PERSPECTIVES

RANKL: Targeting Bone and Cancer to Treat Skeletal Complications of Malignancy

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Abstract

The RANKL signaling pathway is a key mediator of osteoclastic bone resorption in response to stimuli from cancer cells. Bone resorption releases growth factors that then further stimulate cancer growth in a "vicious cycle". RANKL inhibition effectively decreases solid tumor bone metastases, myeloma and hypercalcemia of malignancy in animal studies. In clinical trials, humanized RANKL antibody therapy is effective in all of these settings and in cancer treatment-related bone loss as well. Although there are theoretical concerns that inhibition of RANKL signaling could impact the immune system or increase the incidence of neoplasms, neither of these adverse outcomes have been realized clinically to date. Emerging research suggests that RANKL signaling plays additional roles in cancer cells independent of osteoclasts by increasing homing of cancer cells to bone and invasive potential. Osteoclast eradication caused by RANKL inhibition may also offer advantages in suppression of bone turnover compared to bisphosphonates. Thus, targeting RANKL in skeletal complications of malignancy potently suppresses bone turnover to improve skeletal mortality in those conditions, and may have direct antitumor effects as well. *IBMS BoneKEy*. 2009 September;6(9):323-338.

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Background

Bone is a fertile environment for cancer growth as initially described by the seed and soil hypothesis proposed by Stephen Paget in 1889. Over 100 years later, this theory is unchanged but has evolved to include a greater understanding of the interaction between cancer cells and the cells of the bone microenvironment that results in a "vicious cycle" of cancer growth. Cancer cells hijack osteoblasts and osteoclasts to disrupt normal bone remodeling. Increased bone resorption releases growth factors from the bone matrix that further stimulate cancer cell growth. The balance of resorption and formation determines the radiographic phenotype of osteolytic and osteoblastic bone metastases. Accumulating evidence. clinical and experimental, supports a pivotal role of osteoclasts in the pathogenesis and sequelae of bone metastases regardless of the radiographic phenotype. Bone resorption markers are increased in both osteolytic and osteoblastic solid tumor bone metastases and are higher in the latter (1).

Bisphosphonates decrease skeletal morbidity from bone metastases from solid tumors or myeloma whether osteolytic or osteoblastic (2;3). Finally, the bone resorption marker, N-telopeptide, predicts death and skeletal morbidity from solid tumors and this relationship is stronger in osteoblastic disease due to prostate cancer (4;5). Thus osteoclasts play a pivotal role in bone metastases and their sequelae, and the rationale to target this bone resorbing cell is strong.

Cancer affects the skeleton in several ways: bone metastases, hypercalcemia and osteoporosis due to cancer treatment. Skeletal-related clinical events (SREs) are a direct result of bone metastasis and include bone pain, pathologic fractures and spinal cord compression. SREs contribute significantly to patient morbidity as up to 75% of advanced prostate and breast cancer and 30-40% of lung cancer patients have bone metastases (6). Systemic effects of tumor-derived PTHrP, which promotes calcium mobilization from bone and renal tubular reabsorbtion of calcium, is a major

factor hypercalcemia of causal in malignancy. Multiple myeloma also causes significant osteoclastogenesis and purely lytic lesions resulting in both hypercalcemia and SREs. Finally, cancer treatments such estrogen or androgen blockade, as chemotherapy and radiation therapy result in bone loss. Increased bone resorption by osteoclasts during cancer treatment releases a rich store of growth factors from the mineralized bone matrix and can enhance cancer growth in bone. Inhibiting osteoclast function to reduce bone resorption is paramount in each of these reduce these scenarios to related complications and possibly tumor growth in bone.

Bisphosphonates currently represent the only bone-targeted and approved therapy to treat skeletal complications of cancer and cancer treatment. They effectively inhibit osteoclast function, and decrease the risks of SREs in patients with solid tumor bone metastases. Bisphosphonates are also effective for treatment of hypercalcemia of malignancy, multiple myeloma-related bone resorption and bone loss during cancer treatment. Despite this effectiveness. bisphosphonates do not cause regression of bone metastases, so new therapies are needed. There are several limitations to the use of bisphosphonates: 1) association with osteonecrosis of the jaw, recognized recently with potent IV bisphosphonates. especially in cancer patients: 2) long half-life in bone and long-term potential to suppress bone turnover; 3) possible renal toxicity and contraindication in renal failure: 4) poor oral bioavailability; 5) failure to suppress markers of bone resorption in some patients with myeloma or solid tumor bone metastases (7;8). Thus, better options to target the osteoclast are needed.

An attractive target is the RANKL signaling triad that plays a dominant role in osteoclastogenesis. RANKL, when bound to RANK on the surface of osteoclast precursors, promotes osteoclastogenesis, whereas OPG, a soluble decoy receptor for RANKL, inhibits osteoclast formation. OPG was the first to be identified in this signaling pathway. It caused osteopetrosis in mice when overexpressed and protected mice from ovariectomy-induced bone loss (9). At the same time, another group purified OPG from fibroblasts and used it to significantly inhibit osteoclastogenesis (10). RANKL was then identified as the ligand for OPG and treated mice developed hypercalcemia from increased osteoclastogenesis (11). Further demonstrating a requirement of RANKL signaling for osteoclast development, both RANK and RANKL null transgenic mice lack osteoclasts (12;13).

OPG and RANKL are both expressed by osteoblasts and bone marrow stromal cells and the ratio of these determine the degree of osteoclastogenesis, effectively balancing new bone formation and resorption in healthy bone. In cancer the RANKL/OPG ratio is perturbed by signals from cancer cells and excessive bone resorption or formation occurs. PTHrP, IL-6 and IL-11 are secreted from cancer cells and increase RANKL expression in osteoblasts (14-16). IL-8 increases osteoclastogenesis in both a RANKL-dependent and -independent manner (17). Recent data in osteoarthritis models demonstrate increased RANKL expression in osteoblasts treated with IL-1 β . TNF α , PGE2 and IL-17 as well and these factors could potentially play a role in cancer and bone interactions (18). In addition to enhancing bone resorption in response to cancer cell signals, RANKL, RANK and OPG may also play a role in cancer cell proliferation, migration to and invasion of bone.

Clinical and Experimental Evidence in Cancer

There is accumulating evidence that cancer cells express all components of the RANKL signaling pathway, but the exact function of this pathway may be different in different types of cancers. The relative role of this pathway in cancer cells and the relationship to bone cells is under intense investigation.

Evidence for RANKL: Osteoclastdependent effects

Cancer cells produce many factors that stimulate osteoblast production of RANKL and subsequent osteoclastic bone resorption. These effects are the most recognized role for RANKL in cancer.

