REVIEW

The role of TGF- β in bone metastasis: novel therapeutic perspectives

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The skeleton is a preferred site for cancer metastasis. These bone metastases cause dysregulated bone remodeling and the associated morbidity of fractures, pain, hypercalcemia and catastrophic nerve compression syndromes. Transforming growth factor- β (TGF- β) is stored in mineralized bone matrix, and released and activated by osteoclastic bone resorption. Once activated, TGF- β stimulates nearby metastatic tumor cells within the bone microenvironment to secrete factors that further drive osteolytic destruction of the bone. Therefore, TGF- β and its signaling constitute a critical component driving the feed-forward vicious cycle of cancer growth in bone. Moreover, additional pro-tumorigenic activities attributed to TGF- β include activation of epithelial-to-mesenchymal transition, increased tumor cell invasion, enhanced angiogenesis and various immunomodulatory properties. Blocking the TGF- β signaling pathway to interrupt this vicious cycle and manipulate the bone microenvironment offers a promising area for therapeutic intervention to decrease skeletal metastasis and normalize bone homeostatic mechanisms. In this review, preclinical and clinical data are evaluated for the potential use of TGF- β pathway inhibitors in clinical practice to treat bone metastases and its associated comorbidities.

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Introduction

The skeleton is a preferred site for cancer metastasis. Up to 80% of patients with advanced breast or prostate cancer will develop bone metastases as will 30-40% of patients with lung, renal or thyroid cancer.^{1,2} This devastating complication of cancer causes bone pain, fractures, hypercalcemia and nerve compression syndromes severely diminishing the quality of life.^{3,4} Bone metastases are classified as osteolytic or osteoblastic, based on radiographic appearance. Solid tumors such as breast, lung and renal cancer are typically associated with osteolytic lesions as is the hematologic malignancy multiple myeloma while prostate cancer is associated with osteoblastic bone metastases. Despite these classifications, most patients with solid tumor metastases to bone have components of both bone destruction and new bone formation. Multiple myeloma differs in that it is almost always associated with profound bone destruction and suppressed bone formation.

Perhaps the most devastating consequence is that once cancer metastasizes to bone, it is incurable. Current standard of care to treat bone metastases include antiresorptive therapy to decrease skeletal morbidity; clearly beneficial, but without regression of disease or cure.^{3,4} Patients with cancer metastases to bone,

particularly those with breast and prostate cancer, can survive for many years, during which they will suffer significant morbidity. Thus, better treatments are needed to achieve the long-term goal of preventing or curing bone metastases.

The bone microenvironment is unique and provides a special milieu that metastatic cancer cells can colonize. The mineralized bone matrix is embedded with abundant growth factors and cytokines during the bone formation phase, such as transforming growth factor- β (TGF- β), activins and insulin-like growth factors, which are released and activated upon tumor-induced osteoclastic bone resorption.⁵ High local levels of active TGF- β cause increased invasion, chemotaxis, angiogenesis and immunomodulation. In addition, TGF- β stimulates tumor production of osteolytic factors that further stimulate bone resorption (**Figure 1**).^{6,7} This categorizes TGF- β as a crucial factor responsible for driving the feed-forward vicious cycle of tumor growth in bone. Therefore blocking TGF- β release, its production and/or signaling is a promising strategy to treat bone metastasis.

$TGF-\beta$

TGF- β is a ubiquitously expressed, pluripotent cytokine that controls tissue homeostasis by regulating cellular processes such

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as apoptosis, proliferation and differentiation.⁸ Almost all cells secrete TGF- β and express TGF- β receptors. Therefore, it is not surprising that dysregulation of TGF- β actions has been associated with many disorders, including impaired wound healing, chronic fibrosis, cardiovascular diseases and cancer.^{9,10} The pluripotent nature of TGF- β and its ubiquitous expression provides both opportunities and challenges to neutralize its effects.

TGF- β structure and signaling. Three highly homologous isoforms of TGF- β have been described in humans, TGF- β_1 , TGF- β_2 and TGF- β_3 . The signaling of these isoforms is comparable but the messenger RNA (mRNA) expression levels and/or protein presence differ across various tissues.¹¹ Generally, all of the active TGF- β isoforms bind with high affinity and selectivity to the membrane-spanning serine/threonine kinase receptor TGF- β receptor type II (T β RII), which then recruits and activates TGF- β receptor type I (T β RI, a.k.a. ALK5) (Figure 2).¹² The activated T β RI/ALK5 complex can then phosphorylate the receptor-associated Smads (R-Smads), Smad2 and Smad3.¹³ These activated R-Smads form a stable heterodimeric complex with the common mediator Smad, Smad4, and translocate to the nucleus.^{12,13} Ultimately, the transcriptional outcome on their target genes depends on a host of transcriptional partners (co-activators or corepressors) that Smads interact with. This repertoire of Smad coregulatory proteins is variable, based on the cell type and physiological response being regulated by TGF- β in each cellular niche.

Non-canonical TGF- β signaling can also activate non-Smad signaling pathways including extracellular signal-regulated kinases (ERK-1, ERK-2 and p38), c-Jun amino-terminal kinase

and mitogen-activated protein kinases (MAPKs) in various cell types¹⁴ (**Figure 2**).

TGF- β in Bone Homeostasis

Adult bone is continuously remodeled by the coordinated and balanced activities of bone-resorbing osteoclasts and bone-forming osteoblasts. TGF- β_1 is one of the most abundant growth factors found in bone matrix, 15,16 and the effects of TGF- β on osteoblast, osteoclasts and bone remodeling are both temporal-dependent, spacially and context-dependent. 17

Significant evidence has demonstrated that TGF- β is a key mediator in the dynamic coupling of bone resorption and bone formation.^{18–20} Osteoblasts secrete predominantly TGF- β_1 , where it is embedded as a latent form into the mineralized bone matrix.¹⁷ Osteoclastic bone degradation releases and activates TGF- β , which will result in recruitment of osteoblast precursors to sites of bone resorption.^{18–20} The exposed bone mineral matrix and release of osteotropic factors, such as bone morphogenetic proteins (BMPs), insulin-like growth factor-I and -II, and platelet-derived growth factor, may then promote differentiation of the osteoblast precursor to osteoblasts.²¹ In later phases of osteoblast differentiation, TGF- β has been shown to block osteoblast differentiation, impede bone mineralization^{22,23} and induce osteoblast survival during transdifferentiation into osteocytes.²⁴

Although TGF- β appears to have complex biphasic effects in osteoclastogenesis and bone resorption *in vitro*,^{25,26} studies with genetically modified mice showed that increased levels of TGF- β in bone microenvironment promoted osteoclastogenesis

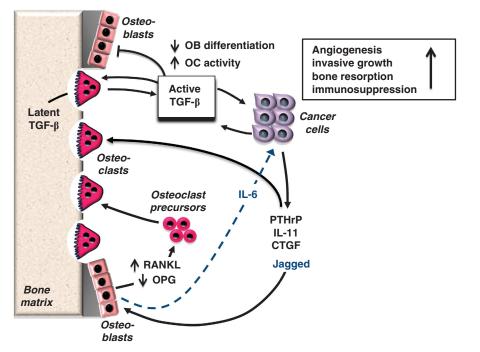


Figure 1 TGF- β in osteolytic bone metastasis. The skeleton is a preferred site for cancer metastasis. In these skeletal metastases, TGF- β is released by osteoclasts from the bone matrix and acts on cancer cells to stimulate the production of osteolytic factors, such as PTHrP, connective tissue growth factor (CTGF) and IL-6 and -11. These factors increase the RANKL/OPG expression ratio in bone stromal cells such as osteoblasts, resulting in osteoclastogenesis. Moreover, active TGF- β stimulates Jagged1 expression in cancer cells, which in turn stimulates Notch signaling in osteoclasts and osteoblasts after direct contact. This results in increased osteoclastogenesis and production of the cytokine IL-6 by osteoblast (blue dotted line), acting as a potent inducer of proliferation of tumor cells. TGF- β itself has also direct effects on bone cells by stimulating osteoclast activity and inhibiting osteoblast differentiation. In aggregate, the effects of TGF- β perpetuate the feed-forward cycle to increase tumor growth in bone.

In support of these data, it has been shown recently that systemic TGF- β inhibition—either by using the orally active T β RI/ALK5 kinase inhibitor, SD-208,²⁹ or the highly selective pan-neutralizing TGF- β antibody, 1D11—resulted in increased trabecular bone volume. This was accompanied by enhanced osteoblast differentiation and subsequent bone formation as well as reduced osteoclast formation and bone resorption.²⁹

TGF- β in Cancer

TGF- β has an essential role in maintaining physiological homeostasis in many tissues through its ability to induce cell cycle arrest, differentiation and apoptosis, thereby preventing uncontrolled proliferation of epithelial, endothelial and hematopoietic cells within the various cellular microenvironment niche(s).^{30,31} However, many cancers often become refractory to this growth inhibition either due to genetic loss of TGF- β signaling components or, more commonly, because of downstream perturbation by other integrated signaling pathways.³² During this time, the protumorigenic actions of TGF- β may prevail, including immunomodulatory properties, induction of angiogenesis and/or promotion of the epithelial-to-mesenchymal transition (EMT) facilitating cancer migration and invasion (reviewed in refs^{33–35}). When TGF- β suppressive effects are lost, TGF- β overproduction and signaling are also commonly observed in many solid tumors.^{36,37}

TGF- β in Bone Metastasis

Metastasis to bone is a multistep process of events.^{3,4,38} First, cancer cells must detach from the primary tumor, enter into the systemic lymphatic or vascular circulation (intravasation), evade the immune system, home and arrest in the bone marrow capillaries or sinusoids, extravasate into the bone marrow and form a micrometastasis. Eventually, some micrometastases may grow into overt bone metastatic lesions.

