Unique pattern of mutations in $\beta$-thalassemia patients in Western Uttar Pradesh

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INTRODUCTION

$\beta$-thalassemia is the most common autosomal single gene disorder across the globe.[1,2] This disorder is caused by a deficient ($\beta^+$ thalassemia) or absent ($\beta^-$ thalassemia) synthesis of one or more of the globin chains. Until date, more than 380 different types of mutations causing this disease have been identified in Online Mendalian Inheritance in Man (OMIM) 2005. The endemic area of the disease is the “thalassemia belt,” which includes the Mediterranean region, part of the Middle-East, the Indian subcontinent and the South-East Asia.[3,4] Immigration plays a major role in both the distribution and the extent of mutations across the globe.[5] There are approximately 45 million carriers with a carrier rate of approximately 1:20, of $\beta$-thalassemia in South Asian countries which include India, Sri Lanka, and Pakistan.[6] Treatment of individuals with $\beta$-thalassemia major, which entails regular blood transfusions and expensive iron chelation regimen, is still not satisfactory. Thus, the disease causes significant morbidity and mortality in affected individuals.[7] $\beta$-thalassemia is a major public-health problem and a burden on the society. The cost for blood
transfusions alone for the disease has been projected at ~3,200 USD (equivalent to approximately 1,76,000 Indian rupees) per child, while the lifetime health-care for a person born with the disease has been projected at 284,154 USD (1,56,28,000 Indian rupees).\[8\]

Though, \(\beta\)-thalassemia is observed in most global populations, each population or sub population has its own unique spectrum of mutations.\[9\] In India, the estimated number of \(\beta\)-thalassemia heterozygous patients is about 29.7 million with a rate of about 7000 homozygous children born with this disease per year.\[10\] The frequency of the heterozygous carriers reaches as high as 40% in some tribal groups, whereas, the incidence of the disease is around 3-4% in the general population.\[11,12\] About 92% of \(\beta\)-thalassemia allele in India is accounted by common mutations such as IVS 1-5 (c. 92 + 5 G > C), IVS 1-1 (c. 92 + 1 G > T), 619 bp deletion, codon 8/9 (c. 27_28insG), and codon 41/42 (c.124_127delTTCT).\[13\] The most common mutation across India is IVS 1-5 (c. 92 + 5 G > C) as also found in Pakistan and Sri Lanka. These mutations are not uniformly distributed across India but have geographic specificity, and each ethnic group has some common mutations and a variable number of rare ones. Regional differences in prevalence across northern, eastern and southern parts of India have also been demonstrated. In the northern states of India, the mutation and allele frequencies are quite similar to those in Pakistan where the two most common mutations are IVS 1-5 (43.5%) and codon 8/9 (38.5%). While in South India, codon 41/42 (c. 124_127delTTCT) is prevalent, in Eastern India mutation HBE (codon 26 G-A) (c. 79G > A) is frequently found.\[15\]

The \(\beta\)-globin chain is encoded by the hemoglobin \(\beta\) (HBB) gene. The \(\beta\)-globin locus harbors, in addition to the HBB gene, the \(\varepsilon\)-globin (HBE) gene, the two \(\gamma\)-globin (HBG) genes (HBG-G and HBG-A) and the \(\delta\)-globin (HBD) gene. The most genes at this locus are provided on the distribution and the frequency of \(\beta\)-thalassemia alleles is useful to establish a program for carrier screening, genetic counseling, prenatal diagnosis, and for physicians to establish specific therapeutic approaches for patients suffering from the disease. Due to lack of information regarding the type of \(\beta\)-thalassemia mutations encountered in Western U.P., an effort was
made to characterize the mutations for \( \beta \)-thalassemia by ARMS-hot start PCR.