Breast Cancer

Metastatic bone lesions are predominately osteolytic in breast cancer. Tumor induction of osteoblast and stromal cell RANKL expression plays a predominant role in osteoclastogenesis and bone destruction. It is possible that breast cancer cells activate osteoclasts directly, but this is likely a small contribution. Only 14% of breast cancers express RANKL (19), and most in vitro systems require osteoblasts to be cultured with breast cancer cells to induce osteoclastogenesis. When MDA-MB-231 breast cancer cells were co-cultured with calvarial osteoblasts, they induced bone resorption and RANKL mRNA expression in osteoblasts whereas MDA-MB-231 cells alone did not express RANKL (20;21). Consistent with this, MDA-MB-231, MCF-7 and T47D breast cancer cells did not express RANKL, but notably did express RANK and OPG. When these cells were cocultured with murine osteoblasts and hematopoietic cells, they induced osteoclast formation. This process PTHrPwas dependent, and bone destruction and RANKL expression by osteoblasts was amplified in PTHrP-overexpressing breast cancer (14). This demonstrates the ability of tumor cells to induce RANKL expression in osteoblasts and stromal cells to indirectly increase osteoclast formation. Since many breast cancers express RANK (14), it also introduces the possibility that RANK expression by cancer cells may play a role in cancer growth in bone.

Myeloma

The vast majority of multiple myeloma cases are characterized by marked osteolysis with suppressed bone formation. Significant bone destruction occurs through alterations in the RANKL signaling pathway. Osteolytic lesions are identified adjacent to nests of myeloma cells and these can result in hypercalcemia and SREs. An increased RANKL/OPG ratio correlated with increased tumor burden in myeloma (22). OPG concentrations were lower in patients with myeloma compared to healthy controls (23).

High RANKL/OPG ratios also predicted poor survival in patients with myeloma (24). Both myeloma cells isolated from patients and established myeloma cell lines express RANKL and can induce osteoclastogenesis directly (25-30). This is evidenced by data showing that myeloma cells and conditioned media are capable of inducing osteoclastogenesis in bone marrow mononuclear populations, bypassing a requirement for osteoblasts or stromal cells (29). Myeloma cells also upregulate RANKL expression in bone marrow stromal cells and osteoblasts (30:31) and in T lymphocytes (32). RANKL expressed by myeloma cells or induced in other cells in the bone microenvironment plays an important role in pathogenesis of myeloma the bone destruction. Since myeloma also suppresses osteoblastogenesis, myeloma expression of RANKL provides an additional route to stimulate osteoclastogenesis and may explain why it remains the malignancy so associated commonly with profound osteolvsis.

Prostate cancer

Prostate cancer bone metastases are predominately osteoblastic, but osteoclastic bone resorption is also a prominent feature in osteoblastic disease and likely contributes to tumor growth in bone. Whereas the RANKL/OPG ratio is increased in predominately osteolytic bone lesions of breast cancer and myeloma, OPG is relatively increased in human prostate cancers. The contribution of this increased OPG to osteoclastic bone resorption or stimulation of bone formation is unclear. Human prostate cancers express all three components of the RANKL signaling triad, RANKL, OPG and RANK, and the expression of these correlated with advanced cancer stage and cancer metastasis (33). Furthermore, the highest concentration of OPG was observed in prostate cancer bone lesions. Consistent with these findings, both OPG and RANKL significantly were increased bv immunohistochemistry in bone metastases compared to paired primary prostate cancers (34). Finally, RANKL expression on the surface of prostate cancer cells predicted recurrence of disease with a hazard ratio of 11.6 (35). Different components of the RANKL signaling pathway are expressed in breast cancer, myeloma and prostate cancer and may have tumor-type specific roles. These tumorspecific roles have yet to be defined.

Hypercalcemia of malignancy

There are no published data on the direct alteration of RANKL signaling in hypercalcemia of malignancy, but there is abundant evidence that tumor products such as PTHrP that mediate hypercalcemia stimulate RANKL. Inhibition of RANKL effectively treats hypercalcemia of malignancy, reviewed later, and provides indirect evidence for a role of the RANKL pathway in the pathogenesis of hypercalcemia associated with malignancy.

Evidence for RANKL: Osteoclastindependent effects

Recent data suggest that the RANKL pathway may have important roles in various aspects of skeletal complications in cancer that do not involve the osteoclast.

Homing of cancer cells to bone

RANKL is clearly important for bone resorption and fueling cancer growth. However, there are data to suggest that bone-derived RANKL may also serve as "soil" or a chemoattractant to bone for RANK-expressing cancer cells independent of bone resorption and osteoclast activity. In normal mammary epithelium, melanoma and breast cancer cells, RANKL induced invasion and migration in vitro. These effects were blocked by OPG, and were not observed in a RANK-negative colon cancer cell line. In a melanoma metastases model that was previously shown not to activate osteoclasts and bone resorption, OPG did decrease bone metastases. Because OPG was effective in a system thought to be devoid of osteoclast activation, the authors concluded that the decrease in bone metastases was caused by inhibition of RANKL-mediated homing of cancer cells to bone and not by increased bone turnover (36). This conclusion is controversial as this mouse melanoma model may have some

degree of osteoclast activation and increased bone turnover, not recognized by the authors. An intratibial inoculation model of these melanoma cells may shed some light on the role of RANKL in cancer cell migration to bone. In this model, there would be no cancer cell migration and OPG efficacy would implicate targets of osteoclast-induced bone resorption or direct anti-tumor effects.

Epithelial to mesenchymal transition (EMT) and invasion

EMT and invasion important are determinants of cancer growth in healthy tissue. Functional RANKL expression by prostate cancer cells is correlated with EMT and a threefold increase in bone metastases (37), supporting another tumor-specific role for RANKL signaling in cancer progression. In PC3 prostate cancer cells, treatment with RANKL increased invasion of collagen matrix and increased expression of MMP-9 and interleukin 6 and these effects were blocked with OPG (38). Consistent with these results, RANKL induced invasion of breast cancer cells in a matrigel invasion assay system (39). The role of RANKL as a chemoattractant or transformant independent of osteoclasts has yet to be established in vivo, but in vitro studies are suggestive.

Apoptosis

Components of the RANKL signaling pathway may have additional effects on tumor cells. There is evidence that blocking RANKL in certain settings with OPG could actually stimulate tumor growth by binding the pro-apoptotic factor TRAIL. Expression of OPG by MDA-MB-231 cells increased tumor inhibition of TRAIL and improved tumor survival (40). OPG over-expression in MCF-7 breast cancer cells also enhanced tumor growth in bone (41) and OPG expression by breast cancer cells correlated with bone homing and colonization potential (42). Recent data also suggest that interaction between TRAIL and OPG could increase RANKL expression in MDA-MB-231 cells (43). These data are in contrast to the inhibition of breast cancer tumor growth in bone by systemic treatment with OPG in mice (44;45) and may be due to the antiapoptotic or perhaps autocrine effects of OPG when produced directly by tumor cells. Although there are concerns that systemic OPG therapy could enhance tumor cell survival, OPG treatment consistently decreases bone metastasis in murine models of breast cancer metastasis.

Angiogenesis

OPG expression in endothelium of malignant breast tumors correlated with higher tumor grade and appears to have proangiogenic effects *in vitro*. OPG was expressed in the endothelium of 59% of malignant breast cancers, but not expressed in endothelium of nonmalignant tissue. OPG also supported endothelial cell survival *in vitro* and promoted cord structures in a matrigel tubule formation assay (46). OPG may decrease apoptosis of both tumor cells and endothelium, simultaneously increasing tumor growth and necessary blood supply.

Inhibition of RANKL: Animal Studies

Inhibition of RANKL *in vivo* is very effective in reducing bone destruction in murine models of breast cancer, myeloma and prostate cancer. In mice, OPG-Fc and RANK-Fc, recombinant chimeric human proteins, have been developed to study this. The fully humanized anti-RANKL antibody (huRANKL MAb), denosumab, does not recognize murine RANKL and is ineffective in mice. Therefore, these *in vivo* studies are a close approximation of huRANKL MAb treatment in humans (Table 1).

