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REVIEW



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Dermatoscopic Features of Non-melanocytic Skin Tumours

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Abstract: Dermatoscopy is a cheap and non-invasive diagnostic technique that improves the diagnostic accuracy of non-pigmented benign and malignant skin tumours. Dermatologist should be aware of dermatoscopic features of non-melanocytic skin tumours to reach the correct diagnosis.

Keywords: dermatoscopy, non-melanocytic skin tumours

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Introduction

Dermatoscopy (also known as dermoscopy, incident light microscopy, epiluminescence microscopy and skin-surface microscopy) is an inexpensive, in vivo and non-invasive technique that permits the visualization of morphologic features that are not visible to the naked eye.¹ Although a 10-fold magnification is sufficient for the assessment of the suspicious skin lesions, magnifications in various dermatoscopy instruments range from 10x to 100x. Dermatoscopy is widely used currently for the diagnosis of pigmented and non-pigmented skin lesions.

There is conflicting data in the literature regarding the history of dermatoscopy. Johan Christophorus Kolhaus investigated small vessels in the nail bed using a microscope in 1636. In 1893, Unna used oil immersion to make the skin more transparent and examined lupus vulgaris lesions.² The German dermatologist, Johann Saphier published four reports on his method adding a built-in light source to the dermatoscope in 1920 and 1921. He was the first to use the term "dermatoscopy". In the 1950s, Goldman coined the term "dermoscopy".³

Dermatoscopy helps in the diagnosis of many pigmented skin lesions such as seborrheic keratosis, pigmented basal cell carcinoma, haemangioma, blue nevus, atypical nevus, and cutaneous melanoma. It is 10% to 27% more sensitive than clinical criteria of ABCD (asymmetry, border regularity, colour distribution, and diameter) in the early diagnosis of cutaneous melanoma.^{4,5} Dermatoscopy of melanocytic lesions increases the presurgical accuracy rate of clinical diagnosis from 50% to 85%.^{6,7}

The accuracy of clinical diagnosis of pigmented Spitz nevi improved from 56% to 93% by using dermatoscopy.^{8,9} Demirtasoglu et al found that dermoscopy raised the rate of diagnostic accuracy for pigmented basal cell carcinoma from 60% to 90% and reported that dermatoscopy is a valuable diagnostic tool in the diagnosis of pigmented basal cell carcinoma.¹⁰ Use of the dermatoscopic methods by experienced physicians increases clinical diagnostic accuracy for haemangioma and angiokeratoma by 87% to 100%.^{11,12}

Basal Cell Carcinoma

Basal cell carcinoma (BCC) is the most common type of skin cancer in humans.¹³ It originates from the basal layer of the epidermis. Non-pigmented basal cell carcinomas

are much more common than pigmented basal cell carcinoma.¹⁴ In the dermatological examination, nonpigmented BCCs can be easily distinguished from any other skin lesion by their asymmetrical arborising vessels, pink colour, and focal ulceration (Fig. 1).¹⁵ White regression areas may be seen.¹⁶

Pigmented BCCs sometimes can be difficult to distinguish clinically from melanoma. Dermatoscopy has been proven to be useful diagnostic tool to distinguish pigmented BCC from other pigmented lesions.^{16–19} Menzies et al proposed a simple dermatoscopic method for diagnosing pigmented BCCs. This method has a sensitivity of 93% and a specificity



Figure 1. Dermatoscopy of non-pigmented BCC—pink colour, absence of pigment network, and arborising vessels.

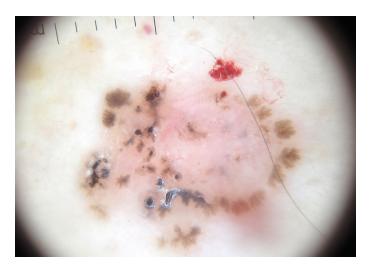


Figure 2. Dermatoscopy of pigmented BCC—structureless areas at the lesion periphery, leaf-like structures, absence of pigmented network, blue-grey globules.



of 89%. In this diagnostic method, a pigmented BCC to be diagnosed must have the negative feature (absence of pigment network) and at least one of the positive features (Table 1).¹⁸

Seborrheic Keratosis

Seborrheic keratosis (SK) is a common benign skin tumour seen mostly among the elderly population.^{20,21} Although diagnosis of SK is generally a clinical diagnosis, sometimes the differentiation between SK and cutaneous melanoma may be difficult in the clinical aspect. Braun et al reported the frequencies of the dermatoscopic structures in SK in a study with 203 patients.²² Although the classical dermatoscopic criteria of SK that includes multiple milia-like cysts and comedo-like openings had a high prevalence, additional structures such as hairpin blood vessels, fissures, sulci and gyri improved the diagnostic accuracy (Figs. 3–5).^{22,23} The dermatoscopic features of SK are easily distinguishable but nonspecific (Table 2).^{22–24}