Numerous studies have shown the importance of the TGF- β signaling pathway for the development of osteolytic bone metastases (**Figure 1**). Yin *et al.*⁷ were the first to demonstrate that blocking TGF- β signaling by stably expressing a dominant-negative T β RII (DNT β RII) in MDA-MB-231 breast cancer cells inhibited TGF- β -induced expression of parathyroid hormone-related protein (PTHrP) production in tumor cells and suppressed the formation of osteolytic bone metastases in a mouse model. *GLI2* appeared to be a TGF- β /Smad target gene^{39,40} that appears, independent of the canonical Hedgehog, to be one of the important mediators in the upregulation of PTHrP.^{41,42} In addition, PTHrP is also increased via Smad-independent pathways (p38 MAPK and to a lesser extend TGF- β activated kinase).^{43,44}

PTHrP has a major role in the development of the osteolytic features of bone metastatic lesions,⁴⁵ and is considered to be responsible for the humoral hypercalcemia of malignancy syndrome (reviewed in refs ^{46,47}). PTHrP stimulates osteo-

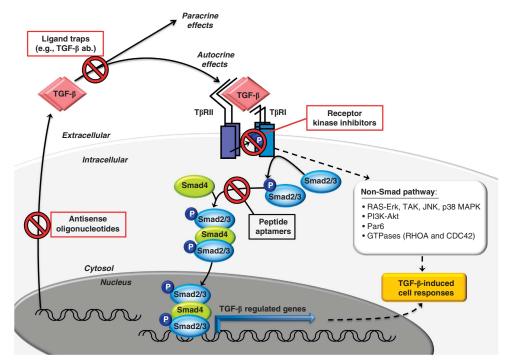


Figure 2 The TGF- β signaling pathway and several therapeutic strategies to intervene. Upon binding of active TGF- β to the TGF- β type I (ALK5) and type II (T β RII) receptor, ALK5 is phosphorylated and activated by T β RII. ALK5, in turn, phosphorylates and activates the R-Smads, Smad2 and Smad3, which form a complex with Smad4. This complex translocates to the nucleus, binds to DNA-binding transcription factors, co-activators and co-repressors regulating TGF- β /Smad target gene expression. In addition, TGF- β is also known to regulate non-Smad pathways, including ERK, p38 MAPK, Jun N-terminal kinase (JNK), PI3K-Akt and small GTPases. Several strategies to inhibit the TGF- β signaling have been preclinically tested, and are currently in clinical trials (see red boxes). Three of these classes of TGF- β inhibitors are tested in the clinic and include the following: ligand traps, including monoclonal-neutralizing TGF- β antibodies that bind and neutralize TGF- β ligand, preventing TGF- β ligand–receptor interactions; receptor kinase inhibitors, which are small molecules that inhibit T β RI (and T β RII) kinase activity, preventing the activation TGF- β R-Smads; and antisense oligonucleotides (ASOs) which inhibit TGF- β expression at transcriptional/translational level, preventing the production and release of TGF- β in microenvironment. Peptide aptamers are a class of TGF- β inhibitors that are not yet tested *in vivo*. They can specifically bind to R-Smads of interest and prevent complex formation with Smad4 offering targeting of specific TGF- β responses.

clast activation by inducing receptor activator of nuclear factor- κ B ligand and downregulating osteoprotegerin in cells of the osteoblast lineage.⁴⁸ In breast cancer, 90% of metastases in bone were found to express PTHrP, compared with only 17% at non-bone sites and 60% of the primary tumors.^{49,50} A large prospective study demonstrated that PTHrP expression in primary breast cancer was significantly associated with fewer (bone) metastases.⁵¹ Therefore, the most likely explanation for the observed increased PTHrP expression in breast cancer bone metastases.^{49,50} is that factors in the bone microenvironment, such as TGF- β , induces cancer cells to express PTHrP rather than the possibility that primary breast cancer cells that express PTHrP have a greater propensity to metastasize to bone.

Transcriptional profiling of various selected subpopulations of human breast cancer cells in a mouse model of MDA-MB-231 bone metastases further illustrated the complexity of bone metastases pathophysiology. Using serial passaging of MDA-MB-231 cells, Kang et al. isolated clones that cause more aggressive bone metastases when compared with its parental line. Microarray analysis identified a number of genes that were selectively upregulated in these aggressive bone metastatic clones. Many of these proteins, including interleukin (IL)-11, CXCR4, connective tissue growth factor and matrix metalloproteinase (MMP)-1, have well-described effects on bone cells. IL-11 stimulates bone resorption by increasing osteoblast production of receptor activator of nuclear factor- κ B ligand.⁶ CXCR4 is a chemokine receptor that binds to stromal-derived factor-1 produced by osteoblasts, and its expression promotes homing of cancer cells to bone. Connective tissue growth factor stimulates osteoblast proliferation as well as angiogenesis, and MMP-1 promotes bone metastasis by activating an EGFR-dependent paracrine signaling cascade suppressing the expression of osteoprotegerin by osteoblasts.^{52,53} When expressed and cosignaling is allowed to occur, these proteins act cooperatively to cause osteolytic metastasis by promoting homing to bone, angiogenesis and invasion. Among the various bone metastasis genes identified, Kang et al.⁶ showed that two of these genes, IL-11 and connective tissue growth factor, were directly regulated by TGF- β via the canonical TGF-β/Smad pathway in metastatic cells. Other studies indicate that CXCR4 and MMP-1 are also regulated by TGF-B.54,55 Using the same mouse model, knockdown of the TGF-β signaling molecule Smad4 inhibited the formation and growth of bone metastases.^{56,57} Recently, it was shown that active TGF-ß stimulates Jagged1 expression in breast cancer bone metastases, which in turn stimulates Notch signaling in osteoclasts and osteoblasts after direct contact.58 This results in increased osteoclastogenesis and the production of the cytokine IL-6 by osteoblast, acting as a potent inducer of proliferation of tumor cells. Collectively, these effects of TGF- β perpetuate the feed-forward cycle to increase tumor growth in bone in a complex and multifaceted way.

These preclinical data also extend to human studies. Plasma TGF- β_1 levels were shown to be elevated in more than half of the 49 (including 23 breast and 15 prostate cancer patients) bone metastasis patients and positively correlated with TGF- β signaling-related markers, including PTHrP and IL-10.⁵⁹ Bone metastases samples from breast cancer patients also displayed enhanced Smad-dependent TGF- β signaling as there is accumulation of phosphorylated Smad2 in the nucleus of tumor cells and cells of the surrounding stroma.^{57, 60} In mouse models of bone

metastases, using live imaging of tumor cells by thymidine kinase activity or dual-bioluminescence, it was also demonstrated that TGF- β signaling is activated in bone metastases, but not in metastases to adrenal glands.^{5,57,61} Furthermore, treatment with a T β RI kinase inhibitor could effectively reduce TGF- β signaling in bone metastasis as could the bisphosphonate, pamidronate, a potent inhibitor of bone resorption.⁵ These data underscore the central role of TGF- β in the pathogenesis of bone metastases as well as the role of bone resorption as the driving source to provide active TGF- β in the bone microenvironment. Furthermore, bone sialoprotein and osteopontin, two secreted glycoproteins regulated by TGF- β , have important roles in bone turnover and were found to be highly expressed in malignant prostate and breast cancer tissue, and correlated with tumor grade. In these patients, serum and mRNA levels of bone sialoprotein and osteopontin were identified to be prognostic indicators for bone metastases.^{62–65}

Hypoxia is observed in most solid tumors and is caused by reduced or inadequate oxygen supply.⁶⁶ Owing to already low oxygen levels (1–7% O₂) in the bone marrow microenvironment, hypoxia and increased expression of hypoxia-inducible factor 1 α are particularly prevalent in bone metastases.^{54,67} Hypoxia also stimulates the expression of *CXCR4* and *DUSP1*,⁶⁸ two genes previously recognized among 11 genes that are most upregulated in MDA-MB-231 bone metastatic clones.⁶ In addition, TGF- β stabilizes hypoxia-inducible factor 1 α by inhibiting its degradation,⁶⁹ and additive responses in the induction of vascular endothelial growth factor and CXCR4 are observed *in vitro*^{54,69} and *in vivo*.⁵⁴ Therefore, the significant interplay of TGF- β signaling and hypoxia may be a key cooperative determinant for various mechanisms of bone metastasis.