Breast cancer

In murine models of breast cancer, OPG-Fc prevented the development of osteolytic bone lesions (44;47) and treatment with OPG-Fc after establishment of osteolytic bone metastases resulted in a significant decrease in bone destruction and lesion growth (44;45;48). However, *in vitro* treatment with OPG did not affect tumor cells suggesting that the antiresorptive effects of OPG *in vivo* are necessary for inhibition of tumor growth (45). Both preventative and treatment models have resulted in improved survival in mice, though

there was no difference in soft tissue tumor burden. OPG caused tumor cell apoptosis in vivo, but not in vitro, once again supporting osteoclast-mediated effects (44). There was increased tumor cell apoptosis in mice treated with OPG-Fc compared to those treated with zoledronic acid. This may be due to the eradication of osteoclasts by OPG-Fc but not by zoledronic acid (49). A study comparing ibandronate and OPG-Fc in mice with breast cancer bone metastases showed similar inhibition of bone destruction, and tumor growth in bone, and once again OPG-Fc treatment resulted in a total absence of osteoclasts (45).

Myeloma

Inhibition of RANKL in murine models of myeloma yields similar results to breast cancer studies. Although OPG and RANK do not have direct effects on multiple myeloma cells *in vitro*, both OPG-Fc and RANK-Fc treatment decrease melanoma burden and bone destruction (31;50-52), with comparable efficacy to zoledronic acid (53).

Prostate cancer

In prostate cancer, both osteoblastic and osteolytic lesions respond to RANKL inhibition. RANK-Fc and OPG-Fc treatment reduced both lesion area and tumor establishment in bone of osteolytic predominate prostate cancer cell lines (54-56). In contrast, osteoblastic lesions became established despite preventative RANKL inhibition, but treatment with RANKL inhibitors decreased growth of established lesions. (55;57). These findings were specific to tumor growth in bone as there was no impact on subcutaneous tumors. These data suggest that establishment of lesions osteoblastic is osteoclastindependent, but that ongoing growth is dependent on bone destruction. Consistent with these findings, OPG-Fc did not inhibit establishment of osteoblastic bone lesions, but did decrease tumor cell growth and PSA (58). As prostate cancer cells often express RANKL, it is important to determine the role of bone-derived and tumor-expressed RANKL. In C4-2B human prostate cancer cells, huRANKL MAb did not inhibit growth

Solid tumor	Agent	Results	Reference
metastases	used		
Breast	OPG-Fc	↓ tumor burden, absent OC with IC MDA-MB-231	(48)
	OPG-Fc	tumor burden and bone destruction of IC MDA-MB-	(47)
		231	()
	OPG-Ec	tumor burden and bone destruction of IT MDA-MB-	(45)
	0.0.0	231 comparable to IBN OC eradicated with OPG	()
		only	
	OPG-Ec	tumor burden and bone destruction 1 survival of	(44)
	01010	$\Gamma_{\rm W}$ (affect barden and bone destruction, Γ survival of $\Gamma_{\rm W}$	()
	Adenoviral	\downarrow tumor burden of IC MDA MB 435 \leftrightarrow visceral mets	(73)
			(73)
Dreatato		tumor growth of IT is tumor of SC C4 2P	(57)
FIUSIALE		↓ tumor growth of 11, ⇔ tumor of 30 04-2B	(57)
	OPG-FC	\downarrow tumor growth \Leftrightarrow tumor establishment of 11 LuCaP	(58)
	0505		(50)
	OPG-FC	\downarrow tumor burden and bone destruction of 11 C4-2,	(56)
		comparable to ZA	
	RANK-Fc	\downarrow tumor growth \Leftrightarrow tumor establishment of IT LAPC-9	(55)
	Hu	↔ tumor or bone destruction of IT C4-2	(59)
	RANKL		
	MAb		
	OPG-Fc	↓ tumor burden of IT PC3	(38)
	RANK-Fc	↓ tumor burden of IT PC3	(61)
	+/-	↑ BV/TV	
	Docetaxel		
	OPG-Fc	↓ tumor burden of IC PC3	(60)
	+/-	↑ survival	
	Docetaxel		
Colon	OPG-Fc	↓ tumor burden of IC colon-26	(48)
Lung	RANK-Fc	↓ tumor burden and bone destruction of IT A549	(62)
Melanoma	OPG-Fc	↓ tumor burden, ↔ visceral mets, ↑ survival with IC	(36)
		B16F10	× ,
Myeloma	RANK-Fc	I myeloma burden and bone destruction with huMM.	(31)
		ARH-77	(-)
	OPG-Ec	Losteolysis with IV 5T2MM	(52)
	RANK-FC	weloma burden and bone destruction with buMM	(52)
		comparable to BP	(00)
		L myoloma hurdon. M protoin	(50)
	OFG-FC		(50)
1 11 18 4			(0.4)
ннм	muRANK-	↓ bone destruction and serum calcium	(64)
		L have destruction and some solutions must ff ((02)
	UPG-FC	bone destruction and serum calcium, greater effect	(63)
<u> </u>			(0.5)
Bone pain	OPG-Fc	↓ bone destruction and movement-evoked pain of	(65)
	1	established osteosarcoma	

Table 1. Animal studies: RANKL inhibition in solid tumor bone metastases, myeloma and SREs.

Breast cancer: MDA-MB-231, MDA-MB-245; Prostate cancer: C4-2, LuCaP, LAPC, PC3; Myeloma: huMM, ARH-77, 5T2MM, 5T33MM. ↔ no change, ↑ increased, ↓ decreased. IT: intratibial; IC: intracardiac; OC: osteoclasts; SC: subcutaneous; IV: intravenous; ZA: zoledronic acid; IBN: ibandronate; BP: bisphosphonate; HHM: humoral hypercalcemia of malignancy.

of tumors in a murine tibial injection model suggesting that the host or bone derived RANKL was critical for tumor growth (59). RANKL inhibition in combination with chemotherapy also improves outcomes. When given with docetaxel, RANK-Fc decreased bone destruction in mice and OPG-Fc improved survival in mice after intracardiac inoculation of PC3 cells (60;61). Inhibition of RANKL decreases tumor

burden in bone and bone destruction in both blastic and lytic bone metastases and may improve survival in mice with solid tumor metastases.

Lung cancer

Mixed blastic and lytic solid tumor bone metastases from a lung cancer cell line, A549, were significantly reduced with RANK-Fc treatment (62). Treatment with RANK-Fc reduced bone metastases and not subcutaneous tumors. These data are consistent with results for both breast and prostate bone metastases and highlight the importance of osteoclastic bone resorption in not only lytic bone metastases, but in blastic bone metastases as well.