Table 1. Dermatoscopic features of pigmented basal cellcarcinoma (Adopted).16,18

Negative feature: Absence of pigment network + at least one of the following positive features

Linear and arborising telangiectasia

Leaf-like or structureless areas on the periphery of the lesion

Multiple blue-grey globules Large blue-grey ovoid nests Focal ulceration Spoke wheel areas

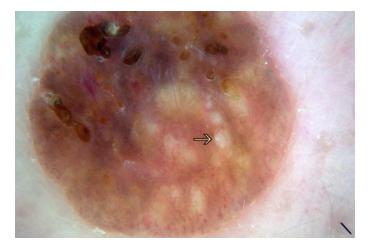






Figure 4. Dermatoscopy of seborrheic keratosis—hyperkeratosis with fissures and ridges.



Figure 5. Dermatoscopy of seborrheic keratosis—cerebriform appearance (sulci and gyri).

Table 2. Dermatoscopic features of seborrheic keratosis.22-24

Multiple milia-like cysts Pseudofollicular (comedo-like) openings Hyperkeratosis/fissures/ridges Light brown finger-like structures Hairpin blood vessels Cerebriform appearance (sulci and gyri)

Actinic Keratosis

Actinic (solar) keratosis (AK) is a direct precursor of squamous cell carcinoma (SCC) and caused by chronic exposure of UV radiation of sunlight that induces



Table 3. Dermatoscopic criteria of facial actinic keratosis.^{30–32}

Pink/red pseudonetwork and erythema surrounding the hair follicles White to yellow surface scale Linear or wavy vessels surrounding the hair follicles

Hair follicle openings filled with yellowish keratotic plugs

abnormal proliferation of epidermal keratinocytes.^{25,26} AK can be pigmented or non-pigmented. Facial AK is a differential diagnosis of cutaneous melanoma (lentigo maligna) since pigmented facial AK may have a brokenup pseudonetwork.^{25–29} Pseudonetwork can be observed in dermatoscopic examination of certain benign pigmented facial lesions such as AK, ephelide, and junctional nevus. Zaluadek et al observed four essential dermatoscopic features in facial AK and defined the combination of these features as "strawberry" pattern (Table 3) (Fig. 6).³⁰

Sebaceous Hyperplasia

Sebaceous hyperplasia is a benign proliferation of sebaceous lobules around the follicular infundibulum.^{33,34} Yellow nodules surrounding a central follicular opening can be seen in dermatoscopic examination (Fig. 7). Sebaceous hyperplasia must be differentiated from small non–pigmented BCC. Dermatoscopic examination of sebaceous hyperplasia can reveal vessels that extend to the centre of the lesion but they are never arborising.¹

Dermatofibroma

Dermatofibroma also known as fibrous histiocytoma is a common benign fibrohistiocytic mesenchymal

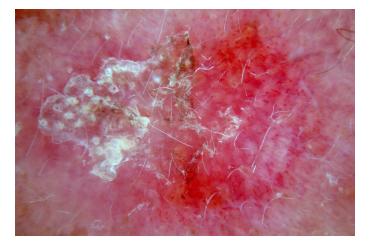


Figure 6. Dermatoscopy of actinic keratosis—white surface scale, ery-thema, pseudonetwork.

growth of the skin. The aetiology of dermatofibroma remains unclear.^{35,36} Dermatofibromas clinically exhibit "dimple sign" with the lateral depression in the overlying skin.^{37,38} Since dermatofibromas may mimic other skin tumours including melanoma the definition of their dermatoscopic features is crucial (Fig. 8) (Table 4). In a recent study of 412 dermatofibromas (from 292 patients) 10 different dermatoscopic patterns were observed. The most common dermatoscopic pattern seen in the study group was central white patch and peripheral pigment network (34.7%).³²

Squamous Cell Carcinoma

The dermatoscopic features of SCC are a non-specific pattern with scales and grouped glomerular blood vessels surrounded by a whitish halo.^{15,39,40} A scaly surface, brown globules and glomerular vessels can be seen in the dermatoscopic examination of pigmented Bowen's disease.

Vascular Lesions

Dermoscopy improves the diagnostic accuracy in the clinical evaluation of pigmented skin lesions, but it is also useful for the assessment of vascular lesions such as haemangioma, solitary angiokeratoma, and pyogenic granuloma.^{1,17} The most typical dermatoscopic features of the vascular lesions are red, blue or black lacunae (Fig. 9) and red-bluish or red-black homogenous areas. Dermatoscopic features of pyogenic granuloma were first studied by Zaballos et al (Table 5).⁴¹



Figure 7. Dermatoscopy of sebaceous hyperplasia—central follicular opening and surrounding yellow lobule.