TGF- β as Therapeutic Target

As a result of its wide variety of effects, blockade of TGF- β or its signaling has provided therapeutic opportunities for the treatment of various different diseases, including fibrotic disease and cancer.^{70–72} For bone metastatic disease, most compounds still remain at the preclinical stages (**Table 1**). As multiple agents under development for the various disease indications other than bone metastatic disease have been described, this might be indicative of the potential tolerability and potential success for safe TGF- β signaling antagonism (**Table 2**). A few of these novel chemical entities will be discussed here in detail while others have been extensively reviewed elsewhere.^{72,73}

Neutralizing antibodies and soluble decoy receptor proteins. One strategy to reduce signaling by the excessive amounts of TGF- β in the (bone) microenvironment is to physically neutralize or trap TGF- β ligand using soluble decoy receptors encompassing the ectodomains from either T β RII or T β RIII/betaglycan protein or via neutralizing TGF- β antibodies (Figure 2). The pan-neutralizing mouse monoclonal antibody 1D11 binds and reduces biological activity of all three TGF- β isoforms and has demonstrated therapeutic potential in both syngeneic and xenograft mouse tumor models, including preclinical bone metastasis models^{74,75} (Table 1).

For use in patients, a fully humanized TGF- β monoclonalneutralizing antibody, GC-1008, is currently being developed by Genzyme (Cambridge, MA, USA)⁷² (**Table 2**). GC-1008 is a pan-neutralizing antibody directed against all three isoforms of

Table 1 Anti-TGF- β compounds successfully used in preclinical bone	
metastasis models	

Agent/target	Target	Company	References
Trap ligands Antibodies 1D11	$TGF-\beta_{1-3}$	Genzyme/ CAT	74, 75, 113
Receptor kinase	inhibitors		
Ki 26894	ALK5	Kirin Brewery	114
LY2109761	ALK5+TGFRII	Company Eli Lilly and Co.	5, 93–95
LY2157299	ALK5	Eli Lilly and Co.	96, 115
LY364947	ALK5	Eli Lilly and Co.	116, 117
SD-208	ALK5	Scios, Inc./ Johnson & Johnson	54, 90–92, 118–120
Combined vaccir 2G7+IL-2	ne/antisense TGF-β ₁₋₃ +IL-2	Genentech	121
Other molecules Halofuginone	antagonizing effec TGF-β effects	Collgard Biophar-	108
BMP7	TGF- β effects	maceuticals Stryker	102, 104

Abbreviation: TGF- β , transforming growth factor- β .

TGF-β, and completed phase I dose-escalation studies in 22 patients with renal cell carcinoma and metastatic melanoma who had failed at least one previous therapy.⁷⁶ No dose-limiting toxicities were observed in these patients. Intriguingly, among patients with malignant melanoma, one patient had a partial response, three had a mixed response and one had stable disease.⁷⁶ PF-03446962 generated by Pfizer (New York City, NY, USA), is an antibody directed against ALK1 and displays potent anti-angiogenic effects *in vitro*.⁷⁷ Currently, this antibody is in phase I clinical trials to evaluate optimal pharmacokinetic parameters for this agent in patients with advanced solid tumors (**Table 2**).

Antisense oligonucleotides Another feasible therapeutic strategy to minimize excessive levels of TGF- β in the local tumor microenvironment is to reduce TGF- β synthesis and secretion by using antisense oligonucleotides (ASOs). ASOs inhibit mRNA function and protein synthesis through modulation of splicing and/or inhibition of translation by disrupting ribosome assembly.^{78,79} Several limitations need to be considered when using ASOs, including stability, RNA-binding affinity, efficiency of delivery to the target cells and associated off-target effects. While TGF- β ASOs should, in theory, not interfere with activated TGF- β proteins released from bone, it is possible that ASO treatment could result in lower concentrations of TGF β being deposited in bone.

Antisense Pharma (Regensburg, Germany) has developed AP-12009 (Trabedersen), an ASO specific for the mRNA of human TGF- β_2 ,^{80,81} for which patients with anaplastic astrocytoma are currently being recruited for a phase III study to evaluate the efficacy and safety of intratumoral treatment with AP-12009 compared with standard chemotherapy (temozolomide) (**Table 2**). In addition, a phase I dose-escalation clinical study is currently ongoing to evaluate the safety and tolera-

bility of intravenously administration of AP-12009 in patients with advanced tumors known to overproduce TGF- β_2 , such as melanoma, pancreatic and colorectal carcinomas (**Table 2**).

Furthermore, TGF- β ASOs are also currently being tested in early-phase clinical trials to generate ASO-modified tumor vaccines with the goal of reducing the immune inhibiting activity at the vaccine sites.⁸²

Receptor kinase inhibitors. While ligand traps and ASOs limit the bioavailability of active TGF- β ligands to directly interact with its cognate receptor, TGF- β receptor kinase inhibitors are a group of small-molecule inhibitors that act via ATP-competitive inhibition of the kinase catalytic activity of T_βRI/ALK5 upon its recruitment, phosphorylation and activation by TGFβ-bound T β RII. While there may be several advantages to the development and scalability of small-molecule inhibitors including potentially optimized pharmacokinetic/pharmacodynamic properties, the potential lack of selectivity of kinase inhibitors continues to be a challenge. Currently, all known small-molecule TβRI/ALK5 inhibitors described in the literature to date, where tested, display equipotent inhibition against ALK4 kinase activity and less inhibition against ALK7.83-87 Interestingly, activin(s) have been shown to require ALK4 for their signaling, and recent animal studies using activin-inhibiting soluble extracellular domain ACVR2A-Fc fusions also display significant anti-resorptive and bone formation activities in various animal models.88,89 Whether the combined inhibition of TGF- β and activin signaling by ALK4/5 inhibitors is advantageous for the treatment of cancer remains to be determined. Of all the different classes of TGF- β antagonists, the T β RI/ALK5 kinase inhibitors are the ones most extensively tested in both syngeneic and xenograft preclinical bone metastasis models (Table 1). For example, the orally active TBRI/ALK5 kinase inhibitor SD-208 inhibited progression and development of melanoma and osteolytic breast cancer bone metastases.^{90,91} In addition, it inhibited bone metastases from osteolytic PC3 prostate cancer cells, but was unable to inhibit tumor growth in bone of LuCaP23.1 prostate cancer cells that form osteoblastic bone metastases.⁹² Therefore, TGF-β blockade may be more effective to treat osteolytic than osteoblastic disease, perhaps owing to the fact that inhibition of TGF- β may result in an additional osteoblastic response.²⁹

LY2109761 (Eli Lilly and Co., Indianapolis, IN, USA), an inhibitor of both the T β RI/ALK5 and T β RII, inhibits the formation of metastases in several mouse models, including colon,⁹³ breast,⁵ prostate⁹⁴ and pancreatic⁹⁵ cancer. While SD-208 successfully inhibited the growth of established MDA-MB-231 bone metastases,⁵⁴ LY2109761 did not,⁵ and suggest that SD-208 may be a more potent or mechanistically distinct versus LY2109761 in inhibiting TGF- β signaling *in vivo*.

LY2157299, under development by Lilly and Co, has entered phase I clinical trials to determine the safety and pharmacokinetics in patients with advanced metastatic malignancies including melanoma, colon cancer, prostate cancer, adrenal gland cancer and breast cancer who have exhausted current standard of care. Daily doses of 40 mg and 80 mg LY2157299 have currently been well tolerated and no drug-related grade 3 or 4 toxicities were observed.⁹⁶ Additional clinical studies are underway in patients with bone metastasis.

Other molecules that antagonize TGF- β . Numerous additional biologic-based molecules, that do not necessarily directly bind

Table 2 Anti-TGF- β compounds currently in clinical trials

Class (compound)	Target	Company	Phase	Disease	References
Trap ligands					
Antibodies Fresolumimab/ GC1008	$TGF\text{-}\beta_{1\text{-}3}$	Genzyme	Phase I (c) Phase I (r)	RCC+Melanoma Metastatic breast cancer	NCT00356460 NCT01401062
Antibody targeting ALK1			Phase I (c)	Advanced melanoma	NCT00923169; 76
PF-03446962	ALK1	Pfizer	Phase I (r)	Advanced solid tumors	NCT00557856; 77
Antisense oligonucleotide Trabedersen (AP12009)	s TGF-β ₂ mRNA	Antisense Pharma	Phase I (c)	Pancreatic + colorectal neoplasms, melanoma	NCT00844064; NCT00431561 NCT00761280
			Phase IIb (c) Phase III (r)	Glioma AA, glioblastoma	NC100701200
Receptor kinase inhibitors					
LY2157299	ALK5	Eli Lilly and Co.	Phase Ib/IIa (r) Phase II (r)	Glioma Hepatocullar carcinoma	NCT01220271 NCT01246986
			Phase Ib/II (r)	Pancreatic cancer	NCT01373164
Combined TGF-β AOS with Lucanix™ (Belagenpumatucel-L)	th a vaccine TGF-β ₂	NovaRx Corp	Phase II (c) Phase III (r)	NSCLC NSCLC	NCT01058785; 122 – 123 NCT006765
TGF- β_2 ASO +GMCSF expression vector	$TGF-\beta_2 + GMCSF$	MCMRC	Phase I (r)	Advanced cancer	NCT00684294

Abbreviations: AA, anaplastic astrocytoma; CRC, colorectal carcinoma; (c), study completed; MCMRC, Mary Crowley Medical Research Centre; NCT#, clinical trial identifier (www.clinicaltrial.gov); NSCLC, non-small-cell lung cancer; (r), study currently recruiting patients; RCC, renal cell carcinoma; TGF-β, transforming growth factor-β.

and inactivate TGF- β , its receptor or its signaling molecules, may still antagonize the effects of TGF- β indirectly. Here, two examples of molecules that antagonize TGF- β function are described that have been extensively tested in bone metastasis models.