Hypercalcemia of malignancy

RANKL inhibition is also effective in models of hypercalcemia of malignancy. In mice with subcutaneous syngeneic colon cancer C-26, a high-PTHrP-expressing cancer cell line, OPG-Fc treatment reduced serum calcium levels significantly more than either zoledronic acid or pamidronate (63). Although there was complete eradication of osteoclasts, there was a gradual increase of serum calcium at the end of the study period that the authors attributed to host immune responses to the subcutaneous delivery of a human protein and a waning of OPG-Fc However, murine RANK-Fc efficacy. treatment showed a similar rise of serum calcium when tested in mice inoculated with lung cancer that was attributed to unabated subcutaneous tumor growth and increasing PTHrP serum concentrations (64). The increasing PTHrP may lead to decreased renal clearance of calcium or increased 1,25 OH₃ vitamin D and intestinal calcium absorption. Consistent with the previous osteoclasts study, were significantly reduced, and the treatment was effective even when started after hypercalcemia had already developed.

Bone pain

Experimental evidence in mice indicates that OPG is also effective in the management of advanced cancer bone pain. In an osteolytic sarcoma model, OPG treatment decreased movement-evoked pain and other pain behaviors when given after significant bone destruction had already occurred (65). There was also decreased tumor growth and neurochemical change at the spinal cord with OPG treatment. RANKL inhibition, due to potent anti-osteoclast activity, is effective for management of both bone pain and hypercalcemia of malignancy.

Clinical Trials

Solid tumor bone metastases and myeloma

Clinical trials with denosumab mirror the data gleaned from in vivo studies with RANK-Fc and OPG-Fc (Table 2). RANKL MAb effectively reduced bone resorption in patients with solid tumor bone metastasis and myeloma in phase II clinical trials. A dosing regimen of 120 mg subcutaneously every four weeks in patients with myeloma or bone metastases from breast cancer rapidly suppressed urine N-telopeptide (uNTx), an effect that was sustained for 13 weeks. In these patients, 74% of patients receiving RANKL MAb had > 65% suppression of uNTx compared to 63% of the bisphosphonate-treated patients (66). SREs were similar between the two groups, 12% of RANKL MAb-treated affecting 16% patients compared to of bisphosphonate-treated patients at 25 weeks (67). However, a wide range of RANKL MAb dosing regimens were used and this may have diluted the effect of the higher dose. The dosing regimen of 120 mg every 4 weeks was the most effective and is now being evaluated in phase III trials. Superiority of denosumab to zoledronic acid was announced recently. Denosumab delayed time to first SRE compared to zoledronic acid with a hazard ratio of 0.82 (95% CI: 0.71-0.95) in 2,049 patients with advanced breast cancer.

RANKL MAb eliminated osteoclast activity in patients who continued to have elevated markers of bone resorption despite intravenous bisphosphonates. In 111 patients with myeloma or bone metastases from breast and prostate cancer fitting these criteria, RANKL MAb suppressed uNTx < 50 nM BCE/mM creatinine in 71% of patients

Cancer studied	N receiving agent	Results	Reference
Myeloma	N = 25	↓ markers of bone resorption	(78)
Breast	N = 29		
Breast	N = 211	↓ markers of bone resorption and	(67)
		SREs	
Myeloma	N = 85	↓ serum M-protein	(68)
Prostate	N = 50	↓ markers of bone resorption and	(8)
Breast	N = 46	SREs	
Myeloma/Other	N = 15		
Giant cell tumor	N = 24	↓ tumor size	(69)
Treatment-induced bone	N = 127	↑ BMD at lumbar spine, total hip and	(74)
loss: Aromatase inhibitor		distal radius	
for nonmetastatic breast			
cancer			

 Table 2. Clinical trials with huRANKL MAb (denosumab)

compared to 29% of patients receiving zoledronic acid (8). In this study, SREs were significantly reduced with RANKL MAb therapy with an incidence of 8% in the RANKL MAb group compared to 17% in the zoledronic acid group. In a separate evaluation of prostate cancer patients in this cohort, 69% had suppression of uNTx < 50 nM BCE/mM creatinine with RANKL MAb treatment compared to 38% of patients receiving zoledronic acid, suggesting that RANKL MAb is effective for both lytic and blastic bone metastases (7). RANKL MAb also stabilized myeloma as measured by serum M protein levels and decreased markers of bone resorption (68). The stable M protein suggests an inhibitory effect of RANKL MAb on myeloma progression. In several studies of a murine model of solid tumor bone metastases or myeloma, RANKL inhibition increased survival (36;44;50;60). Future studies with adequate power and duration will determine whether denosumab improves survival in humans with solid tumor bone metastases.

Other tumors: Giant cell tumors

Giant cell tumors of bone have also been treated effectively with RANKL MAb. RANKL inhibition decreased tumor volume or improved histologic grade in 87% of patients. Dramatic changes could be seen histologically and a tumor response required elimination of greater than 90% of giant cells. This resulted in substantial clinical benefit as measured by decreased pain, improved movement or return to work (69).

Hypercalcemia of Malignancy

Treatment of hypercalcemia of malignancy with RANKL MAb has not been evaluated in a clinical trial at this point, but *in vivo* studies suggest that it would be effective for this as well.

RANKL Inhibition for Cancer Treatment-Related Bone Loss

Estrogen and androgen blockade, common therapies for both breast and prostate cancer, increase bone turnover, cause bone loss and increase fracture risk. This makes bone a more attractive environment for cancer cell growth. In postmenopausal woman, RANKL inhibition increased bone mineral density at the lumbar spine, total hip and distal radius significantly more than placebo. These improvements were similar to or greater than changes observed in the alendronate arm (70). RANKL inhibition has also been studied in ovariectomy and orchiectomy models of bone loss and is very effective in suppressing bone resorption. Compared to alendronate, OPG treatment caused a greater increase in femur mechanical strength, femur bone mineral density and vertebral trabecular bone volume in ovariectomized mice. There was also a greater decrease in osteoclast number with OPG therapy compared to alendronate (71). OPG in orchiectomy models also preserves bone mass (72). In addition to preserving bone in these models, OPG prevented trabecular bone loss in tumor-bearing mice (59;73), and in mice receiving docetaxel (61). In clinical trials

RANKL MAb increased bone mineral density at the lumbar spine by 5.5% and 7.6% at 12 and 24 months, respectively, compared to placebo in women with nonmetastatic breast cancer treated with aromatase inhibitors (74). Inhibition of RANKL in settings of low estrogen or during cancer treatment effectively inhibits bone resorption, making bone a less fertile environment for cancer cell growth.

Adverse Effects

Determining adverse effects of RANKL MAb has relied on clinical studies. In mice, intact RANKL signaling is required for normal immune system development and absence of either RANKL or RANK causes lymph node agenesis and lymphocyte dysfunction, which was recently reviewed (75). RANK is expressed on the surface of T-cells and dendritic cells and inhibition of RANKL theoretically poses a risk of immune dysregulation. However, inhibition of RANKL after embryogenesis does not disrupt the immune system. Mice treated with OPG-Fc and RANK-Fc had a significant reduction of osteoclasts, but were still able to mount a normal immune response to influenza (76). Clinical trials do not show clinically immune disruption significant with denosumab. In 49 patients treated with a single dose of RANKL MAb, there were no decreases in lymphocyte number (77). After 2 years of RANKL MAb treatment in 319 patients, there was a small but significant increase in urinary tract infections and six cases of community-acquired infections that did not reach statistical significance and were easily treated with standard antibiotics. Acute phase reactions with myalgias and pyrexia were similar in the RANKL MAb and placebo groups. Other adverse effects may include hypertension that was more frequent in patients receiving MAb or alendronate either RANKL compared to placebo (70). RANKL MAb did not increase the incidence of neoplasm in this study, making potential interaction between RANKL inhibition and TRAIL less concernina. To date. no cases of osteonecrosis of the jaw have been reported with RANKL MAb therapy, but more frequent dosing schedules have been evaluated in only a limited number of patients. In 197

patients, there were transient and asymptomatic decreases in serum calcium at 2 weeks in 8% of RANKL MAb patients compared to 5% of patients receiving bisphosphonates. Finally several studies have documented absence of antidenosumab antibodies with therapy (66;78). RANKL MAb is well-tolerated by patients and has not resulted in serious adverse events.

