

ORIGINAL RESEARCH

OPEN ACCESS Full open access to this and thousands of other papers at http://www.la-press.com.

Do Hormonal Characteristics of the Luteal Phase Affect the Conception Rate of Women Undergoing IUI Treatment in the Following Menstrual Cycle?

Misao Fukuda¹, Kiyomi Fukuda¹, Anne Grete Byskov² and Claus Yding Andersen²

¹M&K Health Institute, 30-9 Kariya, Ako, Hyogo 678-0239, Japan. ²Laboratory of Reproductive Biology, Juliane Marie Center for Children, Women and Reproduction, Rigshospitalet, Section 5712, University Hospital of Copenhagen, Blegdamsvej 9, DK-2100, Copenhagen, Denmark. Corresponding author email: web@fukuda8767.com

Abstract

Objective: The present study was performed to evaluate whether the hormone profiles of the mid-luteal phase impact on the chances of conceiving during the following menstrual cycle in connection with intrauterine insemination (IUI) treatment. **Design:** Observational clinical study.

Design: Observational clinical si

Setting: Infertility clinic.

Patient(s): 92 women underwent a total of 288 IUI treatment cycles, without use of exogenous hormones.

Intervention(s): The mid-luteal hormone profiles including levels of luteinizing hormone (LH), follicle-stimulating hormone (FSH), prolactin, oestradiol, progesterone, androstenedione and testosterone, were measured in the preceding cycle of IUI treatment.

Main outcome measure(s): The mid-luteal hormone profiles were correlated to whether the women conceived in the following natural menstrual cycle in which IUI treatment was performed and to whether ovulation occurred on the same ovary in two consecutive cycles (ipsilateral ovulation) or jumped from one ovary to the other (contralateral ovulation).

Result(s): When ovulation occurred from the same ovary in two consecutive cycles mid-luteal levels of progesterone of the preceeding cycle were significantly lower in conceptional than non-conceptional cycles. No significant differences of LH, FSH, estradiol, androstendione and testosterone were observed when correlating the different parameters.

Conclusion(s): Lower mid-luteal progesterone levels seem to enhance chances of conception when ovulation occurs in the same ovary for two consecutive cycles.

Keywords: conceptional cycle, mid-luteal hormone profile, natural menstrual cycle, ovulation pattern, progesterone

Reproductive Biology Insights 2011:4 11–15

doi: 10.4137/RBI.S6824

This article is available from http://www.la-press.com.

© the author(s), publisher and licensee Libertas Academica Ltd.

This is an open access article. Unrestricted non-commercial use is permitted provided the original work is properly cited.

Introduction

In natural menstrual cycles conditions in one cycle may affect conditions of the following cycle. When ovulations alternate between the two ovaries the follicular phase length is shorter compared to when ovulation occur from the same ovary.^{1–3} Moreover, ovulation jumping between the ovaries in two consecutive menstrual cycles seemed to be accompanied by increased chances of conception in the latter of the two cycles as compared to two ovulations originating in the same ovary.^{3–8} There is also some evidence to suggest that oocytes released from the right ovary possess a higher pregnancy potential than oocytes released from the left ovary.⁹

Previously we found that hormonal characteristics on cycle day 3 in the normal menstrual cycle correlated to the likelihood of achieving conception in connection with IUI treatment, expressed by increased levels of oestradiol and an increased oestradiol/androgen and oestradiol/FSH ratios.¹⁰ These observations have in the present study been extended to include measurements of hormonal profiles of the midluteal phase of the preceding cycle and have been related to whether the ovulation occurred in the same ovary or jumped between the ovaries.

The aim of the present study was to understand further how hormones of the luteal phase affect the conception rate of women undergoing IUI treatment in the following menstrual cycle.