**Materials And Methods**

**Specimens**

The sample consisted of 48 unrelated individuals (32 males and 16 females) with age ranging between 2 years and 16 years. The patients included in this study were all transfusion dependent. Age, sex, history, and consanguinity between the parents were recorded by reviewing the patients' files after obtaining permission from their parents. The age at presentation of the disease was as early as 6 months. Most of these patients had received transfusion 14-16 times/year, but only a few of them had received transfusion at irregular intervals due to economic and other constraints. Samples of venous blood (2 ml) from each patient was collected in tubes containing ethylene diamine tetraacetic acid (EDTA) (Volex, Italy) prior to blood transfusion during thalassemia camps organized by the Thalassemia Society, Bareilly between July 2011 and May 2012.

**DNA extraction and characterization of mutations by ARMS-PCR**

DNA was isolated from whole blood using HiPurA blood genomic DNA midiprep purification Spin Kit (Himedia, Mumbai). RNA was digested with 200 \( \mu \text{l} \) of RNase A solution (20 mg/ml, Himedia, Mumbai). The isolated DNA was evaluated by agarose gel electrophoresis after staining with ethidium bromide and exposure to ultraviolet (UV) light and stored at – 20\(^\circ\)C until mutation analysis. All the samples were then screened for the five most common mutations previously reported in Indian population by ARMS-hot start PCR.

Hot start-PCR reactions were carried in two separate tubes for each sample, one tube for the amplification by the normal ARMS primer and the 2\(^{nd}\) for the amplification by the mutant ARMS primer. However, amplification in both tubes was carried out using a common reverse primer. To monitor false-negative results, an internal PCR control, which amplified another region of the genome was included in the reaction. ARMS-PCR primers were synthesized by Bangalore genei, Bangalore. The sequences of allele-specific oligonucleotide primers (normal and mutant) are listed in Table 1.

A total of 25 \( \mu \text{l} \) final PCR reaction volume was made in two separate tubes for each sample. The reaction mixture comprised 0.5 \( \mu \text{g} \) of the DNA template, 10 pmol of each of the four primers (two control primers, one mutant ARMS primer or one normal ARMS primer and its corresponding reverse primer for the reaction) in a solution of 10 mM Tris-HCl, 50 mM KCl and 1.2 mM MgCl\(_2\) (Himedia, Mumbai). For hot start PCR, Taq DNA polymerase 2.5 units (Himedia, Mumbai) and 0.2 mM of each deoxyribonucleotide triphosphate (Bangalore