Summary

RANKL plays a major role in solid tumor metastases. myeloma, cancer bone treatment-induced osteoporosis and hypercalcemia of malignancy and the rationale to target RANKL in these disorders is strong. Fig. 1 summarizes potential mechanistic roles of RANKL in these skeletal complications of malignancy. Myeloma and solid tumor bone metastases are incurable and result in significant patient morbidity. In early clinical trials evaluating solid tumor bone metastases and multiple myeloma, RANKL MAb reduces bone resorption and SREs more effectively than bisphosphonates. Recent results from a large phase III trial also show superiority of RANKL MAb to zoledronic acid. In vivo studies show powerful reduction of calcium with RANKL inhibition in models of hypercalcemia of malignancy. RANKL inhibition also arrests bone loss in the setting of estrogen depletion with aromatase inhibitors. Thus RANKL inhibition effectively osteoclast-dependent inhibits skeletal complications of malignancy. In vivo studies suggest that RANKL inhibition completely eradicates osteoclasts from the bone environment. in contrast to it possible bisphosphonates. ls that osteoclasts provide stimulus to cancer cells independent of bone resorption? Are osteoclasts a source of important tumor growth factors? In addition to the known osteoclast-dependent effects, bone-derived RANKL may enhance homing of RANKpositive cancer cells independent of osteoclast activity or bone resorption. Tumor-derived RANKL may also play a role in EMT, providing another target for RANKL inhibition. Despite concerns about potential interference with TRAIL and the immune



Fig. 1. Mechanistic roles of RANKL in skeletal complications of malignancy. Role of RANKL in solid tumor bone metastases and myeloma (top panel), hypercalcemia of malignancy (middle panel) and treatment-induced bone loss (bottom panel). Targeting RANKL effectively reduces bone resorption and associated complications in cancer and may also have direct anti-tumor effects.

system, RANKL inhibition does not appear to increase the incidence of neoplasm or interfere with lymphocytes. RANKL inhibitors are not stored in bone like bisphosphonates, and suppression of bone turnover is reversible. It is not apparent whether RANKL will associated with inhibitors be osteonecrosis of the jaw, but perhaps the reversibility will be advantageous and reduce this risk. There does appear to be increased risk of urinary tract infections with the use of RANKL MAb, and a higher frequency of hypertension that was comparable to alendronate. Longer and larger studies will need to be completed to further evaluate these potential concerns, and to determine whether RANKL inhibition has anti-tumor effects or impacts survival of patients with multiple myeloma or solid tumor bone metastases.

Conflict of Interest: Dr. Guise reports that she is on the advisory board, and consults, for Amgen and Novartis, and that she owns stock in Amgen. Dr. Crook reports no conflicts of interest.

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References

- Demers LM. Costa L, Chinchilli VM, Gaydos L, Curley E, Lipton A. Biochemical markers of bone turnover in patients with metastatic bone disease. *Clin Chem.* 1995 Oct;41(10):1489-94.
- Lipton A, Small E, Saad F, Gleason D, Gordon D, Smith M, Rosen L, Kowalski MO, Reitsma D, Seaman J. The new bisphosphonate, Zometa (zoledronic acid), decreases skeletal complications in both osteolytic and osteoblastic lesions: a comparison to pamidronate. *Cancer Invest*. 2002;20 Suppl 2:45-54.
- Saad F, Gleason DM, Murray R, Tchekmedyian S, Venner P, Lacombe L, Chin JL, Vinholes JJ, Goas JA, Chen B; Zoledronic Acid Prostate Cancer Study Group. A randomized, placebocontrolled trial of zoledronic acid in patients with hormone-refractory metastatic prostate carcinoma. J Natl Cancer Inst. 2002 Oct 2;94(19):1458-68.

- Brown JE, Cook RJ, Major P, Lipton A, Saad F, Smith M, Lee KA, Zheng M, Hei YJ, Coleman RE. Bone turnover markers as predictors of skeletal complications in prostate cancer, lung cancer, and other solid tumors. J Natl Cancer Inst. 2005 Jan 5;97(1):59-69.
- Cook RJ, Coleman R, Brown J, Lipton A, Major P, Hei YJ, Saad F, Smith MR. Markers of bone metabolism and survival in men with hormone-refractory metastatic prostate cancer. *Clin Cancer Res.* 2006 Jun 1;12(11 Pt 1):3361-7.
- Coleman RE. Bisphosphonates: clinical experience. *Oncologist*. 2004;9 Suppl 4:14-27.
- Fizazi K, Lipton A, Mariette X, Suarez T, Body J, Rahim Y, Gralow JR, Gao G, Wu L. Denosumab in patients with bone metastases from prostate, breast and other cancers and elevated urinary Ntelopeptide during intravenous bisphosphonate therapy: Final results of a randomized, phase II study. *J Clin Oncol.* 2008;26(May 20 Suppl):#3596.
- Fizazi K, Lipton A, Mariette X, Body JJ, Rahim Y, Gralow JR, Gao G, Wu L, Sohn W, Jun S. Randomized phase II trial of denosumab in patients with bone metastases from prostate cancer, breast cancer, or other neoplasms after intravenous bisphosphonates. *J Clin Oncol.* 2009 Apr 1;27(10):1564-71.
- 9. Simonet WS. Lacev DL. Dunstan CR. Kelley M, Chang MS, Luthy R, Nguyen HQ, Wooden S, Bennett L, Boone T, Shimamoto G, DeRose M, Elliott R. Colombero A, Tan HL, Trail G, Sullivan J, Davy E, Bucay N, Renshaw-Gegg L, Hughes TM, Hill D, Pattison W, Campbell P, Sander S, Van G, Tarpley J, Derby P, Lee R, Boyle WJ. Osteoprotegerin: a novel secreted protein involved in the regulation of bone density. Cell. 1997 Apr 18;89(2):309-19.
- Tsuda E, Goto M, Mochizuki S, Yano K, Kobayashi F, Morinaga T, Higashio K. Isolation of a novel cytokine from human

fibroblasts that specifically inhibits osteoclastogenesis. *Biochem Biophys Res Commun.* 1997 May 8;234(1):137-42.