BMP7 is another member of the TGF- β superfamily and signals via different type I and II receptor kinases activating the R-Smads, Smad1, -5 and -8.97,98 It has long been recognized that BMP7 can counteract TGF-β-induced EMT, and induce the opposite process, a mesenchymal-to-epithelial transition in embryonic kidney and eye development.⁹⁹⁻¹⁰¹ More recently, it was demonstrated that BMP7 could also counteract the TGFβ-induced EMT in breast and prostate cancer. In addition, pretreatment, forced overexpression or systemic administration of BMP7 inhibited the formation of bone metastases from osteolytic breast and prostate cancer.^{60,102–105} It is tempting to speculate that BMP7, by counteracting TGF- β -induced EMT, may act as a differentiation-inducing agent that targets the cancer stem cell (CSC) fraction in breast and prostate cancer.^{105,106} However, more research is certainly warranted to address whether BMP7 is a valid novel approach in the treatment of breast and prostate cancer. As a potent inducer of bone formation, BMP7 is currently approved for clinical use in open fractures of long bones, non-unions and spinal fusion.¹⁰⁷

Halofuginone (Hfg) is a natural product derivative that is known to inhibit TGF- β signaling. It recently completed phase II clinical trials for the treatment of sarcoma (**Table 2**). Unpublished data from our laboratory show that Hfg inhibits TGF- β signaling *in vitro* in several cell types, and that the systemic daily treatment of Hfg in mice significantly inhibits the formation of osteolytic lesions and bone metastases after intracardiac inoculation of MDA-MB-231 breast, PC3 prostate cancer and 1205Lu melanoma cells.¹⁰⁸ Although the exact mechanism remains to be investigated, Hfg treatment represents another novel agent that may inhibit TGF- β signaling in bone metastasis.

Combination therapy. An attractive approach to increase treatment efficacy for patients with bone metastases is to combine treatments that antagonize the effects of TGF- β with other therapies. For example, targeting TGF- β signaling can enhance the therapeutic efficacy of various cytotoxic agents such as rapamycin¹⁰⁹ and doxorubicin.^{110,111} Unpublished studies in our laboratory show that SD-208 dosed in combination with an inhibitor of bone resorption, zoledronic acid, reduces the progression of established osteolytic metastases from breast cancer more effectively than either therapy alone.¹¹² Using the same bone metastasis model of MDA-MB-231 human breast cancer cells, we tested the effects of a combined treatment of SD-208 and 2-methoxyestradiol, an inhibitor of hypoxia-inducible factor 1a. Combined treatment with these agents reduced osteolytic lesions and tumor burden, and improved survival of mice more effectively than either treatment alone.⁵⁴

Risks, Limitations and Opportunities

As a result of its biological importance and wide variety of effect, blockade of TGF- β or its signaling provides intriguing therapeutic opportunities for the treatment of many different disease indications. However, potent and/or chronic inhibition of this wide-spread biologically important molecule may also potentially result in a variety of undesirable side effects.

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Risks. Knockout mice for TGF- β_1 display reduced numbers of regulatory T-lymphocytes, uncontrolled activation of the immune system and loss of immune tolerance resulting in generalized inflammation and autoimmunity.^{124–127} TGF- β blockade may also paradoxically increase the risk of tumorigenesis, 128, 129 inhibit wound healing¹³⁰ and is likely to be teratogenic to the fetus.¹³¹ However, severe toxicity has not been observed in animals when TGF- β is blocked by lifetime expression of a soluble T_βRII, under the regulation of the mammary-selective MMTV-LTR promoter,¹³² or when TGF-β-neutralizing antibodies are chronically administered for long periods of time.¹³³ In contrast to the severe chronic deficiency of TGF-B observed in knockout mouse models, neutralizing antibodies and soluble receptor decoys typically achieve only a partial deficiency of TGF- β , and may only interfere with the excessive TGF- β activity observed in the setting of cancer without significantly altering normal homeostatic TGF- β signaling. In clinical trials, TGF- β neutralizing antibodies, TGF-β ASOs and TGF-β receptor kinase inhibitors have been documented to be generally well tolerated. One patient developed premalignant skin lesion testing a TGF- β antibody from Genzyme, but upon discontinuation of the drug, this effect resolved with time. Furthermore, acquired resistance to the TGF-β receptor kinase inhibitor LY2109761 has recently been observed in a mouse skin model of de novo chemically induced carcinogenesis. The resultant carcinomas were more aggressive and inflammatory in nature, with de-localized and elevated expression of MMPs.¹³⁴ In addition, some toxicities may differ in rodent compared with primate models.

Limitations and opportunities. TGF-ß receptor kinase inhibitors are most suitable for oral dosing, have no loss of efficacy due to neutralizing antibody generation and better tissue penetration than observed with biologic-based agents.⁷² The wide variety of chemical pharmacophores and binding modes may eventually yield selective kinome profiles that may prove safe and effective for their designed purpose.^{72,135} However, the TGF- β receptor kinase inhibitors described to date are less selective than the current TGF- β ASOs or the biologic-based TGF- β -directed therapies. For example, the T β RI/ALK5 small-molecule kinase inhibitors can also potently inhibit ALK4 and less so ALK7 owing to their very close catalytic domain homology to TBRI/ALK5.¹¹⁷ In line with these observations, it may be challenging to dissect TGF- β -dependent toxicity from the off-target effects of these compounds. For this, biologic-based inhibitors may prove to be more indicative of true TGF- β -dependent toxicities owing to their highly selective nature.

The effects of blocking TGF- β on angiogenesis can be dependent on what type of inhibitor is used. TGF- β can indirectly stimulate angiogenesis by upregulating tumor production of vascular endothelial growth factor, which can be blocked with treatment with TGF- β antibodies, TGF- β ASOs or T β RI/ALK5 kinase inhibitors. Interestingly, while TGF- β antibodies and TGF- β ASOs block both the pro-(TGF- β /ALK1-mediated) as well as the anti-(TGF- β /ALK5-mediated) angiogenic effects in endothelial cells, T β RI/ALK5 kinase inhibitors block only the anti-angiogenic effects in endothelial cells. Indeed, several studies have demonstrated that treatment with T β RI/ALK5 kinase inhibitors can even promote angiogenesis.^{34,136,137} Therefore, the therapeutic strategies to target TGF- β must be carefully considered. Furthermore, it is interesting to speculate to what extend the mechanistic distinction may be due to the inhibition

of canonical versus non-canonical TGF- β signaling by these various agents.

It is imperative to keep in mind that the different therapeutic strategies to target TGF- β described in this review have fundamentally different properties regarding to the mechanism of action, pharmacokinetic properties and delivery challenges. Besides specificity, one of the main challenges with TGF- β ASOs will be its mode of administration to bone metastases. In addition, drug delivery challenges are also faced with neutralizing TGF- β antibodies and other large molecules such as T β RII/III:Fc fusion proteins.

Although speculative at present, some of the tumor-inhibiting effects of anti-TGF- β compounds described in this review may also be attributed to inhibitory effects on CSCs. To this end, as a proof-of-principle, treatment with the T β RI/ALK5 inhibitor LY2109761 inhibited TGF- β signaling in CD44+/CD24-/low breast CSC-like cells and reversed their mesenchymal and tumorigenic stem cell-like phenotype to a more epithelioid CD44-/low/CD24+ phenotype.¹⁴⁰ New studies are warranted to address whether TGF- β inhibitors also affect this CSC subpopulation in primary tumors and bone metastasis in both clinical and preclinical models.

A promising novel approach to overcome off-target tissue toxicity and poor drug exposure to tumor cells in bone metastatic disease are the use of bisphosphonate-coated liposomes or nanoparticles, which may be useful as a targeting device to sites of high bone turnover, including sites with bone metastatic disease.¹³⁸ Potentially, these bone-targeted liposomes may allow for a more prolonged local exposure to higher concentrations of the bioactive compounds described in this review, thereby enhancing therapeutic efficacy and minimizing systemic side effects. In addition, these bioactive compounds of interest could be delivered to bone metastatic sites in combination with other anti-cancer agents with synergistic action.