**Table 1: Oligonucleotide primers used for mutation detection by ARMS**

<table>
<thead>
<tr>
<th>Primer name</th>
<th>ARMS primer (5’-3’)</th>
<th>Mutation detected</th>
<th>Amplified product size (bp)</th>
</tr>
</thead>
<tbody>
<tr>
<td>BG-3 (N)</td>
<td>CTC CTT AAA CCT GTC TTG TAA CCT TGT TAssC</td>
<td>IVS1-5ntG-C (N)</td>
<td>285</td>
</tr>
<tr>
<td>BG-4 (M)</td>
<td>CTC CTT AA A CCT GTC TTG TAA CCT TGT TAG</td>
<td>IVS1-5ntG-C (M)</td>
<td>285</td>
</tr>
<tr>
<td>BG-5 (N)</td>
<td>CCT TGC CCC ACA GGG CAG TAA CGG CAC ACT</td>
<td>Cd.8/9+G (N)</td>
<td>225</td>
</tr>
<tr>
<td>BG-6 (M)</td>
<td>CCT TGC CCC ACA GGG CAG TAA CGG CAC ACC</td>
<td>Cd.8/9+G (M)</td>
<td>225</td>
</tr>
<tr>
<td>BG-7 (N)</td>
<td>GAG TGG ACA GAT CCC CAA AGG ACT CAA AGA</td>
<td>Cd.41/42 (−TCTT) (N)</td>
<td>439</td>
</tr>
<tr>
<td>BG-8 (M)</td>
<td>GAG TGG ACA GAT CCC CAA AGG ACT CAA CCT</td>
<td>Cd.41/42 (−TCTT) (M)</td>
<td>439</td>
</tr>
<tr>
<td>BG-1 (N)</td>
<td>GAT GAA GTT GGT GGT GAC GCC CTG GGT AGG</td>
<td>IVS1-Int.G-T (N)</td>
<td>450</td>
</tr>
<tr>
<td>BG-2 (M)</td>
<td>TTA AAC CTG TCT TGT AAC CTT CAT GAC AAA</td>
<td>IVS1-Int.G-T (M)</td>
<td>281</td>
</tr>
<tr>
<td>BG-9 (N)</td>
<td>TAA CCT TGA TAC CAA CCT GCC CAG GCC GTC</td>
<td>HBE Cd.26G-A (N)</td>
<td>236</td>
</tr>
<tr>
<td>BG-10 (M)</td>
<td>TAA CCT TGA TAC CAA CCT GCC CAG GCC GTT</td>
<td>HBE Cd.26G-A (M)</td>
<td>236</td>
</tr>
<tr>
<td>BG 11 Int.C1</td>
<td>CAA TGT ATC ATG CCT CTT TGC ACC</td>
<td>PCR internal control primers (861 bp)</td>
<td></td>
</tr>
<tr>
<td>BG 12 Int.C2</td>
<td>GAG TCA AGG CTG AGA GAT GCA GGA</td>
<td></td>
<td></td>
</tr>
<tr>
<td>BG 13 Rev.Pr</td>
<td>ACC TCA CCC TGT GGA GCC AC</td>
<td>Common reverse primer A</td>
<td></td>
</tr>
<tr>
<td>*BG 14 Rev.Pri B</td>
<td>CCC CTT CCT ATG ACA TGA ACT TAA</td>
<td>Reverse primer B</td>
<td></td>
</tr>
</tbody>
</table>

*For BG 1 IVS1-1 (N) only, PCR: Polymerase chain reaction, bp: Base pair, ARMS: Amplification refractory mutation system, IVS: Intervening Sequences; HBE: \( \varepsilon \) globin gene, BG: \( \beta \) globin
Christopher, et al.: Screening of common β-thalassemia mutations in western Uttar Pradesh

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genei, Bangalore) were added into each tube after the initial denaturation step (96°C for 1 min) on a thermal cycler (XP thermal cycler, Bioer, Japan). The reaction tubes were then subjected to thermal cycling regimen consisting of 25 cycles; denaturation at 94°C for 1 min, primer annealing at 65°C for 1 min and extension at 72°C for 1.5 min with the final extension at 72°C for 3 min.

The electrophoresis condition was as follows. Fifteen microlitres of the PCR products was removed and mixed with 3 μl of a loading buffer and then loaded on a 2% agarose gel and the gel was set at 100 volts for 1 h and then stained with ethidium bromide. After staining, the bands were visualized under UV light. Images of the gels were acquired on DigiDoc-It™ imaging system (UVP, USA) and molecular weight of the PCR products (test mutation) were determined by comparing them with the 100 base pair (bp) DNA marker using the Doc-It® LS image acquisition and analysis software (UVP, USA).

Results

The DNA extracted [Figure 1] from the whole blood samples of transfusion dependent β-thalassemia patients were subjected to ARMS-hot start PCR for detection of mutations. This technique is one of the most commonly used techniques for the diagnosis of the disease. The ARMS-hot start PCR results revealed the presence of 4 mutations [Table 2]. Internal control band of 861 bp was present in all ARMS-PCR reactions. Presence of an amplified product by the mutant ARMS primer on 2% agarose gel indicated the presence of the mutant allele. In the present study, we were able to identify the different β-thalassemia mutations in 28 patients (58%), while mutations were uncharacterized in 20 patients (42%). The reason for the uncharacterized mutations in these patients could be the presence of mutation other than the five selected mutations or that these patients harbored other hemoglobinopathies. Molecular analysis showed that IVS1-5(G-C) was the commonest β-thalassemic mutation (46%) followed by Fr8/9 (+G), which had a frequency of 21%. The frequency of other mutations was IVS1-1 (12%) and Cd 41/42 (4%). We were not able to find mutation for Cd26 (HBE), which is the 2nd most widespread mutation in the north eastern states of India [Table 2].