Figure 8. Dermatoscopy of a typical dermatofibroma—Central scar-like patch and peripheral delicate network.



Figure 10. Dermatoscopy of pyogenic granuloma—red lagoons, appearance of white collarette and white "rail lines" that intersect the lesion.

 Table 4. Dermatoscopic features of dermatofibroma.^{31,32}

Peripheral pigment network Central white scar-like patch Different vascular structures White network Absence of melanocytic features



Figure 9. Dermatoscopy of haemangioma—red homogeneous area.

Table 5. Dermatoscopic features of pyogenic granuloma.

Reddish homogenous areas

White collarette

Ulceration

White rail lines intersecting the lesion

Conclusion

Dermatoscopy improves the diagnostic accuracy in melanocytic and non-melanocytic skin lesions. Thus, every dermatologist should acquire more in-depth knowledge relating to the dermatoscopic features and patterns of the benign and malignant skin lesions.

Disclosures

This manuscript has been read and approved by the author. This paper is unique and is not under consideration by any other publication and has not been published elsewhere. The author reports no conflicts of interest.

References

- Zalaudek I, Argenziano G, Di Stefani A, Ferrara G, Marghoob AA, Hofmann-Wellenhof R, et al. Dermoscopy in general dermatology. *Dermatology*. 2006; 212:7–18.
- Paschoal FM. Early diagnosis of melanoma by surface microscopy (dermatoscopy). Sao Paulo Med J. 1996;114:1220–1.
- Friedman RJ, Rigel DS, Silverman MK, Kopf AW, Vossaert KA. Malignant melanoma in the 1990s: the continued importance of early detection and the role of physician examination and self-examination of the skin. *CA Cancer J Clin.* 1991;41:201–26.
- Piccolo D, Smolle J, Argenziano G, Wolf IH, Braun R, Cerroni L, et al. Teledermoscopy—results of a multicentre study on 43 pigmented skin lesions. *J Telemed Telecare*. 2000;6:132–7.
- Soyer HP, Kenet RO, Wolf IH, Kenet BJ, Cerroni L. Clinicopathological correlation of pigmented skin lesions using dermoscopy. *Eur J Dermatol.* 2000;10:22–8.
- Nilles M, Boedeker RH, Schill WB. Surface microscopy of naevi and melanomas—clues to melanoma. *Br J Dermatol*. 1994;130:349–55.
- Soyer HP, Smolle J, Hodl S, Pachernegg H, Kerl H. Surface microscopy. A new approach to the diagnosis of cutaneous pigmented tumors. *Am J Dermatopathol.* 1989;11:1–10.
- Steiner A, Pehamberger H, Binder M, Wolff K. Pigmented Spitz nevi: improvement of the diagnostic accuracy by epiluminescence microscopy. *J Am Acad Dermatol.* 1992;27:697–701.