Another approach to target bone metastases was recently shown by using an oncolytic adenovirus expressing soluble T β RII-Fc.¹³⁹ This resulted in inhibition of established MDA-MB-231 bone metastases, but whether this activity may extend into the clinical situation remains to be studied and is unknown.

Biomarkers and patient stratification. Clinical trials are currently addressing whether plasma TGF- β levels in bone metastasis patients may be indicative of TGF- β -dependent metastatic disease and may be useful as a biomarker to predict the success of treatment with TGF- β antagonists in metastatic disease.

Immunohistochemical studies have also demonstrated that increased expression of TGF- β_1 and phosphorylated Smad2/3 in clinical samples from primary tumors are correlated with a high incidence of metastasis.^{140,141} However, immunohistochemical analyses only provide static snapshots of TGF- β signaling activity and fail to provide any information on the eventual gene expression changes induced by TGF- β . It is possible that mutations found downstream of the Smad proteins may not allow for appropriate TGF- β -induced gene responses. To circumvent this limitation, Padua *et al.* applied bioinformatics tools to define a TGF- β response signature that can be used to more accurately assess the TGF- β response status of human breast tumors.¹⁴² Analysis of clinical samples revealed that ~40% of human breast tumors actively responded to TGF- β signals. In addition, a validated signature of TGF- β upregulated

genes is clearly associated with poor prognosis of the subsets of basal-like- and Her2-positive breast cancers.¹⁴³ These findings suggest that the presence of a TGF- β response signature might drive tumor progression in estrogen-independent cancer, but may also reflect a suppressive host cell response in estrogen-dependent luminal cancers. Taken together, patient stratification based on the presence of the TGF- β response signature signature in primary tumor samples might help to identify a rationally tailored subset of patients that are likely to respond to anti-TGF- β therapies.

Conclusion

TGF- β is a multifunctional cytokine with a prominent role in cancer progression and bone metastasis. TGF- β is a major component of the bone and a central mediator in driving a feedforward cycle of tumor growth in bone. This has provided a powerful rationale to develop agents that inhibit TGF-B activity to prevent and/or treat bone metastases. Three therapeutic modalities targeting TGF- β have been pursued and are currently being tested in clinical trials in cancer patients (including bone metastatic disease): TGF- β antibodies, TGF- β receptor kinase inhibitors and TGF-B ASOs. All three modalities have fundamentally different pharmacokinetic/pharmacodynamic properties and mechanisms of actions, providing a broad range of limitations and opportunities, particularly with respect to specificity and toxicity. One benefit is that blockade of TGF- β signaling may increase bone mass in cancer patients. This may be particularly beneficial in a cancer population with already a low bone mineral density baseline resulting from either gonadal status or due to an oncolytic therapeutic regime including hormone-, radiation- or chemo-therapy. However, concerns about the impact on immune function and/or growthpromoting effects of these agents continue to remain. The next decade promises new and exciting clinical data that will determine which TGF- β therapeutic strategies will be most effective to prevent and/or cure bone metastatic disease coupled with a tolerable side-effect profile.

Conflict of Interest

TAG is a consultant for Amgen, Roche and Novartis, and has stock ownership of Amgen. KRS is an employee of Eli Lilly & Company. JTB has no potential conflict of interests.

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References

- 1. Coleman RE. Skeletal complications of malignancy. Cancer 1997;80:1588–1594.
- Jemal A, Siegel R, Xu J, Ward E. Cancer statistics, 2010. CA Cancer J Clin 2010;60:277–300.
 Mundy GR. Metastasis to bone: causes, consequences and therapeutic opportunities. Nat Rev
- Cancer 2002;2:584-593.
- 4. Roodman GD. Mechanisms of bone metastasis. N Engl J Med 2004;350:1655–1664.
- Korpal M, Yan J, Lu X, Xu S, Lerit DA, Kang Y. Imaging transforming growth factor-beta signaling dynamics and therapeutic response in breast cancer bone metastasis. *Nat Med* 2009;15: 960–966.

- Kang Y, Siegel PM, Shu W, Drobnjak M, Kakonen SM, Cordon-Cardo C et al. A multigenic program mediating breast cancer metastasis to bone. Cancer Cell 2003;3:537–549.
- Yin JJ, Selander K, Chirgwin JM, Dallas M, Grubbs BG, Wieser R *et al.* TGF-beta signaling blockade inhibits PTHrP secretion by breast cancer cells and bone metastases development. *J Clin Invest* 1999;103:197–206.
- Massague J, Blain SW, Lo RS. TGFbeta signaling in growth control, cancer, and heritable disorders. *Cell* 2000;103:295–309.
- Blobe GC, Schiemann WP, Lodish HF. Role of transforming growth factor beta in human disease. N Engl J Med 2000;342:1350–1358.
- ten Dijke P, Arthur HM. Extracellular control of TGFbeta signalling in vascular development and disease. Nat Rev Mol Cell Biol 2007;8:857–869.
- Wu MY, Hill CS. Tgf-beta superfamily signaling in embryonic development and homeostasis. Dev Cell 2009;16:329–343.
- Derynck R, Zhang YE. Smad-dependent and Smad-independent pathways in TGF-beta family signalling. Nature 2003;425:577–584.
- Shi Y, Massague J. Mechanisms of TGF-beta signaling from cell membrane to the nucleus. Cell 2003;113:685–700.
- Siegel PM, Shu W, Massague J. Mad upregulation and Id2 repression accompany transforming growth factor (TGF)-beta-mediated epithelial cell growth suppression. J Biol Chem 2003;278:35444–35450.
- Seyedin SM, Thomas TC, Thompson AY, Rosen DM, Piez KA. Purification and characterization of two cartilage-inducing factors from bovine demineralized bone. *Proc Natl Acad Sci USA* 1985;82:2267–2271.
- Seyedin SM, Thompson AY, Bentz H, Rosen DM, McPherson JM, Conti A et al. Cartilageinducing factor-A. Apparent identity to transforming growth factor-beta. J Biol Chem 1986;261:5693–5695.
- Janssens K, ten Dijke P, Janssens S, Van Hul W. Transforming growth factor-beta1 to the bone. Endocr Rev 2005;26:743–774.
- 18. Iqbal J, Sun L, Zaidi M. Coupling bone degradation to formation. Nat Med 2009;15:729-731.
- Tang Y, Wu X, Lei W, Pang L, Wan C, Shi Z et al. TGF-beta1-induced migration of bone mesenchymal stem cells couples bone resorption with formation. Nat Med 2009;15:757–765.
- Wu X, Pang L, Lei W, Lu W, Li J, Li Z et al. Inhibition of sca-1-positive skeletal stem cell recruitment by alendronate blunts the anabolic effects of parathyroid hormone on bone remodeling. *Cell Stem Cell* 2010;7:571–580.
- 21. Canalis E. Growth factor control of bone mass. J Cell Biochem 2009; 108:769-777.
- Alliston T, Choy L, Ducy P, Karsenty G, Derynck R. TGF-beta-induced repression of CBFA1 by Smad3 decreases cbfa1 and osteocalcin expression and inhibits osteoblast differentiation. *EMBO J* 2001;20:2254–2272.
- Maeda S, Hayashi M, Komiya S, Imamura T, Miyazono K. Endogenous TGF-beta signaling suppresses maturation of osteoblastic mesenchymal cells. *EMBO* J2004;23:552–563.
- Karsdal MA, Andersen TA, Bonewald L, Christiansen C. Matrix metalloproteinases (MMPs) safeguard osteoblasts from apoptosis during transdifferentiation into osteocytes: MT1-MMP maintains osteocyte viability. DNA Cell Biol 2004;23:155–165.
- Quinn JM, Itoh K, Udagawa N, Hausler K, Yasuda H, Shima N et al. Transforming growth factor beta affects osteoclast differentiation via direct and indirect actions. J Bone Miner Res 2001;16:1787–1794.
- Sells Galvin RJ, Gatlin CL, Horn JW, Fuson TR. TGF-beta enhances osteoclast differentiation in hematopoietic cell cultures stimulated with RANKL and M-CSF. *Biochem Biophys Res Commun* 1999;265:233–239.
- Balooch G, Balooch M, Nalla RK, Schilling S, Filvaroff EH, Marshall GW et al. TGF-beta regulates the mechanical properties and composition of bone matrix. Proc Natl Acad Sci USA 2005;102:18813–18818.
- Erlebacher A, Derynck R. Increased expression of TGF-beta 2 in osteoblasts results in an osteoporosis-like phenotype. J Cell Biol 1996;132:195–210.
- Mohammad KS, Chen CG, Balooch G, Stebbins E, McKenna CR, Davis H et al. Pharmacologic inhibition of the TGF-beta type I receptor kinase has anabolic and anti-catabolic effects on bone. PLoS ONE 2009;4:e5275.
- Derynck R, Akhurst RJ. Differentiation plasticity regulated by TGF-beta family proteins in development and disease. Nat Cell Biol 2007;9:1000–1004.
- 31. Massague J, Chen YG. Controlling TGF-beta signaling. Genes Dev 2000;14:627-644.
- Calonge MJ, Massague J. Smad4/DPC4 silencing and hyperactive Ras jointly disrupt transforming growth factor-beta antiproliferative responses in colon cancer cells. J Biol Chem 1999;274:33637–33643.
- 33. Massague J. TGFbeta in Cancer. Cell 2008;134:215-230.
- Pardali E, Goumans MJ, ten Dijke P. Signaling by members of the TGF-beta family in vascular morphogenesis and disease. *Trends Cell Biol* 2010;20:556–567.
- Thiery JP, Acloque H, Huang RY, Nieto MA. Epithelial-mesenchymal transitions in development and disease. *Cell* 2009;139:871–890.
- Gorsch SM, Memoli VA, Stukel TA, Gold LI, Arrick BA. Immunohistochemical staining for transforming growth factor beta 1 associates with disease progression in human breast cancer. *Cancer Res* 1992;52:6949–6952.
- Ivanovic V, Todorovic-Rakovic N, Demajo M, Neskovic-Konstantinovic Z, Subota V, Ivanisevic-Milovanovic O et al. Elevated plasma levels of transforming growth factor-beta 1 (TGF-beta 1) in patients with advanced breast cancer: association with disease progression. Eur J Cancer 2003;39:454–461.
- Buijs JT, van der Pluijm G. Osteotropic cancers: from primary tumor to bone. Cancer Lett 2009;273:177–193.
- Alexaki VI, Javelaud D, Van Kempen LC, Mohammad KS, Dennler S, Luciani F et al. GLI2-mediated melanoma invasion and metastasis. J Natl Cancer Inst 2010;102:1148–1159.

