

**STEREOSELECTIVE SYNTHESSES OF 2,5-DISUBSTITUTED
HYDROXYFURAN DERIVATIVES AS SYNTHON OF POLYETHER
ANTIBIOTICS[†]**

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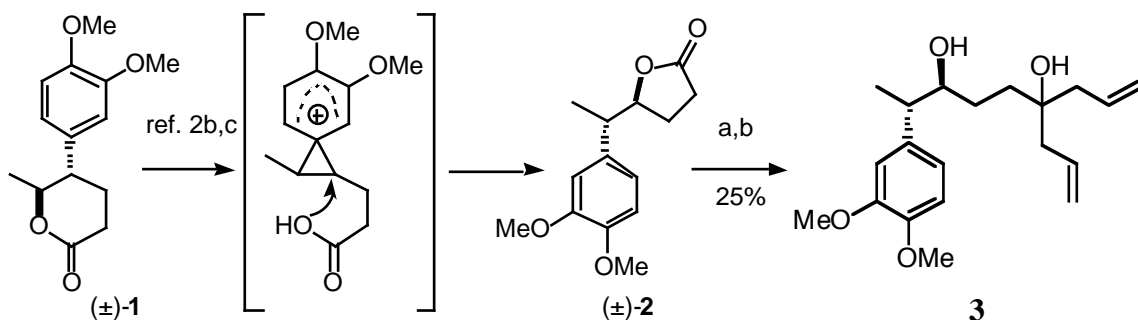
Abstract- Stereoselective syntheses of optically active 2,5-dialkyltetrahydrofurans (**8**), (**11**) and (**12**), which correspond to a central moiety of pamamycin 607, have been achieved on the basis of lactone-ring transformation *via* a phenonium ion, intramolecular Friedel-Crafts reaction, oxidative decomposition of an aromatic ring and differentiation of two ester groups by a chemical or chemo-enzymatic method.

Nature abounds with compounds containing 2,5-disubstituted tetrahydrofurans as polyether antibiotics. Since these compounds are important in many biological processes, there is much interest in stereoselective preparation of such a moiety.¹ Recently, we developed a lactone-ring transformation of α -lactone (\pm)-(**1**) into β -lactone (\pm)-(**2**) *via* a phenonium ion proposed by Cram (Scheme 1).^{2,3} It is noteworthy that the benzylic asymmetric center on the ring can be stereospecifically transferred to a side chain in this ring transformation. The successful conversion stimulated us to carry out stereoselective installation of the second alkyl side chain with a view to developing a new route for synthesis of polyether antibiotics. Our initial attempt to install the second alkyl chain into the 5-membered ring was carried out using Kishi's method,⁴ which was applied to the preparation of *cis*-2,6-disubstituted tetrahydropyran. Alkylation of (\pm)-**2** with allyl Grignard reagent followed by reduction with $\text{Et}_3\text{SiH}/\text{BF}_3 \cdot \text{Et}_2\text{O}$ afforded diol (**3**) in low yield.

Faced with this problem, we decided to use an alternative method for installation of the second side chain. We report here the stereoselective syntheses of optically active hydrofurans (**8**), (**11**) and (**12**), which correspond to a central moiety of pamamycin 607 isolated from *S. alboniger* IFO 12738,^{6,7} *via* lactone-ring transformation, intramolecular Friedel-Crafts reaction and decomposition of an aromatic ring.

[†] This paper is dedicated to Prof. Yuichi Kanaoka in memory of his 75th birthday.