Materials and Methods

Data for this study was collected between August 1997 and November 2002. This is an observational study. A total of 92 women receiving treatment with IUI were included. All women showed regular menstrual cycles (29.4 \pm 2.4 days, range 25–35) with an intercycle variation of less than 7 days. They all had two intact ovaries without ovarian cysts and none had ovulation disorders. All these 92 women underwent sonohysterosalpingography and they showed bilateral tubal patency and no endometrial cavity abnormalities.¹¹ The IUI treatment resulted in 44 women conceived (age: 29.8 ± 3.8 , mean \pm SD, range 22-38). In total these 44 women had undergone a total of 117 treatment cycles (ie, 44 conceptional and 73 non-conceptional). The remaining 48 women who did not conceive underwent a total of 171

conceived were male factor, 34 and unknown, 10. Those among the 48 women who did not conceive were male factor, 37 and unknown, 11. We compared women's age, body mass index (BMI), husband's age and semen analysis between pregnant women and non-pregnant women. In the present study we used 'pregnant woman' who had conceptional cycle and 'non-pregnant woman' who did not show any conceptional cycle but showed only non-conceptional cycles during IUI treatment. None of the women received any exogenous medication. Following informed consent blood samples were drawn for assessing the hormone profiles 7 days after ovulation (defined by follicle disappearance as observed with transvaginal ultrasound) in the first of the two consecutive cycles and treatment was performed in connection with the second cycle. Monitoring included assessment of concentrations of serum luteinizing hormone (LH: mIU/ml), follicle-stimulating hormone (FSH: mIU/ml), prolactin (ng/ml), oestradiol (E2: pg/ml), progesterone (P: ng/ml), androstenedione (A: ng/ml) and testosterone (T: ng/dl) 7 days after ovulation. Hormone concentrations were measured using automated chemiluminescence system (Bayer-Medical Ltd, Tokyo, Japan) for LH, FSH, prolactin, E2, P and T and radioimmunoassay for A. Intraassay variance was within 5% and interassay variance was within 6%. IUI was performed on a next day following a positive urinary LH test or on the same day when the LH test was strongly positive. Clinical pregnancy was confirmed by the presence of a gestational sac with transvaginal ultrasound. The study protocol was approved by the Institutional Review Board of M&K Health Institute.

treatment cycles (age 30.3 ± 3.8 years, range 23-38). The causes of infertility among the 44 women who

Statistical evaluation of multiple groups was performed using ANOVA first and when a statistical difference was detected Tukey-Kramer procedure was used to perform one by one comparison among four groups. Differences were considered significant at P < 0.05. Results are presented as mean \pm SD.

Results

As shown in Table 1 there were no significant differences of women's age, BMI, husband's age and semen analysis between pregnant women and nonpregnant women.





Table 1. Comparison of women's age, BMI, husband's age and semen analysis between pregnant women and non-pregnant women.

	Pregnant women (n = 44)	Non-pregnant women (n = 48)	P-value
Women's	29.8 ± 3.8	30.3 ± 3.8	<i>P</i> = 0.6041
age (years)	(22–38)	(23–38)	
BMI	20.5 ± 2.7	20.7 ± 2.6	<i>P</i> = 0.7371
(kg/m ²)	(17.1–30.3)	(17.3–30.0)	
Husband's age (years)	33.5 ± 4.5 (25–43)	33.4 ± 5.1 (24–42)	<i>P</i> = 0.8663
Sperm	13.1 ± 9.7	13.4 ± 10.0	<i>P</i> = 0.8924
count (10 ⁶ /ml)	(3–50)	(2–60)	
% motility	25.9 ± 19.6	26.3 ± 18.5	<i>P</i> = 0.9322
(%)	(4–70)	(5–75)	

Note: Mean \pm SD.

The mid-luteal serum concentrations of LH, FSH, prolactin, E2, P, A and T of the preceding cycle prior to the IUI treatment cycle in which ovulation was either occurring contralateral (C) or ipsilateral (I) side in relation to previous ovulation irrespective of conception are shown in Table 2. There were no significant differences of different hormone levels between (C) and (I).