- Johnson RW, Nguyen MP, Padalecki SS, Grubbs BG, Merkel AR, Oyajobi BO et al. TGF-beta promotion of Gli2-induced expression of parathyroid hormone-related protein, an important osteolytic factor in bone metastasis, is independent of canonical Hedgehog signaling. *Cancer* Res 2011;71:822–831.
- Sterling JA, Oyajobi BO, Grubbs B, Padalecki SS, Munoz SA, Gupta A et al. The hedgehog signaling molecule Gli2 induces parathyroid hormone-related peptide expression and osteolysis in metastatic human breast cancer cells. *Cancer Res* 2006;66:7548–7553.
- Kakonen SM, Selander KS, Chirgwin JM, Yin JJ, Burns S, Rankin WA et al. Transforming growth factor-beta stimulates parathyroid hormone-related protein and osteolytic metastases via Smad and mitogen-activated protein kinase signaling pathways. J Biol Chem 2002;277:24571–24578.
- Safina A, Sotomayor P, Limoge M, Morrison C, Bakin AV. TAK1-TAB2 signaling contributes to bone destruction by breast carcinoma cells. *Mol Cancer Res* 2011;9:1042–1053.
- Guise TA, Yin JJ, Taylor SD, Kumagai Y, Dallas M, Boyce BF *et al.* Evidence for a causal role of parathyroid hormone-related protein in the pathogenesis of human breast cancer-mediated osteolysis. *J Clin Invest* 1996;98:1544–1549.
- Clines GA, Guise TA. Hypercalcaemia of malignancy and basic research on mechanisms responsible for osteolytic and osteoblastic metastasis to bone. *Endocr Relat Cancer* 2005;12:549–583.
- Mundy GR, Edwards JR. PTH-related peptide (PTHrP) in hypercalcemia. J Am Soc Nephrol 2008;19:672–675.
- Thomas RJ, Guise TA, Yin JJ, Elliott J, Horwood NJ, Martin TJ et al. Breast cancer cells interact with osteoblasts to support osteoclast formation. Endocrinology 1999;140:4451–4458.
- Powell GJ, Southby J, Danks JA, Stillwell RG, Hayman JA, Henderson MA et al. Localization of parathyroid hormone-related protein in breast cancer metastases: increased incidence in bone compared with other sites. *Cancer Res* 1991;51:3059–3061.
- Southby J, Kissin MW, Danks JA, Hayman JA, Moseley JM, Henderson MA et al. Immunohistochemical localization of parathyroid hormone-related protein in human breast cancer. Cancer Res 1990;50:7710–7716.
- Henderson MA, Danks JA, Slavin JL, Byrnes GB, Choong PF, Spillane JB *et al.* Parathyroid hormone-related protein localization in breast cancers predict improved prognosis. *Cancer Res* 2006;66:2250–2256.
- Guise TA. Breaking down bone: new insight into site-specific mechanisms of breast cancer osteolysis mediated by metalloproteinases. *Genes Dev* 2009;23:2117–2123.
- Lu X, Wang Q, Hu G, Van Poznak C, Fleisher M, Reiss M et al. ADAMTS1 and MMP1 proteolytically engage EGF-like ligands in an osteolytic signaling cascade for bone metastasis. *Genes Dev* 2009;23:1882–1894.
- Dunn LK, Mohammad KS, Fournier PG, McKenna CR, Davis HW, Niewolna M et al. Hypoxia and TGF-beta drive breast cancer bone metastases through parallel signaling pathways in tumor cells and the bone microenvironment. PLoS ONE 2009;4:e6896.
- Kang Y, Massague J. Epithelial-mesenchymal transitions: twist in development and metastasis. *Cell* 2004;118:277–279.
- 56. Deckers M, van Dinther M, Buijs J, Que I, Lowik C, van der Pluijm G et al. The tumor suppressor Smad4 is required for transforming growth factor beta-induced epithelial to mesenchymal transition and bone metastasis of breast cancer cells. Cancer Res 2006;66:2202–2209.
- Kang Y, He W, Tulley S, Gupta GP, Serganova I, Chen CR et al. Breast cancer bone metastasis mediated by the Smad tumor suppressor pathway. Proc Natl Acad Sci USA 2005;102: 13909–13914.
- Sethi N, Dai X, Winter CG, Kang Y. Tumor-derived JAGGED1 promotes osteolytic bone metastasis of breast cancer by engaging notch signaling in bone cells. *Cancer Cell* 2011;19: 192–205.
- Baselga J, Rothenberg ML, Tabernero J, Seoane J, Daly T, Cleverly A et al. TGF-beta signallingrelated markers in cancer patients with bone metastasis. *Biomarkers* 2008;13:217–236.
- Buijs JT, Henriquez NV, van Overveld PG, van der Horst G, ten Dijke P, van der Pluijm G. TGF-beta and BMP7 interactions in tumour progression and bone metastasis. *Clin Exp Metastasis* 2007;24:609–617.
- Serganova I, Moroz E, Vider J, Gogiberidze G, Moroz M, Pillarsetty N et al. Multimodality imaging of TGFbeta signaling in breast cancer metastases. FASEB J 2009;23:2662–2672.
- Bellahcene A, Castronovo V, Ogbureke KU, Fisher LW, Fedarko NS. Small integrin-binding ligand N-linked glycoproteins (SIBLINGs): multifunctional proteins in cancer. Nat Rev Cancer 2008;8:212–226.
- Bellahcene A, Kroll M, Liebens F, Castronovo V. Bone sialoprotein expression in primary human breast cancer is associated with bone metastases development. J Bone Miner Res 1996;11: 665–670.
- Hotte SJ, Winquist EW, Stitt L, Wilson SM, Chambers AF. Plasma osteopontin: associations with survival and metastasis to bone in men with hormone-refractory prostate carcinoma. *Cancer* 2002;95:506–512.
- Zhang L, Hou X, Lu S, Rao H, Hou J, Luo R et al. Predictive significance of bone sialoprotein and osteopontin for bone metastases in resected Chinese non-small-cell lung cancer patients: a large cohort retrospective study. Lung Cancer 2010;67:114–119.
- Brown JM, Wilson WR. Exploiting tumour hypoxia in cancer treatment. Nat Rev Cancer 2004;4:437–447.
- Hiraga T, Kizaka-Kondoh S, Hirota K, Hiraoka M, Yoneda T. Hypoxia and hypoxia-inducible factor-1 expression enhance osteolytic bone metastases of breast cancer. *Cancer Res* 2007;67:4157–4163.
- Lu X, Yan CH, Yuan M, Wei Y, Hu G, Kang Y. In vivo dynamics and distinct functions of hypoxia in primary tumor growth and organotropic metastasis of breast cancer. *Cancer Res* 2010;**70**: 3905–3914.