- Lacey DL, Timms E, Tan HL, Kelley MJ, Dunstan CR, Burgess T, Elliott R, Colombero A, Elliott G, Scully S, Hsu H, Sullivan J, Hawkins N, Davy E, Capparelli C, Eli A, Qian YX, Kaufman S, Sarosi I, Shalhoub V, Senaldi G, Guo J, Delaney J, Boyle WJ. Osteoprotegerin ligand is a cytokine that regulates osteoclast differentiation and activation. *Cell*. 1998 Apr 17;93(2):165-76.
- 12. Kong YY, Yoshida H, Sarosi I, Tan HL, Timms E, Capparelli C, Morony S, Oliveira-dos-Santos AJ, Van G, Itie A, Khoo W, Wakeham A, Dunstan CR, Lacey DL, Mak TW, Boyle WJ, Penninger JM. OPGL is a key regulator osteoclastogenesis, lymphocyte of development and lymph-node organogenesis. Nature. 1999 Jan 28;397(6717):315-23.
- Dougall WC, Glaccum M, Charrier K, Rohrbach K, Brasel K, De Smedt T, Daro E, Smith J, Tometsko ME, Maliszewski CR, Armstrong A, Shen V, Bain S, Cosman D, Anderson D, Morrissey PJ, Peschon JJ, Schuh J. RANK is essential for osteoclast and lymph node development. *Genes Dev*. 1999 Sep 15;13(18):2412-24.
- 14. Thomas RJ, Guise TA, Yin JJ, Elliott J, Horwood NJ, Martin TJ, Gillespie MT. Breast cancer cells interact with osteoblasts to support osteoclast formation. *Endocrinology*. 1999 Oct;140(10):4451-8.
- Palmqvist P, Persson E, Conaway HH, Lerner UH. IL-6, leukemia inhibitory factor, and oncostatin M stimulate bone resorption and regulate the expression of receptor activator of NF-kappa B ligand, osteoprotegerin, and receptor activator of NF-kappa B in mouse calvariae. *J Immunol.* 2002 Sep 15;169(6):3353-62.

- Horwood NJ, Elliott J, Martin TJ, Gillespie MT. Osteotropic agents regulate the expression of osteoclast differentiation factor and osteoprotegerin in osteoblastic stromal cells. *Endocrinology*. 1998 Nov;139(11):4743-6.
- 17. Bendre MS, Montague DC, Peery T, Akel NS, Gaddy D, Suva LJ. Interleukin-8 stimulation of osteoclastogenesis and bone resorption is a mechanism for the increased osteolysis of metastatic bone disease. *Bone*. 2003 Jul;33(1):28-37.
- Tat SK, Pelletier JP, Lajeunesse D, Fahmi H, Duval N, Martel-Pelletier J. Differential modulation of RANKL isoforms by human osteoarthritic subchondral bone osteoblasts: influence of osteotropic factors. *Bone*. 2008 Aug;43(2):284-91.
- 19. Cross RF. SS, Harrison Balasubramanian SP. Lippitt JM. Evans CA, Reed MW, Holen I. Expression of receptor activator of nuclear factor kappabeta ligand (RANKL) and tumour factor related, apoptosis necrosis inducing ligand (TRAIL) in breast cancer. and their relations with osteoprotegerin, oestrogen receptor, and clinicopathological variables. J Clin Pathol. 2006 Jul;59(7):716-20.
- Park HR, Min SK, Cho HD, Kim DH, Shin HS, Park YE. Expression of osteoprotegerin and RANK ligand in breast cancer bone metastasis. J Korean Med Sci. 2003 Aug;18(4):541-6.
- Ohshiba T, Miyaura C, Inada M, Ito A. Role of RANKL-induced osteoclast formation and MMP-dependent matrix degradation in bone destruction by breast cancer metastasis. *Br J Cancer*. 2003 Apr 22;88(8):1318-26.
- 22. Goranova-Marinova V, Goranov S, Pavlov P, Tzvetkova T. Serum levels of OPG, RANKL and RANKL/OPG ratio in newly-diagnosed patients with multiple myeloma. Clinical correlations. *Haematologica*. 2007 Jul;92(7):1000-1.

- Seidel C, Hjertner O, Abildgaard N, Heickendorff L, Hjorth M, Westin J, Nielsen JL, Hjorth-Hansen H, Waage A, Sundan A, Borset M. Serum osteoprotegerin levels are reduced in patients with multiple myeloma with lytic bone disease. *Blood*. 2001 Oct 1;98(7):2269-71.
- 24. Terpos E, Szydlo R, Apperley JF, Hatjiharissi E, Politou M, Meletis J, Viniou N, Yataganas X, Goldman JM, Rahemtulla A. Soluble receptor activator of nuclear factor kappaB ligandosteoprotegerin ratio predicts survival in multiple myeloma: proposal for a novel prognostic index. *Blood.* 2003 Aug 1;102(3):1064-9.
- 25. Sezer O, Heider U, Jakob C, Zavrski I, Eucker J, Possinger K, Sers C, Krenn V. Immunocytochemistry reveals RANKL expression of myeloma cells. *Blood*. 2002 Jun 15;99(12):4646-7; author reply 4647.
- Sezer O, Heider U, Jakob C, Eucker J, Possinger K. Human bone marrow myeloma cells express RANKL. J Clin Oncol. 2002 Jan 1;20(1):353-4.
- 27. Farrugia AN, Atkins GJ, To LB, Pan B, Horvath N, Kostakis P, Findlay DM, Bardy P, Zannettino AC. Receptor activator of nuclear factor-kappaB ligand expression by human myeloma cells mediates osteoclast formation in vitro and correlates with bone destruction in vivo. *Cancer Res.* 2003 Sep 1;63(17):5438-45.
- Heider U, Zavrski I, Jakob C, Bangeroth K, Fleissner C, Langelotz C, Possinger K, Hofbauer LC, Viereck V, Sezer O. Expression of receptor activator of NF-kappaB ligand (RANKL) mRNA in human multiple myeloma cells. *J Cancer Res Clin Oncol.* 2004 Aug;130(8):469-74.
- Lai FP, Cole-Sinclair M, Cheng WJ, Quinn JM, Gillespie MT, Sentry JW, Schneider HG. Myeloma cells can directly contribute to the pool of RANKL in bone bypassing the classic stromal

and osteoblast pathway of osteoclast stimulation. *Br J Haematol*. 2004 Jul;126(2):192-201.

- Roux S, Meignin V, Quillard J, Meduri G, Guiochon-Mantel A, Fermand JP, Milgrom E, Mariette X. RANK (receptor activator of nuclear factor-kappaB) and RANKL expression in multiple myeloma. *Br J Haematol*. 2002 Apr;117(1):86-92.
- Pearse RN, Sordillo EM, Yaccoby S, Wong BR, Liau DF, Colman N, Michaeli J, Epstein J, Choi Y. Multiple myeloma disrupts the TRANCE/osteoprotegerin cytokine axis to trigger bone destruction and promote tumor progression. *Proc Natl Acad Sci U S A*. 2001 Sep 25;98(20):11581-6.
- Giuliani N, Colla S, Sala R, Moroni M, Lazzaretti M, La Monica S, Bonomini S, Hojden M, Sammarelli G, Barille S, Bataille R, Rizzoli V. Human myeloma cells stimulate the receptor activator of nuclear factor-kappa B ligand (RANKL) in T lymphocytes: a potential role in multiple myeloma bone disease. *Blood*. 2002 Dec 15;100(13):4615-21.
- Chen G, Sircar K, Aprikian A, Potti A, Goltzman D, Rabbani SA. Expression of RANKL/RANK/OPG in primary and metastatic human prostate cancer as markers of disease stage and functional regulation. *Cancer*. 2006 Jul 15;107(2):289-98.
- Brown JM, Corey E, Lee ZD, True LD, Yun TJ, Tondravi M, Vessella RL. Osteoprotegerin and rank ligand expression in prostate cancer. *Urology*. 2001 Apr;57(4):611-6.
- Perez-Martinez FC, Alonso V, Sarasa JL, Manzarbeitia F, Vela-Navarrete R, Calahorra FJ, Esbrit P. Receptor activator of nuclear factor-kappaB ligand (RANKL) as a novel prognostic marker in prostate carcinoma. *Histol Histopathol.* 2008 Jun;23(6):709-15.
- Jones DH, Nakashima T, Sanchez OH, Kozieradzki I, Komarova SV, Sarosi I, Morony S, Rubin E, Sarao R, Hojilla CV,

Komnenovic V, Kong YY, Schreiber M, Dixon SJ, Sims SM, Khokha R, Wada T, Penninger JM. Regulation of cancer cell migration and bone metastasis by RANKL. *Nature*. 2006 Mar 30;440(7084):692-6.