The mid-luteal serum concentrations of the preceding cycle in relation to the (C) and (I) ovulation pattern of conceptional and non-conceptional cycles in the group of women who became pregnant and in those who did not conceive are shown in Table 3. With regard to (C) and (I) ovulation patterns there were no significant differences in levels of LH, FSH, prolactin, E2, A and T between conceptional and nonconceptional cycles. *P* levels of ipsilateral ovulation in conceptional cycles were significantly lower than that of non-conceptional cycles among both pregnant and non-pregnant women (AC vs. AI, P = 0.0454; AI vs. BI, P = 0.0335).

Discussion

The present study shows that conditions created by the corpus luteum is likely to affect the following menstrual cycle and have an impact of whether or not conception will occur, at least in connection with IUI treatment. When ovulation occurs on the same ovary in two consecutive menstrual cycles lower mid-luteal progesterone levels of the preceding cycle seem to augment chances of pregnancy during the subsequent cycle, although concentrations of a number of other hormones remain similar. This may express a local inhibitory effect of progesterone on follicular development, which is then manifested in the following cycle as a reduced chance of conception.

Serum progesterone levels have been used to measure luteal function. The peak of progesterone level is seen 7 to 8 days after ovulation and therefore we measured the hormone profiles 7 days after follicle disappearance by transvaginal ultrasound. Because the progesterone secretion from corpus luteum is pulsatile in nature three measurements between 5 to 9 days after ovulation is recommended.¹² However, in order to reduce patients' pain we measured one sample of progesterone levels during the morning hours which are generally the highest in nature.¹³

There is ample evidence to suggest that progesterone may have a local inhibitory effect on follicular growth in animal studies^{14,15} and in humans.^{16,17} The effect of progesterone is expressed through a progesterone-binding protein with receptor like features located on the surface of rat granulosa cells.^{18,19} Progesterone suppressed the binding of ¹²⁵I-FSH to porcine granulosa cells in vitro.²⁰ In addition, progesterone caused atresia of oocytes and follicles using mouse ovarian fragments in vitro.²¹ Progesterone inhibited FSH-stimulated oestrogen production through inhibition of aromatase activity

	LH (iu/ml)	FSH (iu/ml)	Prolactin (ng/ml)	E2 (pg/ml)	P (ng/ml)	A (ng/ml)	T (ng/dl)
С	4.4 ± 2.6 (n = 155)	2.8 ± 1.2 (n = 155)	20 ± 11 (n = 155)	142 ± 44 (n = 157)	21.2 ± 7.3 (n = 157)	1.88 ± 0.66 (n = 126)	26.8 ± 13.6 (n = 123)
I	4.5 ± 2.6 (n = 131)	2.7 ± 1.0 (n = 131)	21 ± 10 ((n = 131)	144 ± 53 (n = 131)	21.2 ± 8.1 (n = 131)	1.93 ± 0.64 (n = 113)	26.9 ± 12.5 (n = 109)
Total	(n = 286)	(n = 101) 2.8 ± 1.1 (n = 286)	20 ± 10 (n = 286)	143 ± 48 (n = 288)	(n = 288)	(n = 1.0) 1.90 ± 0.65 (n = 239)	(n = 100) 26.9 ± 13.1 (n = 232)

Table 2. Mid-luteal hormone profiles of the former cycle of contralateral (C) and ipsilateral (I) ovulations of all cycles.

Note: Mean \pm SD.