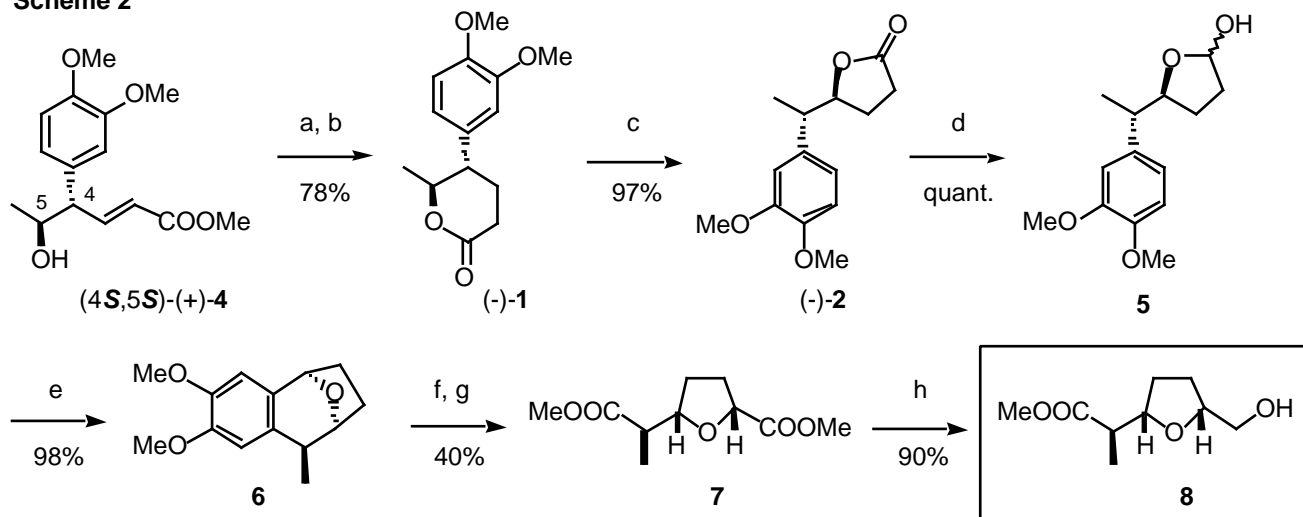
Scheme 1



Reagents: (a) $\text{CH}_2=\text{CHCH}_2\text{MgBr}$, -78°C (b) Et_3SiH , $\text{BF}_3\cdot\text{Et}_2\text{O}$

At first, we prepared optically active α -lactone $(-)$ -**1**⁵ by hydrogenation of $(4S,5S)$ -**4**⁸ and subsequent acidic lactonization. Treatment of $(-)$ -**1** with $\text{TsOH}\cdot\text{H}_2\text{O}$ in MeNO_2 at 70°C gave $(-)$ -**2**⁵ in 97% yield. After reduction of $(-)$ -**2** with DIBALH, treatment of the resulting lactol (**5**)⁵ with $\text{BF}_3\cdot\text{Et}_2\text{O}$ resulted in an intramolecular Friedel-Crafts reaction,⁹ giving a bridgehead tricyclic compound (**6**)⁵ in 98% yield. The structure of **6** was determined by ^1H and ^{13}C NMR and other spectral data. The ^1H NMR spectrum exhibited only two aromatic singlet resonances (δ 6.72, 6.48) and benzyl methine proton resonance (δ 4.95, $J = 5.9$ Hz). A newly generated asymmetric center could be completely controlled by use of the nature of the bridgehead ring. Next, we attempted the oxidative decomposition of the aromatic ring using Sharpless's method.¹⁰ Treatment of **6** with a catalytic amount of RuCl_3 in the presence of NaIO_4 in MeCN , CCl_4 and H_2O followed by methylation with CH_2N_2 produced *cis*-diester (**7**)⁵ in 40% yield (2 steps).

