz, H-1'), 5.45 (dt, 1H,  $J_{2',\text{F}} = 50.8$  Hz, H-2'), 4.86 (d, 2H,  $J = 7.6$  Hz,  $\text{PhCH}_2-$ ), 4.42 (dt, 1H,  $J = 4.8, 17.6$  Hz, H-3'), 4.32-4.30 (m, 1H, H-4'), 4.20-4.02 (m, 2H, H-5').  $^{13}\text{C}$  NMR (100 MHz,  $\text{D}_2\text{O}$ ):  $\delta$  165.8, 146.8,

144.0, 141.8, 137.6, 133.9, 129.2, 129.1, 128.8, 128.7, 128.3, 95.1 (d,  $J_{C-2'-F} = 196.0$  Hz, C-2'), 94.5 (d,  $J_{C-1'-F} = 16.0$  Hz, C-1'), 84.6 (C-4'), 71.9 (d,  $J_{C-3'-F} = 24.0$  Hz, C-3'), 68.4, 63.8 (C-5').  $^{19}\text{F}$  NMR (470 MHz,  $\text{D}_2\text{O}$ ,  $\text{CF}_3\text{CO}_2\text{H}$ ):  $\delta$  -198.7 (m, C-F).  $^{31}\text{P}$  NMR (122 MHz,  $\text{D}_2\text{O}$ ):  $\delta$  0.89 (s). HRMS (ESI-TOF<sup>+</sup>): calcd for  $\text{C}_{18}\text{H}_{20}\text{N}_2\text{O}_7\text{FP}$  (M + H)<sup>+</sup> 427.1065, found 427.1047.

### Antagonist Assay

NADase activity was performed by recombinant CD38 prepared by a yeast expression system as described in reference.<sup>30</sup> Recombinant CD38 (0.015  $\mu\text{g}/\text{mL}$ ) and BSA (50  $\mu\text{g}/\text{ml}$ ) were added to different concentrations of antagonist and incubated for 2 h at room temperature in dark. NAD (2  $\mu\text{M}$ ) was added to initiate the reactions and aliquots of samples were collected at T = 0, 4, 8 and 12 mins respectively. Samples were immediately stopped with equal volume of 0.6 M HCl followed by neutralization with 2 times volume of 0.5 M sodium phosphate buffer (pH = 8). The amounts of NAD were measured by cycling assay.<sup>31</sup> NADase activity was showed as the percentage of inhibition as compared to control without antagonist.

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