	LH (iu/ml)	FSH (iu/ml)	Prolactin (ng/ml)	E2 (pg/ml)	P (ng/ml)	A (ng/ml)	T (ng/dl)
Α							
C I	$\begin{array}{c} 4.3 \pm 2.8 \\ (n=25) \\ 4.7 \pm 2.9 \\ (n=17) \end{array}$	3.0 ± 1.2 (n = 25) 2.8 ± 1.0 (n = 17)	$\begin{array}{l} 17 \pm 9 \\ (n = 25) \\ 22 \pm 11 \\ (n = 17) \end{array}$	$\begin{array}{c} 132 \pm 45 \\ (n=27) \\ 154 \pm 76 \\ (n=17) \end{array}$	$\begin{array}{l} 22.8 \pm 8.0^{\#} \\ (n=27) \\ 16.5 \pm 7.8^{\#} \\ (n=17) \end{array}$	$\begin{array}{c} 1.81 \pm 0.74 \\ (n=20) \\ 1.72 \pm 0.47 \\ (n=16) \end{array}$	$\begin{array}{c} 26.9 \pm 15.4 \\ (n=20) \\ 21.6 \pm 12.9 \\ (n=16) \end{array}$
B C I	$\begin{array}{l} 4.3 \pm 2.6 \\ (n = 130) \\ 4.4 \pm 2.6 \\ (n = 114) \end{array}$	2.8 ± 1.2 (n = 130) 2.7 ± 1.1 (n = 114)	21 ± 11 (n = 130) 21 ± 10 (n = 114)	145 ± 44 (n = 130) 142 \pm 50 (n = 114)	$\begin{array}{l} 20.9 \pm 7.3 \\ (n=130) \\ 22.0 \pm 7.9^{\#} \\ (n=114) \end{array}$	$\begin{array}{l} 1.90 \pm 0.63 \\ (n=106) \\ 1.97 \pm 0.66 \\ (n=97) \end{array}$	$\begin{array}{c} 26.8 \pm 12.6 \\ (n=103) \\ 27.8 \pm 12.2 \\ (n=93) \end{array}$

Table 3. Mid-luteal hormone profiles of the former cycle of contralateral (C) and ipsilateral (I) ovulations of conceptional (**A**) and non-conceptional cycles (**B**).

Notes: Mean ± SD. ANOVA: P(Progesterone), P = 0.0364; #AC vs. AI, P = 0.0454; AI vs. BI, P = 0.0335.

in cultured rat granulosa cells, suggesting a direct but reversible inhibitory action of progesterone on follicular development.^{22–24} Also, progesterone administered before pregnant mares' serum (PMS) injection appears to inhibit early follicular growth and causes atresia, suppressing the proliferation of granulosa cells, and consequently suppresses superovulation induced by PMS and human chorionic gonadotropin (HCG) in hypophysectomized rats.²⁵ Taken together, results from the present study in combination with results from animal studies suggest that progesterone exerts a local paracrine effect on ovaries that reduces the optimal developmental capacity of subsequent follicles of that ovary during the following cycle.

A number of studies suggest a local antifolliculogenic effect of the corpus luteum.^{26–31} Also, a negative effect of the corpus luteum on follicular growth is already present in the actual luteal phase, since the diameter of the largest follicle in the ipsilateral ovary containing the corpus luteum is smaller than that of the contralateral ovary without corpus luteum.³² The present study corroborates these earlier findings and warrants further studies to elucidate the mechanisms by which progesterone and the corpus luteum in humans are capable of affecting follicular development prior to the gonadotrophin dependent stage.

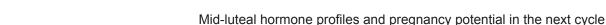
In conclusion, lower mid-luteal progesterone levels are accompanied by a higher chance of becoming pregnant in the next cycle, especially in connection with ipsilateral ovulations. Corpus luteum, with progesterone as a likely mediator substance, may affect the health status of developing follicles locally and may subsequently affect the likelihood of releasing an oocyte with an optimal pregnancy potential. Thus, there seems to exist a link between midluteal hormone profiles (conditions of corpus luteum) and pregnancy potential of the oocyte of the dominant follicle of the next following cycle. The present study therefore suggests a renewed interest in the actions of progesterone exerted on small antal follicles that may further explain the present findings.

Acknowledgements

We are grateful to Mr. Yamauchi, Mr. Mizugami, Ms. Suzuki, and Mr. Moori of ASKA Pharmaceutical for their kind assistance.