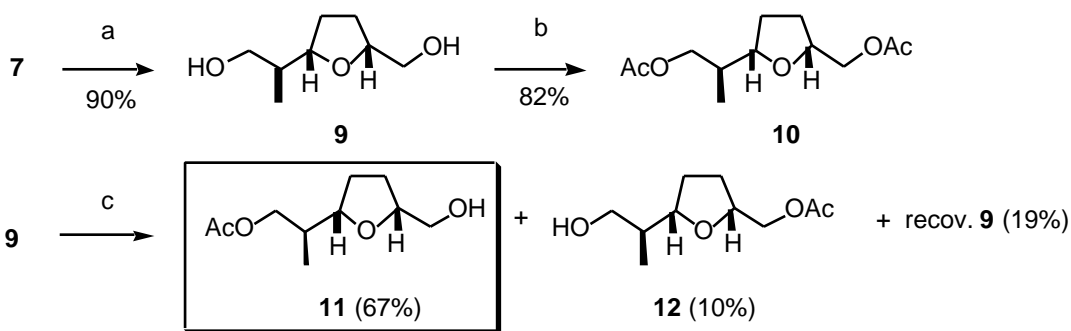
Scheme 2



Reagents: (a) 20% $\text{Pd}(\text{OH})_2\text{-C}$, H_2 (b) 2N HCl (c) $p\text{-TsOH}\cdot\text{H}_2\text{O}$, MeNO_2 , 70°C (d) DIBALH, -78°C (e) $\text{BF}_3\cdot\text{Et}_2\text{O}$, MeCN , 0°C (f) $\text{RuCl}_3\cdot n\text{H}_2\text{O}$, NaIO_4 , MeCN , CCl_4 , H_2O , 0°C (g) CH_2N_2 , Et_2O , 0°C (h) NaBH_4 , MeOH , 0°C

The final step toward our goal was differentiation of the two ester groups of **7**. We used two methods. One of them was a chemical method. When **7** was treated with NaBH_4 in MeOH at 0°C , reduction proceeded regioselectively to give **8**⁵ in 90% yield.¹¹ As an alternative effort, we confirmed the regioselective enzymatic hydrolysis^{12,13} of diacetate (**10**). Dimethyl ester (**7**) was converted into **10**⁵ by LAH reduction of **7** followed by conventional acetylation. When **10** was treated with lipase Amamo PS (*Pseudomonas* sp.) in phosphate buffer at 33°C for 4 h, hydrolysis proceeded with fairly good regioselectivity to give monoacetate (**11**)⁵ in 67% yield along with **12**⁵ (10%). Fortunately, these compounds could be separated by column chromatography with silica gel. Compounds (**8**), (**11**) and (**12**) can be used in the synthetic studies of pamamycin 607.

Scheme 3



Reagents: (a) LiAlH_4 , THF, 0°C (b) Ac_2O , DMAP, Py, 0°C (c) lipase Amamo PS, 0.1 M phosphate buffer, 33°C , 4 h

In conclusion, we synthesized **8**, **11** and **12** based on lactone-ring transformation *via* a phenonium ion, intramolecular Friedel-Crafts reaction, oxidative decomposition of an aromatic ring and differentiation of diester by a chemical or chemo-enzymatic method. Further attempts to convert of the compounds into pamamycin 607 are in progress.