- 37. Odero-Marah VA, Wang R, Chu G, Zayzafoon M, Xu J, Shi C, Marshall FF, Zhau HE, Chung LW. Receptor activator of NF-kappaB Ligand (RANKL) expression is associated with epithelial to mesenchymal transition in human prostate cancer cells. *Cell Res.* 2008 Aug;18(8):858-70.
- Armstrong AP, Miller RE, Jones JC, Zhang J, Keller ET, Dougall WC. RANKL acts directly on RANKexpressing prostate tumor cells and mediates migration and expression of tumor metastasis genes. *Prostate*. 2008 Jan 1;68(1):92-104.
- Hwang C, Chung H, Kang Y, Moon I, Yim C, Han K, Jang H, Park W, Yoon H, Han I. Effects of RANKL on breast cancer cell lines. *J Bone Miner Res.* 2002 Sep;17(Suppl 1):S310.
- 40. Holen I, Cross SS, Neville-Webbe HL, Cross NA, Balasubramanian SP, Croucher PI, Evans CA, Lippitt JM, Coleman RE, Eaton CL. Osteoprotegerin (OPG) expression by breast cancer cells in vitro and breast tumours in vivo--a role in tumour cell survival? *Breast Cancer Res Treat.* 2005 Aug;92(3):207-15.
- 41. Fisher JL, Thomas-Mudge RJ, Elliott J, Hards DK, Sims NA, Slavin J, Martin TJ, Gillespie MT. Osteoprotegerin overexpression by breast cancer cells enhances orthotopic and osseous tumor growth and contrasts with that delivered therapeutically. *Cancer Res.* 2006 Apr 1;66(7):3620-8.
- 42. Kapoor P, Suva LJ, Welch DR, Donahue HJ. Osteoprotegerin and the bone homing and colonization potential of breast cancer cells. *J Cell Biochem*. 2008 Jan 1;103(1):30-41.

- Nicolin V, Narducci P. Soluble TRAIL could enhance bone destruction acting on Rank-ligand in estrogen-independent human breast cancer cell line MDA-MB-231. Acta Histochem. 2008 Dec 4. [Epub ahead of print]
- 44. Canon JR, Roudier M, Bryant R, Morony S, Stolina M, Kostenuik PJ, Dougall WC. Inhibition of RANKL blocks skeletal tumor progression and improves survival in a mouse model of breast cancer bone metastasis. *Clin Exp Metastasis*. 2008;25(2):119-29.
- 45. Zheng Y, Zhou H, Brennan K, Blair JM, Modzelewski JR, Seibel MJ, Dunstan CR. Inhibition of bone resorption, rather than direct cytotoxicity, mediates the anti-tumour actions of ibandronate and osteoprotegerin in a murine model of breast cancer bone metastasis. *Bone*. 2007 Feb;40(2):471-8.
- 46. Cross SS, Yang Z, Brown NJ, Balasubramanian SP, Evans CA, Woodward JK, Neville-Webbe HL, Lippitt JM, Reed MW, Coleman RE, Holen I. Osteoprotegerin (OPG)--a potential new role in the regulation of endothelial cell phenotype and tumour angiogenesis? *Int J Cancer*. 2006 Apr 15;118(8):1901-8.
- 47. Morony S, Warmington K, Tan H, Shalhoub V, Chow G, Dunstan CR, Lacey DL, Kostenuik PJ. OPG inhibits the progression of bone destruction and skeletal tumor burden in mice with established osteolytic MDA-231 breast cancer metastases. *J Bone Miner Res.* 2002 Sep;17(Suppl 1):S147.
- Morony S, Capparelli C, Sarosi I, Lacey DL, Dunstan CR, Kostenuik PJ. Osteoprotegerin inhibits osteolysis and decreases skeletal tumor burden in syngeneic and nude mouse models of experimental bone metastasis. *Cancer Res.* 2001 Jun 1;61(11):4432-6.
- 49. Tometsko M, Roudier M, Canon J, Bryant B, Miller R, Jones J, Armstrong A, Dougall W, Chaisson ML. RANKL inhibition causes a greater suppression

of tumor-induced osteoclastogenesis than zoledronate treatment in vivo and RANKL rescues osteoclasts from zoledronate killing in vitro. *J Bone Miner Res.* 2006 Sep;21(Suppl 1):S346.

- Vanderkerken K, De Leenheer E, Shipman C, Asosingh K, Willems A, Van Camp B, Croucher P. Recombinant osteoprotegerin decreases tumor burden and increases survival in a murine model of multiple myeloma. *Cancer Res.* 2003 Jan 15;63(2):287-9.
- 51. Sordillo EM, Pearse RN. RANK-Fc: a therapeutic antagonist for RANK-L in myeloma. *Cancer*. 2003 Feb 1;97(3 Suppl):802-12.
- 52. Croucher PI, Shipman CM, Van Camp B, Vanderkerken K. Bisphosphonates and osteoprotegerin as inhibitors of myeloma bone disease. *Cancer*. 2003 Feb 1;97(3 Suppl):818-24.
- 53. Yaccoby S, Pearse RN, Johnson CL, Barlogie B, Choi Y, Epstein J. Myeloma interacts with the bone marrow microenvironment to induce osteoclastogenesis and is dependent on osteoclast activity. *Br J Haematol*. 2002 Feb;116(2):278-90.
- 54. Zhang J, Dai J, Qi Y, Lin DL, Smith P, Strayhorn C, Mizokami A, Fu Z, Westman J, Keller ET. Osteoprotegerin inhibits prostate cancer-induced osteoclastogenesis and prevents prostate tumor growth in the bone. J *Clin Invest*. 2001 May;107(10):1235-44.
- 55. Whang PG, Schwarz EM, Gamradt SC, Dougall WC, Lieberman JR. The effects of RANK blockade and osteoclast depletion in a model of pure osteoblastic prostate cancer metastasis in bone. *J Orthop Res.* 2005 Nov;23(6):1475-83.
- 56. Quinn JE, Brown LG, Zhang J, Keller ET, Vessella RL, Corey E. Comparison of Fc-osteoprotegerin and zoledronic acid activities suggests that zoledronic acid inhibits prostate cancer in bone by indirect mechanisms. *Prostate Cancer Prostatic Dis.* 2005;8(3):253-9.