- McMahon S, Charbonneau M, Grandmont S, Richard DE, Dubois CM. Transforming growth factor beta1 induces hypoxia-inducible factor-1 stabilization through selective inhibition of PHD2 expression. J Biol Chem 2006;281:24171–24181.
- Buijs JT, Kuijpers CC, van der Pluijm G. Targeted therapy options for treatment of bone metastases; beyond bisphosphonates. *Curr Pharm Des* 2010;16:3015–3027.
- Kelly RJ, Morris JC. Transforming growth factor-beta: a target for cancer therapy. J Immunotoxicol 2010;7:15–26.
- Yingling JM, Blanchard KL, Sawyer JS. Development of TGF-beta signalling inhibitors for cancer therapy. Nat Rev Drug Discov 2004;3:1011–1022.
- Buijs JT, Stayrook KR, Guise TA. TGF-beta in the bone microenvironment: role in breast cancer metastases. *Cancer Microenviron* 2011;4:261–281.
- Biswas S, Nyman JS, Alvarez J, Chakrabarti A, Ayres A, Sterling J et al. Anti-transforming growth factor-beta antibody treatment rescues bone loss and prevents breast cancer metastasis to bone. PLoS One 2011;6:e27090.
- Ganapathy V, Ge R, Grazioli A, Xie W, Banach-Petrosky W, Kang Y et al. Targeting the transforming growth factor-beta pathway inhibits human basal-like breast cancer metastasis. *Mol Cancer* 2010;9:122.
- Morris JC, Shapiro GI, Tan AR, Lawrence DP, Olencki TE, Dezube BJ et al. Phase I/II study of GC1008: a human anti-transforming growth factor-beta (TGF-β) monoclonal antibody (MAb) in patients with advanced malignant melanoma (MM) or renal cell carcinoma (RCC). J Clin Oncol 2008;26:9028.
- Mancuso P, Shalinsky DR, Calleri A, Quarna J, Antoniotti P, Jilani I *et al.* Evaluation of ALK-1 expression in circulating endothelial cells (CECs) as an exploratory biomarker for PF-03446962 undergoing phase I trial in cancer patients. *J Clin Oncol* 2009;27:15s: abstr 3573.
- Bennett CF, Swayze EE. RNA targeting therapeutics: molecular mechanisms of antisense oligonucleotides as a therapeutic platform. *Annu Rev Pharmacol Toxicol* 2010;50:259–293.
- 79. Crooke ST. Progress in antisense technology. *Annu Rev Med* 2004;**55**:61–95.
- Hau P, Jachimczak P, Schlingensiepen R, Schulmeyer F, Jauch T, Steinbrecher A et al. Inhibition of TGF-beta2 with AP 12009 in recurrent malignant gliomas: from preclinical to phase I/II studies. Oligonucleotides 2007;17:201–212.
- Bogdahn U, Hau P, Stockhammer G, Venkataramana NK, Mahapatra AK, Suri A *et al.* Targeted therapy for high-grade glioma with the TGF-beta2 inhibitor trabedersen: results of a randomized and controlled phase IIb study. *Neuro Oncol* 2011;13:132–142.
- Fakhrai H, Mantil JC, Liu L, Nicholson GL, Murphy-Satter CS, Ruppert J et al. Phase I clinical trial of a TGF-beta antisense-modified tumor cell vaccine in patients with advanced glioma. *Cancer Gene Ther* 2006;13:1052–1060.
- Byfield SD, Roberts AB. Lateral signaling enhances TGF-beta response complexity. Trends Cell Biol 2004;14:107–111.
- Fu K, Corbley MJ, Sun L, Friedman JE, Shan F, Papadatos JL et al. SM16, an orally active TGF-beta type I receptor inhibitor prevents myofibroblast induction and vascular fibrosis in the rat carotid injury model. Arterioscler Thromb Vasc Biol 2008;28:665–671.
- Inman GJ, Nicolas FJ, Callahan JF, Harling JD, Gaster LM, Reith AD et al. SB-431542 is a potent and specific inhibitor of transforming growth factor-beta superfamily type I activin receptor-like kinase (ALK) receptors ALK4, ALK5, and ALK7. Mol Pharmacol 2002;62:65–74.
- Petersen M, Thorikay M, Deckers M, van Dinther M, Grygielko ET, Gellibert F et al. Oral administration of GW788388, an inhibitor of TGF-beta type I and II receptor kinases, decreases renal fibrosis. *Kidney Int* 2008;**73**:705–715.
- Tojo M, Hamashima Y, Hanyu A, Kajimoto T, Saitoh M, Miyazono K *et al.* The ALK-5 inhibitor A-83-01 inhibits Smad signaling and epithelial-to-mesenchymal transition by transforming growth factor-beta. *Cancer Sci* 2005;**96**:791–800.
- Eijken M, Swagemakers S, Koedam M, Steenbergen C, Derkx P, Uitterlinden AG et al. The activin A-follistatin system: potent regulator of human extracellular matrix mineralization. FASEB J 2007;21:2949–2960.
- Pearsall RS, Canalis E, Cornwall-Brady M, Underwood KW, Haigis B, Ucran J et al. A soluble activin type IIA receptor induces bone formation and improves skeletal integrity. Proc Natl Acad Sci USA 2008;105:7082–7087.
- Mohammad KS, Javelaud D, Fournier PG, Niewolna M, McKenna CR, Peng XH et al. TGF-beta-RI kinase inhibitor SD-208 reduces the development and progression of melanoma bone metastases. Cancer Res 2011;71:175–184.
- Stebbins EG, Mohammad KS, Niewolna M, McKenna CR, Mison AP, Schimmoller F et al. SD-208, a small molecule inhibitor of transforming growth factor β receptor I kinase reduces breast cancer metastases to bone and improves survival in a mouse model. J Bone Miner Res 2005;20:1216 (abstract).
- Fournier PJ, Mohammad KS, McKenna CR, Peng X, Chirgwin JM, Guise TA.TGF-beta blockade inhibits osteolytic metastases but not osteoblastic prostate cancer metastases. #SA274. *American Society for Bone and Mineral Research* 30th annual meeting; 2008; Montreal, QC, Canada; 2008.
- Zhang B, Halder SK, Zhang S, Datta PK. Targeting transforming growth factor-beta signaling in liver metastasis of colon cancer. *Cancer Lett* 2009;277:114–120.
- Wan X, Li ZG, Yingling JM, Yang J, Starbuck MW, Ravoori MK et al. Effect of transforming growth factor beta (TGF-beta) receptor I kinase inhibitor on prostate cancer bone growth. *Bone* 2012;50:695–703.
- Melisi D, Ishiyama S, Sclabas GM, Fleming JB, Xia Q, Tortora G et al. LY2109761, a novel transforming growth factor beta receptor type I and type II dual inhibitor, as a therapeutic approach to suppressing pancreatic cancer metastasis. Mol Cancer Ther 2008;7:829–840.
- Calvo-Aller E, Baselga J, Glatt S, Cleverly A, Lahn M, Arteaga CL *et al.* First human dose escalation study in patients with metastatic malignancies to determine safety and pharmacokinetics of LY2157299, a small molecule inhibitor of the transforming growth factorbeta receptor I kinase. *J Clin Oncol* 2008;26:14554 (abstract).