ACKNOWLEDGEMENTS

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- 5 All new compounds were identified by spectroscopic data. For representative compounds, **6** (white needle): mp 89-91°C (AcOEt-hexane); $[\alpha]_D^{20} -13.4^\circ$ ($c = 1.03$, CHCl_3); EI-MS m/z 234 (M^+), 205; HR-MS m/z 234.1284 (Calcd for $\text{C}_{14}\text{H}_{18}\text{O}_3$: 234.1255); $^1\text{H-NMR}$ (270 MHz, CDCl_3) 6.72 (1H, s), 6.48 (1H, s), 4.95 (1H, d, $J = 5.9$ Hz), 4.51-4.43 (1H, m), 3.85 (3H, s), 3.84 (3H, s), 3.45-3.32 (1H, m), 2.20-2.03 (1H, m), 1.94-1.83 (3H, m), 1.20 (3H, d, $J = 7.3$ Hz); $^{13}\text{C-NMR}$ (68 MHz, CDCl_3) 148.07 (s), 147.00 (s), 132.23 (s), 128.92 (s), 110.12 (d), 107.24 (d), 79.09 (d), 77.15 (d), 55.97 (q), 55.92 (q), 36.74 (d), 35.69 (t), 22.34 (t), 14.78 (q). **7** (colorless oil): $[\alpha]_D^{20} -25.1^\circ$ ($c = 1.01$, CHCl_3); IR (neat) 1738 cm^{-1} ; EI-MS m/z 185 ($\text{M}^+ - \text{OCH}_3$), 157; HR-MS m/z 185.0807 (Calcd for $\text{C}_9\text{H}_{13}\text{O}_4$ ($\text{M}^+ - \text{OCH}_3$): 185.0813); $^1\text{H-NMR}$ (270 MHz, CDCl_3) 4.47 (1H, dd, $J = 8.2, 4.6$ Hz), 4.11 (1H, dt, $J = 8.2, 6.3$ Hz), 3.73 (3H, s), 3.69 (3H, s), 2.72 (1H, quintet, $J = 7.0$ Hz), 2.32-2.15 (1H, m), 2.14-1.97 (2H, m), 1.83-1.66 (1H, m), 1.32 (1H, d, $J = 7.0$ Hz); $^{13}\text{C-NMR}$ (68 MHz, CDCl_3) 174.40 (s), 172.96 (s), 81.83 (d), 76.58 (d), 51.46 (q), 51.13 (q), 44.28 (d), 29.79 (t), 28.56 (t), 13.98 (q). **8** (colorless oil): $[\alpha]_D^{20} -36.5^\circ$ ($c = 1.00$, CHCl_3); IR (neat) $3454, 1738\text{ cm}^{-1}$; EI-MS m/z 157 ($\text{M}^+ - \text{CH}_2\text{OH}$), 125; HR-MS m/z 157.0860 (Calcd for $\text{C}_8\text{H}_{13}\text{O}_3$ ($\text{M}^+ - \text{CH}_2\text{OH}$): 157.0864); $^1\text{H-NMR}$ (270 MHz, CDCl_3) 4.10-3.98 (2H, m), 3.79-3.67 (1H, m), 3.69 (3H, s), 3.45 (1H, dd, $J = 11.5, 5.5$ Hz), 2.65 (1H, quintet, $J = 6.9$ Hz), 2.25-1.65 (5H, m), 1.23 (3H, d, $J = 6.9$ Hz); $^{13}\text{C-NMR}$ (68 MHz, CDCl_3) 174.92 (s), 80.69 (d), 79.59 (d), 64.91 (t), 51.63 (q), 44.12 (d), 28.74 (t), 26.82 (t), 13.65 (q). **11** (colorless oil): $[\alpha]_D^{20} -4.9^\circ$ ($c = 1.27$, CHCl_3); IR (neat) $3470, 1734\text{ cm}^{-1}$; EI-MS m/z 171 ($\text{M}^+ - \text{CH}_2\text{OH}$), 149; HR-MS m/z 171.1028 (Calcd for $\text{C}_9\text{H}_{15}\text{O}_3$ ($\text{M}^+ - \text{CH}_2\text{OH}$): 171.1020); $^1\text{H-NMR}$ (270 MHz, CDCl_3) 4.07 (1H, dd, $J = 11.2, 6.3$ Hz), 3.98 (1H, dd, $J = 11.2, 6.6$ Hz), 4.05-3.94 (1H, m), 3.88-3.78 (1H, m), 3.70 (1H, br d, $J = 11.2$ Hz), 3.48 (1H, dd, $J = 11.2, 5.9$ Hz), 2.06 (3H, s), 2.05-1.87 (3H, m), 1.77-1.56 (3H, m), 1.01 (3H, d, $J = 6.9$ Hz).

¹³C-NMR (68 MHz, CDCl₃) 171.12 (s), 80.97 (d), 78.98 (d), 66.78 (t), 65.20 (t), 37.18 (d), 28.79 (t), 27.12 (t), 20.94 (q), 13.01 (q). **12** (colorless oil): [α]_D²⁰ -11.4° (c = 1.03, CHCl₃); IR (neat) 3430, 1744 cm⁻¹; EI-MS *m/z* 143 (M⁺ - CH(CH₃)CH₂OH), 129; HR-MS *m/z* 143.0709 (Calcd for C₇H₁₁O₃ (M⁺ - CH(CH₃)CH₂OH): 143.0708); ¹H-NMR (270 MHz, CDCl₃) 4.17 (1H, dd, *J* = 10.6, 3.0 Hz), 4.15-3.99 (2H, m), 4.02 (1H, dd, *J* = 10.6, 5.6 Hz), 3.69 (1H, dd, *J* = 10.9, 7.3 Hz), 3.56 (1H, dd, *J* = 10.9, 4.3 Hz), 2.54 (1H, br), 2.13-1.60 (5H, m), 2.08 (3H, s), 0.91 (3H, d, *J* = 7.3 Hz); ¹³C-NMR (68 MHz, CDCl₃) 170.99 (s), 83.36 (d), 76.45 (d), 66.13 (t), 65.86 (t), 37.97 (d), 27.68 (t), 26.68 (t), 20.84 (q), 12.09 (q).

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