- Zhang J, Dai J, Yao Z, Lu Y, Dougall W, Keller ET. Soluble receptor activator of nuclear factor kappaB Fc diminishes prostate cancer progression in bone. *Cancer Res.* 2003 Nov 15;63(22):7883-90.
- 58. Kiefer JA, Vessella RL, Quinn JE, Odman AM, Zhang J, Keller ET, Kostenuik PJ, Dunstan CR, Corey E. The effect of osteoprotegerin administration on the intra-tibial growth of the osteoblastic LuCaP 23.1 prostate cancer xenograft. *Clin Exp Metastasis*. 2004;21(5):381-7.
- 59. Morrissey C, Kostenuik PL, Brown LG, Vessella RL, Corey E. Host-derived RANKL is responsible for osteolysis in a C4-2 human prostate cancer xenograft model of experimental bone metastases. *BMC Cancer*. 2007 Aug 3;7:148.
- Miller RE, Roudier M, Jones J, Armstrong A, Canon J, Dougall WC. RANK ligand inhibition plus docetaxel improves survival and reduces tumor burden in a murine model of prostate cancer bone metastasis. *Mol Cancer Ther.* 2008 Jul;7(7):2160-9.
- Ignatoski KM, Escara-Wilke JF, Dai JL, Lui A, Dougall W, Daignault S, Yao Z, Zhang J, Day ML, Sargent EE, Keller ET. RANKL inhibition is an effective adjuvant for docetaxel in a prostate cancer bone metastases model. *Prostate*. 2008 Jun 1;68(8):820-9.
- 62. Feeley BT, Liu NQ, Conduah AH, Krenek L, Roth K, Dougall WC, Huard J, Dubinett S, Lieberman JR. Mixed metastatic lung cancer lesions in bone are inhibited by noggin overexpression and Rank:Fc administration. *J Bone Miner Res*. 2006 Oct;21(10):1571-80.
- 63. Morony S, Warmington K, Adamu S, Asuncion F, Geng Z, Grisanti M, Tan HL, Capparelli C, Starnes C, Weimann B, Dunstan CR, Kostenuik PJ. The inhibition of RANKL causes greater suppression of bone resorption and hypercalcemia compared with

bisphosphonates in two models of humoral hypercalcemia of malignancy. *Endocrinology*. 2005 Aug;146(8):3235-43.

- 64. Oyajobi BO, Anderson DM, Traianedes K, Williams PJ, Yoneda T, Mundy GR. Therapeutic efficacy of a soluble receptor activator of nuclear factor kappaB-IgG Fc fusion protein in suppressing bone resorption and hypercalcemia in a model of humoral hypercalcemia of malignancy. *Cancer Res.* 2001 Mar 15;61(6):2572-8.
- Luger NM, Honore P, Sabino MA, Schwei MJ, Rogers SD, Mach DB, Clohisy DR, Mantyh PW. Osteoprotegerin diminishes advanced bone cancer pain. *Cancer Res.* 2001 May 15;61(10):4038-47.
- 66. Lipton A, Steger GG, Figueroa J, Alvarado C, Solal-Celigny P, Body JJ, de Boer R, Berardi R, Gascon P, Tonkin KS, Coleman R, Paterson AH, Peterson MC, Fan M, Kinsey A, Jun S. Randomized active-controlled phase II study of denosumab efficacy and safety in patients with breast cancer-related bone metastases. J Clin Oncol. 2007 Oct 1;25(28):4431-7.
- 67. Lipton A, Steger GG, Figueroa J, Alvarado C, Solal-Celigny P, Body JJ, de Boer R, Berardi R, Gascon P, Tonkin KS, Coleman RE, Paterson AH, Gao GM, Kinsey AC, Peterson MC, Jun S. Extended efficacy and safety of denosumab in breast cancer patients with bone metastases not receiving prior bisphosphonate therapy. *Clin Cancer Res.* 2008 Oct 15;14(20):6690-6.
- Vij R, Horvath N, Spencer A, Taylor K, Vadhan-Raj S, Smith J, Jun S. An openlabel, phase II trial of denosumab in the treatment of relapsed of plateau-phase multiple myeloma. *Blood*. 2007; 110:#3604.
- 69. Thomas D, Chawla SP, Skubitz K, Staddon AP, Henshaw R, Blay JY, Smith J, Ye Z, Roudier M, Jun S. Denosumab treatment of giant cell 337

tumor of bone: Interim analysis of an open-label phase II study. *J Clin Oncol.* 2008;26(May 20 Suppl):#10500.

- Lewiecki EM, Miller PD, McClung MR, Cohen SB, Bolognese MA, Liu Y, Wang A, Siddhanti S, Fitzpatrick LA. Two-year treatment with denosumab (AMG 162) in a randomized phase 2 study of postmenopausal women with low BMD. *J Bone Miner Res.* 2007 Dec;22(12):1832-41.
- 71. Samadfam R, Xia Q, Goltzman D. Cotreatment of PTH with osteoprotegerin or alendronate increases its anabolic effect on the skeleton of oophorectomized mice. *J Bone Miner Res.* 2007 Jan;22(1):55-63.
- 72. Li X, Ominsky MS, Asuncion F, Grisanti M, Tan HL, Warminton KS, Kostenuik PJ. RANKL inhibition by OPG pevents bone loss and preserves trabecular architecture in orchiectomized rats. *Calcif Tissue Int.* 2006;78:S144-S145.
- 73. Chanda D, Isayeva T, Kumar S, Siegal GP, Szafran AA, Zinn KR, Reddy VV, Ponnazhagan S. Systemic osteoprotegerin gene therapy restores tumor-induced bone loss in а therapeutic model of breast cancer bone metastasis. Mol Ther. 2008 May;16(5):871-8.
- 74. Ellis GK, Bone HG, Chlebowski R, Paul D, Spadafora S, Smith J, Fan M, Jun S. Randomized trial of denosumab in patients receiving adjuvant aromatase

inhibitors for nonmetastatic breast cancer. *J Clin Oncol.* 2008 Oct 20;26(30):4875-82.

- Baud'huin M, Lamoureux F, Duplomb L, Redini F, Heymann D. RANKL, RANK, osteoprotegerin: key partners of osteoimmunology and vascular diseases. *Cell Mol Life Sci.* 2007 Sep;64(18):2334-50.
- 76. Miller RE, Branstetter D, Armstrong A, Kennedy B, Jones J, Cowan L, Bussiere J, Dougall WC. Receptor activator of NF-kappa B ligand inhibition suppresses bone resorption and hypercalcemia but does not affect host immune responses to influenza infection. *J Immunol*. 2007 Jul 1;179(1):266-74.
- 77. Bekker PJ, Holloway DL, Rasmussen AS, Murphy R, Martin SW, Leese PT, Holmes GB, Dunstan CR, DePaoli AM. A single-dose placebo-controlled study of AMG 162, a fully human monoclonal antibody to RANKL, in postmenopausal women. *J Bone Miner Res.* 2004 Jul;19(7):1059-66.
- 78. Body JJ, Facon T, Coleman RE, Lipton A, Geurs F, Fan M, Holloway D, Peterson MC, Bekker PJ. A study of the biological receptor activator of nuclear factor-kappaB ligand inhibitor, denosumab, in patients with multiple myeloma or bone metastases from breast cancer. *Clin Cancer Res.* 2006 Feb 15;12(4):1221-8.