- 97. Chen D, Zhao M, Mundy GR. Bone morphogenetic proteins. Growth Factors 2004;22: 233-241.
- ten Dijke P. Bone morphogenetic protein signal transduction in bone. Curr Med Res Opin 2006;22 (Suppl 1): S7–S11.
- Dudley AT, Lyons KM, Robertson EJ. A requirement for bone morphogenetic protein-7 during development of the mammalian kidney and eye. *Genes Dev* 1995;9:2795–2807.
- Luo G, Hofmann C, Bronckers AL, Sohocki M, Bradley A, Karsenty G. BMP-7 is an inducer of nephrogenesis, and is also required for eye development and skeletal patterning. *Genes Dev* 1995;9:2808–2820.
- Simic P, Vukicevic S. Bone morphogenetic proteins in development and homeostasis of kidney. Cytokine Growth Factor Rev 2005;16:299–308.
- Buijs JT, Henriquez NV, van Overveld PG, van der Horst G, Que I, Schwaninger R et al. Bone morphogenetic protein 7 in the development and treatment of bone metastases from breast cancer. Cancer Res 2007;67:8742–8751.
- Buijs JT, Petersen M, van der Horst G, van der Pluijm G. Bone morphogenetic proteins and its receptors; therapeutic targets in cancer progression and bone metastasis? *Curr Pharm Des* 2010;16:1291–1300.
- Buijs JT, Rentsch CA, van der Horst G, van Overveld PG, Wetterwald A, Schwaninger R et al. BMP7, a putative regulator of epithelial homeostasis in the human prostate, is a potent inhibitor of prostate cancer bone metastasis in vivo. Am J Pathol 2007;171:1047–1057.
- Buijs JT, van der Horst G, van den Hoogen C, Cheung H, de Rooij B, Kroon J *et al.* The BMP2/7 heterodimer inhibits the human breast cancer stem cell subpopulation and bone metastases formation. *Oncogene* 2012;31:2164–2174.
- van der Pluijm G. Epithelial plasticity, cancer stem cells and bone metastasis formation. Bone 2011;48:37–43.
- Gautschi OP, Frey SP, Zellweger R. Bone morphogenetic proteins in clinical applications. ANZ J Surg 2007;77:626–631.
- Juarez P, Chirgwin J, Mohammad KS, Ryan C, Walton H, Niewolna M et al. Halofuginone decreases osteolytic bone metastases. #MO0109. J Bone Miner Res 2009;24:1.
- Gadir N, Jackson DN, Lee E, Foster DA. Defective TGF-beta signaling sensitizes human cancer cells to rapamycin. Oncogene 2008;27:1055–1062.
- Filyak Y, Filyak O, Stoika P. Transforming growth factor beta-1 enhances cytotoxic effect of doxorubicin in human lung adenocarcinoma cells of A549 line. Cell Biol Int 2007;31:851–855.
- Taniguchi Y, Kawano K, Minowa T, Sugino T, Shimojo Y, Maitani Y. Enhanced antitumor efficacy of folate-linked liposomal doxorubicin with TGF-beta type I receptor inhibitor. *Cancer Sci* 2010;101:7.
- 112. Mohammad KS, Stebbins EG, Kingsley L, Fournier PGJ, Niewolna M, McKenna CR et al. Combined transforming growth factor β receptor I kinase inhibitor and biphosphonates are additive to reduce breast cancer bone metastases. J Bone Miner Res 2008;23:F275 (Abstract).
- Dasch JR, Pace DR, Waegell W, Inenaga D, Ellingsworth L. Monoclonal antibodies recognizing transforming growth factor-beta. Bioactivity neutralization and transforming growth factor beta 2 affinity purification. *J Immunol* 1989;142:1536–1541.
- 114. Ehata S, Hanyu A, Fujime M, Katsuno Y, Fukunaga E, Goto K et al. Ki26894, a novel transforming growth factor-beta type I receptor kinase inhibitor, inhibits in vitro invasion and in vivo bone metastasis of a human breast cancer cell line. Cancer Sci 2007;98:127–133.
- Bueno L, de Alwis DP, Pitou C, Yingling J, Lahn M, Glatt S et al. Semi-mechanistic modelling of the tumour growth inhibitory effects of LY2157299, a new type I receptor TGF-beta kinase antagonist, in mice. Eur J Cancer 2008;44:142–150.
- Bandyopadhyay A, Agyin JK, Wang L, Tang Y, Lei X, Story BM *et al.* Inhibition of pulmonary and skeletal metastasis by a transforming growth factor-beta type I receptor kinase inhibitor. *Cancer Res* 2006;66:6714–6721.
- Peng SB, Yan L, Xia X, Watkins SA, Brooks HB, Beight D et al. Kinetic characterization of novel pyrazole TGF-beta receptor I kinase inhibitors and their blockade of the epithelialmesenchymal transition. Biochemistry 2005;44:2293–2304.
- Bonniaud P, Margetts PJ, Kolb M, Schroeder JA, Kapoun AM, Damm D *et al.* Progressive transforming growth factor beta1-induced lung fibrosis is blocked by an orally active ALK5 kinase inhibitor. *Am J Respir Crit Care Med* 2005;**171**:889–898.
- 119. Ge R, Rajeev V, Ray P, Lattime E, Rittling S, Medicherla S *et al.* Inhibition of growth and metastasis of mouse mammary carcinoma by selective inhibitor of transforming growth factor-beta type I receptor kinase *in vivo. Clin Cancer Res* 2006;**12**:4315–4330.
- 120. Uhl M, Aulwurm S, Wischhusen J, Weiler M, Ma JY, Almirez R et al. SD-208, a novel transforming growth factor beta receptor I kinase inhibitor, inhibits growth and invasiveness and enhances immunogenicity of murine and human glioma cells in vitro and in vivo. Cancer Res 2004;64:7954–7961.

- Wojtowicz-Praga S, Verma UN, Wakefield L, Esteban JM, Hartmann D, Mazumder A. Modulation of B16 melanoma growth and metastasis by anti-transforming growth factor beta antibody and interleukin-2. *J Immunother Emphasis Tumor Immunol* 1996;19:169–175.
- Nemunaitis J, Dillman RO, Schwarzenberger PO, Senzer N, Cunningham C, Cutler J et al. Phase II study of belagenpumatucel-L, a transforming growth factor beta-2 antisense gene-modified allogeneic tumor cell vaccine in non-small-cell lung cancer. J Clin Oncol 2006;24:4721–4730.
- 123. Nemunaitis J, Nemunaitis M, Senzer N, Snitz P, Bedell C, Kumar P et al. Phase II trial of Belagenpumatucel-L, a TGF-beta2 antisense gene modified allogeneic tumor vaccine in advanced non small cell lung cancer (NSCLC) patients. *Cancer Gene Ther* 2009;16: 620–624.
- Gorelik L, Flavell RA. Abrogation of TGFbeta signaling in T cells leads to spontaneous T cell differentiation and autoimmune disease. *Immunity* 2000;12:171–181.
- 125. Kulkarni AB, Huh CG, Becker D, Geiser A, Lyght M, Flanders KC *et al.* Transforming growth factor beta 1 null mutation in mice causes excessive inflammatory response and early death. *Proc Natl Acad Sci USA* 1993;**90**:770–774.
- Shull MM, Ormsby I, Kier AB, Pawlowski S, Diebold RJ, Yin M *et al.* Targeted disruption of the mouse transforming growth factor-beta 1 gene results in multifocal inflammatory disease. *Nature* 1992;359:693–699.
- Yaswen L, Kulkarni AB, Fredrickson T, Mittleman B, Schiffman R, Payne S *et al*. Autoimmune manifestations in the transforming growth factor-beta 1 knockout mouse. *Blood* 1996;87: 1439–1445.
- Bhowmick NA, Chytil A, Plieth D, Gorska AE, Dumont N, Shappell S et al. TGF-beta signaling in fibroblasts modulates the oncogenic potential of adjacent epithelia. Science 2004;303: 848–851.
- Cheng N, Chytil A, Shyr Y, Joly A, Moses HL. Transforming growth factor-beta signalingdeficient fibroblasts enhance hepatocyte growth factor signaling in mammary carcinoma cells to promote scattering and invasion. *Mol Cancer Res* 2008;6:1521–1533.
- Prud'homme GJ. Pathobiology of transforming growth factor beta in cancer, fibrosis and immunologic disease, and therapeutic considerations. *Lab Invest* 2007;87:1077–1091.
- Klopcic B, Maass T, Meyer E, Lehr HA, Metzger D, Chambon P et al. TGF-beta superfamily signaling is essential for tooth and hair morphogenesis and differentiation. Eur J Cell Biol 2007;86:781–799.
- Yang YA, Dukhanina O, Tang B, Mamura M, Letterio JJ, MacGregor J et al. Lifetime exposure to a soluble TGF-beta antagonist protects mice against metastasis without adverse side effects. J Clin Invest 2002;109:1607–1615.
- Ruzek MC, Hawes M, Pratt B, McPherson J, Ledbetter S, Richards SM et al. Minimal effects on immune parameters following chronic anti-TGF-beta monoclonal antibody administration to normal mice. Immunopharmacol Immunotoxicol 2003;25:235–257.
- Connolly EC, Saunier EF, Quigley D, Luu MT, De Sapio A, Hann B et al. Outgrowth of drugresistant carcinomas expressing markers of tumor aggression after long-term TbetaRI/II kinase inhibition with LY2109761. Cancer Res 2011;71:2339–2349.
- Bonafoux D, Lee WC. Strategies for TGF-beta modulation: a review of recent patents. Expert Opin Ther Pat 2009;19:1759–1769.
- Liu Z, Kobayashi K, van Dinther M, van Heiningen SH, Valdimarsdottir G, van Laar T *et al.* VEGF and inhibitors of TGFbeta type-I receptor kinase synergistically promote blood-vessel formation by inducing alpha5-integrin expression. *J Cell Sci* 2009;**122**:3294–3302.
- Watabe T, Nishihara A, Mishima K, Yamashita J, Shimizu K, Miyazawa K et al. TGF-beta receptor kinase inhibitor enhances growth and integrity of embryonic stem cell-derived endothelial cells. J Cell Biol 2003;163:1303–1311.
- Hengst V, Oussoren C, Kissel T, Storm G. Bone targeting potential of bisphosphonate-targeted liposomes. Preparation, characterization and hydroxyapatite binding *in vitro*. Int J Pharm 2007;331:224–227.
- 139. Hu Z, Gerseny H, Zhang Z, Chen YJ, Berg A, Stock S et al. Oncolytic adenovirus expressing soluble TGFbeta receptor II-Fc-mediated inhibition of established bone metastases: a safe and effective systemic therapeutic approach for breast cancer. Mol Ther 2011;19:1609–1618.
- 140. Dalal BI, Keown PA, Greenberg AH. Immunocytochemical localization of secreted transforming growth factor-beta 1 to the advancing edges of primary tumors and to lymph node metastases of human mammary carcinoma. *Am J Pathol* 1993;**143**:381–389.
- Shinto O, Yashiro M, Toyokawa T, Nishii T, Kaizaki R, Matsuzaki T et al. Phosphorylated smad2 in advanced stage gastric carcinoma. BMC Cancer 2010;10:652.
- Padua D, Zhang XH, Wang Q, Nadal C, Gerald WL, Gomis RR et al. TGFbeta primes breast tumors for lung metastasis seeding through angiopoietin-like 4. Cell 2008;133:66–77.
- Tan AR, Alexe G, Reiss M. Transforming growth factor-beta signaling: emerging stem cell target in metastatic breast cancer? *Breast Cancer Res Treat* 2009;115:453–495.