- Steiner A, Pehamberger H, Wolff K. Improvement of the diagnostic accuracy in pigmented skin lesions by epiluminescent light microscopy. *Anticancer Res.* 1987;7:433–4.
- Demirtasoglu M, Ilknur T, Lebe B, Kusku E, Akarsu S, Ozkan S. Evaluation of dermoscopic and histopathologic features and their correlations in pigmented basal cell carcinomas. *J Eur Acad Dermatol Venereol*. 2006;20: 916–20.
- Steiner A, Pehamberger H, Wolff K. In vivo epiluminescence microscopy of pigmented skin lesions. II. Diagnosis of small pigmented skin lesions and early detection of malignant melanoma. *J Am Acad Dermatol.* 1987;17:584–91.
- Pehamberger H, Steiner A, Wolff K. In vivo epiluminescence microscopy of pigmented skin lesions. I. Pattern analysis of pigmented skin lesions. J Am Acad Dermatol. 1987;17:571–83.
- Wong CS, Strange RC, Lear JT. Basal cell carcinoma. *BMJ*. 2003;327: 794–8.
- 14. Brooke RC. Basal cell carcinoma. Clin Med. 2005;5:551-4.
- Felder S, Rabinovitz H, Oliviero M, Kopf A. Dermoscopic differentiation of a superficial basal cell carcinoma and squamous cell carcinoma in situ. *Dermatol Surg.* 2006;32:423–5.
- Menzies SW. Dermoscopy of pigmented basal cell carcinoma. *Clin Dermatol.* 2002;20:268–9.
- 17. Kreusch JF. Vascular patterns in skin tumors. *Clin Dermatol*. 2002;20: 248–54.
- Menzies SW, Westerhoff K, Rabinovitz H, Kopf AW, McCarthy WH, Katz B. Surface microscopy of pigmented basal cell carcinoma. *Arch Dermatol.* 2000;136:1012–6.
- Terstappen K, Larko O, Wennberg AM. Pigmented basal cell carcinoma comparing the diagnostic methods of SIAscopy and dermoscopy. *Acta Derm Venereol*. 2007;87:238–42.
- Kettler AH, Goldberg LH. Seborrheic keratoses. Am Fam Physician. 1986;34: 147–52.
- Cashmore RW, Perry HO. Differentiating seborrheic keratosis from skin neoplasm. *Geriatrics*. 1985;40:69–71, 4–5.
- Braun RP, Rabinovitz HS, Krischer J, Kreusch J, Oliviero M, Naldi L, et al. Dermoscopy of pigmented seborrheic keratosis: a morphological study. *Arch Dermatol.* 2002;138:1556–60.
- Braun RP, Rabinovitz H, Oliviero M, Kopf AW, Saurat JH. Dermoscopic diagnosis of seborrheic keratosis. *Clin Dermatol.* 2002;20:270–2.
- Sahin MT, Ozturkcan S, Ermertcan AT, Gunes AT. A comparison of dermoscopic features among lentigo senilis/initial seborrheic keratosis, seborrheic keratosis, lentigo maligna and lentigo maligna melanoma on the face. *J Dermatol.* 2004;31:884–9.
- Callen JP, Bickers DR, Moy RL. Actinic keratoses. J Am Acad Dermatol. 1997;36:650–3.
- Rossi R, Mori M, Lotti T. Actinic keratosis. Int J Dermatol. 2007;46: 895–904.
- Schwartz RA, Bridges TM, Butani AK, Ehrlich A. Actinic keratosis: an occupational and environmental disorder. *J Eur Acad Dermatol Venereol*. 2008.
- Piaserico S, Belloni Fortina A, Rigotti P, Rossi B, Baldan N, Alaibac M, et al. Topical photodynamic therapy of actinic keratosis in renal transplant recipients. *Transplant Proc.* 2007;39:1847–50.

- Dinehart SM. The treatment of actinic keratoses. J Am Acad Dermatol. 2000;42:25–8.
- Zalaudek I, Giacomel J, Argenziano G, Hofmann-Wellenhof R, Micantonio T, Di Stefani A, et al. Dermoscopy of facial nonpigmented actinic keratosis. *Br J Dermatol*. 2006;155:951–6.
- Arpaia N, Cassano N, Vena GA. Dermoscopic patterns of dermatofibroma. Dermatol Surg. 2005;31:1336–9.
- Zaballos P, Puig S, Llambrich A, Malvehy J. Dermoscopy of dermatofibromas: a prospective morphological study of 412 cases. *Arch Dermatol.* 2008;144: 75–83.
- Boonchai W, Leenutaphong V. Familial presenile sebaceous gland hyperplasia. J Am Acad Dermatol. 1997;36:120–2.
- Zouboulis CC, Boschnakow A. Chronological ageing and photoageing of the human sebaceous gland. *Clin Exp Dermatol.* 2001;26:600–7.
- Cerio R, Spaull J, Jones EW. Histiocytoma cutis: a tumour of dermal dendrocytes (dermal dendrocytoma). Br J Dermatol. 1989;120:197–206.
- Hui P, Glusac EJ, Sinard JH, Perkins AS. Clonal analysis of cutaneous fibrous histiocytoma (dermatofibroma). J Cutan Pathol. 2002;29:385–9.
- Zelger B. Pigmented atypical fibroxanthoma, a dermatofibroma variant? *Am J Dermatopathol*. 2004;26:84–6; author reply 6–7.
- Zelger B, Zelger BG, Burgdorf WH. Dermatofibroma-a critical evaluation. Int J Surg Pathol. 2004;12:333–44.
- Bugatti L, Filosa G, De Angelis R. The specific dermoscopical criteria of Bowen's disease. J Eur Acad Dermatol Venereol. 2007;21:700–1.
- Cabrijan L, Lipozencic J, Batinac T, Lenkovic M, Gruber F, Stanic Zgombic Z. Correlation between clinical-dermatoscopic and histopathologic diagnosis of skin tumors in our patients. *Coll Antropol.* 2008;32 Suppl 2:195–7.
- Zaballos P, Llambrich A, Cuellar F, Puig S, Malvehy J. Dermoscopic findings in pyogenic granuloma. *Br J Dermatol.* 2006;154:1108–11.

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