Disclosures

Author(s) have provided signed confirmations to the publisher of their compliance with all applicable legal and ethical obligations in respect to declaration of conflicts of interest, funding, authorship and contributorship, and compliance with ethical requirements in respect to treatment of human and animal test subjects. If this article contains identifiable human subject(s) author(s) were required to supply signed patient consent prior to publication. Author(s) have confirmed that the published article is unique and not under consideration nor published by any other publication and that they have consent to reproduce any copyrighted material. The peer reviewers declared no conflicts of interest.



References

- 1. Wallach EE, Virutamasen P, Wright KH. 1973 Menstrual cycle characteristics and side of ovulation in the Rhesus monkey. *Fertil Steril*. 1973;24:715–21.
- Potashnik G, Insler V, Meizner I. Frequency, sequence, and side of ovulation in women menstruating normally. *Br Med J.* 1987;294:219.
- Fukuda M, Fukuda K, Yding Andersen C, Byskov AG. Contralateral selection of dominant follicle favours pre-embryo development. *Hum Reprod.* 1996;11:1958–62.
- Fukuda M, Fukuda K, Yding Andersen C, Byskov AG. Anovulations in an ovary during two menstrual cycles enhance the pregnancy potential of oocytes matured in that ovary during the following third cycle. *Hum Reprod.* 1999;14:96–100.
- Fukuda M, Fukuda K, Yding Andersen C, Byskov AG. Does anovulation induced by oral contraceptives favor pregnancy during the following two menstrual cycles? *Fertil Steril*. 2000;73:742–7.
- Fukuda M, Fukuda K, Yding Andersen C, Byskov AG. Characteristics of human ovulation in natural cycles correlated to age and achievement of pregnancy. *Hum Reprod.* 2001;16:2501–7.
- Fukuda M, Fukuda K, Yding Andersen C, Byskov AG. Ovulation jumping from the left to the right ovary in two successive cycles may increase the chances of pregnancy during intrauterine insemination and/or in vitro fertilization natural cycles. *Fertil Steril.* 2006;85:514–7.
- 8. Fukuda M, Fukuda K, Tatsumi K, et al. The ovulation pattern during three consecutive menstrual cycles has a significant impact on pregnancy rate and sex of the offspring. *Fertil Steril*. 2011;95:2545–7.
- Fukuda M, Fukuda K, Yding Andersen C, Byskov AG. Right-sided ovulation favours pregnancy more than left-sided ovulation. *Hum Reprod.* 2000;15:1921–6.
- Fukuda M, Fukuda K, Yding Andersen C, Byskov AG. Do basal oestradiol and oestradiol:androgens and oestradiol:FSH ratios reflect pregnancy potential of women receiving intrauterine insemination during natural cycles? *Reprod Biomed Online*. 2003;6:452–5.
- Fukuda M, Shimizu T, Fukuda K, Yomura W, Shimizu S. Transvaginal hysterosonography for differential diagnosis between submucous and intramural myoma. *Gynecol Obstet Invest*. 1993;35:236–9.
- Jordan J, Craig K, Clifton DK, Soules MR. Luteal phase defect: the sensitivity and specificity of diagnostic methods in common clinical use. *Fertil Steril.* 1994;62:54–62.
- Syrop CH, Hammond MG. Diurnal variation in midluteal serum progesterone measurements. *Fertil Steril*. 1987;47:47–67.
- Moore PJ, Greenwald GS. Effect of hypophysectomy and gonadotropin treatment on follicular development and ovulation in the hamster. *Am J Anat.* 1974;139:37–8.
- Mizuno T, Wada H, Nakano R, Tojo S, Washio M. Effects of estrogen and progesterone on follicular growth in hypophysectomized rats. *Folia Endocrinol Jpn.* 1976;52:372, Abstract 213 [in Japanese].
- Hoffmann F. Uber die Wirkung des Progesterons auf das Follikelwachstum im Zyklus und seine Bedeutung für die hormonale Steuerung des Ovarialzyklus der Frau. *Geburtshilfe Frauenheilkd.* 1962;22:433–40.
- Bäckström T, Carlstrom K, von Schoultz B, Toivonen J. Effect of progesterone, administered via intravaginal rings, on serum concentration of oestradiol, FSH, LH and prolactin in women. *J Reprod Fertil.* 1982;64:53–8.
- Peluso JJ, Pappalardo A. Progesterone mediates its anti-mitotic and antiapoptotic actions in rat granulosa cells through a progesterone-binding protein with gamma aminobutyric acid receptor-like features. *Biol Reprod.* 1998;58:1131–7.
- Peluso JJ. Multiplicity of progesterone's actions and receptors in the mammalian ovary. *Biol Reprod.* 2006;75:2–8.
- 20. Akahori T. The effect of follicular maturation and sex steroids upon LH and FSH binding to porcine granulosa cells. *Acta Obstet Gynaecol Jpn*. 1978;30:191–8.
- Tyler JPP, Smith DM, Biggers JD. Effect of steroids on oocyte maturation and atresia in mouse ovarian fragments in vitro. *J Reprod Fertil.* 1980; 58:203–12.