Discussion

Of the 48 patients, we were able to detect four different type of mutations viz. IVS 1-5, CD 41/42, Fr 8/9 and IVS 1-1 in 28 (58%) patients. The most common mutation found in our study was IVS 1-5 (46%), which is in conformity with the previously reported studies as the most common mutation in the Indian population.\[15,19\] Therefore, it can fairly be considered as the most common mutation in India.

Mutation Fr 8/9 was found to be the 2nd most common mutation in our study with a frequency of 21%. This finding is in contrast with the previously reported data, which showed it as the fourth most common mutation in India.\[10\] Moreover, this mutation was not among the top three mutations in several other parts of the country, viz.

Table 2: Frequency of β-thalassemia mutations detected

<table>
<thead>
<tr>
<th>Mutation</th>
<th>No. of patients detected with mutation</th>
<th>Frequency (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>IVS 1-5 (G-C)</td>
<td>22</td>
<td>46</td>
</tr>
<tr>
<td>Fr 8/9 (+G)</td>
<td>10</td>
<td>21</td>
</tr>
<tr>
<td>IVS 1-1 (G-T)</td>
<td>6</td>
<td>12</td>
</tr>
<tr>
<td>Fr 41/42 (−TTCT)</td>
<td>2</td>
<td>4</td>
</tr>
<tr>
<td>Cd 26 HBE</td>
<td>0</td>
<td>0</td>
</tr>
</tbody>
</table>

HBE: ε globin gene; TTCT: Thymine, Cytosine; IVS: Intervening Sequences
Delhi,[20] Chandigarh,[21] Gujarat,[7] and Western India.[22] However, in Eastern India, this mutation ranked as the third most common, HBE being the 2nd most common mutation,[15] while in South India, the same mutation has a very low frequency of occurrence. Hence, this mutation can be considered unique to Western U.P. High frequency of IVS 1-5 and Fr 8/9 mutations in our study suggests that these may be the oldest β-thalassemia mutations in Western U.P.

The 3rd most common mutation in Western UP was IVS 1-1 (12%). This conforms with the data previously reported as the third most common mutation in India.[1] However, this mutation was not detected in Southern and Eastern India.[14,15] The least common mutation detected in the present study was Cd 41/42 (4%); the same was reported from Delhi[20] but is in contrast with the data reported from Chandigarh.[21]

An important finding of the present study is that the distribution of Fr 8/9 mutation is high in comparison with the other regions of the country. To the best of our knowledge, this mutation as the 2nd most prevalent mutation has never been reported previously from India. Hence, this mutation can be considered unique to Western U.P as HBE is unique to Eastern India.

β-thalassemia has been reported as probably the most common inherited hemoglobin disorder in the Indian subcontinent.[23] Since, no viable forms of treatment are available, the best course is prevention through prenatal diagnosis. Premarital screening in Indian population is still considered controversial, while the same is practiced successfully in Iran and Turkey.[24,25] Although, the frequency of various β-thalassemia mutations in major cities in different states of India have already been reported, yet the common mutations in various rural areas of the country still need to be explored. Moreover, no attempt of molecular characterization of β-thalassemia has been carried out in Western U.P region. As each population or sub population has its own unique spectrum of mutations, in the present study, the prevalence of β-thalassemia mutations was investigated in-patients of this area. Through ARMS technique, we were able to analyze mutations in transfusion dependent β-thalassemia patients.

Conclusion

The people in U.P exhibit a genetic heterogeneity that is unparalleled in other regions of India. As only a few prevalent HBB mutations underlie the majority of patients with β-thalassemia in this area, a relatively cost-effective dedicated carrier screening method could be implemented. Furthermore, prenatal diagnosis and other preventive approaches may be the other important strategies in this regard.

Acknowledgment

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