- Schreiber JR, Nakamura K, Erickson GF. Progestins inhibit FSH-stimulated steroidogenesis in cultured rat granulosa cells. *Mol Cell Endocrinol*. 1980; 19:165–73.
- Schreiber JR, Nakamura K, Erickson GF. Progestins inhibit FSH-stimulated granulosaestrogenproductionatapost-cAMPsite. *Mol Cell Endocrinol*. 1981; 21:161–70.
- Fortune JE, Vincent SE. Progesterone inhibits the induction of aromatase activity in rat granulosa cells in vitro. *Biol Reprod.* 1983;28:1078–89.
- Fukuda M, Katayama K, Tojo S. Inhibitory effect of progesterone on follicular growth and induced superovulation in the rat. *Arch Gynecol.* 1980; 230:77–87.
- Goodman AL, Nixon WE, Johnson DK, Hodgen GD. Regulation of folliculogenesis in the cycling rhesus monkey: selection of dominant follicle. *Endocrinology*. 1977;100:155–61.
- Tyndale-Biscoe CH, Hawkins J. The corpora lutea of marsupials: aspects of function and control. In: Calaby JH, Tyndale-Biscoe CH, editors, *Reproduction and Evolution*. Australian Academy of Science, Canberra. 1977:245–52.
- diZerega GS, Hodgen GD. Folliculogenesis in the primate ovarian cycle. Endocrine Reviews. 1981;2:27–49.
- Renfree MB, Wallace GI, Young IR. Effects progesterone, oestradiol-17B and androstenedione on follicular growth after removal of the corpus luteum during lactational and seasonal quiescence in the Tammar Wallaby. *J Endocrinol*. 1982;92:397–403.
- Tsuji K, Sowa M, Nakano R. Relationship among the status of the human oocyte, the 17B-estradiol concentration in the antral fluid and the follicular size. *Endocrinol Japon*. 1983;30:251–4.
- Fukuda M, Katayama K, Tsujimoto D, Akahori T, Tojo S. Inhibitory effect of corpus luteum on follicular growth and induced superovulation in immature rats. *Jpn J Fertil Steril*. 1983;28:45–9.
- Fukuda M, Fukuda K, Yding Andersen C, Byskov AG. Does corpus luteum locally affect follicular growth negatively? *Hum Reprod.* 1997;12:1024–7.

Publish with Libertas Academica and every scientist working in your field can read your article

"I would like to say that this is the most author-friendly editing process I have experienced in over 150 publications. Thank you most sincerely."

"The communication between your staff and me has been terrific. Whenever progress is made with the manuscript, I receive notice. Quite honestly, I've never had such complete communication with a journal."

"LA is different, and hopefully represents a kind of scientific publication machinery that removes the hurdles from free flow of scientific thought."

Your paper will be:

- Available to your entire community free of charge
- Fairly and quickly peer reviewed
- Yours! You retain copyright

http://www.la-press.com